



Global and countrywide prevalence of subclinical and clinical mastitis in dairy cattle and buffaloes by systematic review and meta-analysis

Paramanandham Krishnamoorthy^{*}, Akshata Lokanath Goudar, Kuralayanapalya Puttahonnappa Suresh, Parimal Roy

Pathoepidemiology Laboratory, ICAR- National Institute of Veterinary Epidemiology and Disease Informatics (NIVEDI), Post Box No.6450, Ramagondanahalli, Yelahanka, Bengaluru 560064, India

ARTICLE INFO

Keywords:
Subclinical mastitis
Clinical mastitis
Prevalence
World
India
Meta-analysis

ABSTRACT

In the present study, subclinical mastitis (SCM) and clinical mastitis (CM) prevalence for various countries in the World were calculated by using online and offline databases. The SCM and CM prevalence studies reported during 1967-2019 were collected, reviewed, and a meta-analysis was done in R-Software. A total of 222 and 150 studies from the World and 103 and 37 studies from India on SCM and CM, respectively were included. The pooled prevalence of SCM and CM were 42% [Confidence Interval (CI) 38-45%, Prediction Interval (PI) 10-83%] and 15% [CI 12-19%, PI 1-81%] in the World respectively, 45% [CI 40-49%, PI 11-84%] and 18% [CI 14-23%, PI 3-60%] in India respectively. Continent-wise analysis indicated a higher prevalence of SCM in North America and CM in Europe and among the countries, a higher SCM prevalence in Uganda and CM in the United Kingdom was observed. Further, species-wise indicated a higher SCM and CM prevalence in buffaloes of the World than the cattle. Based on method-wise, SCM and CM prevalence were high in somatic cell count and clinical examination, respectively in the World. The SCM prevalence was higher than CM and indicated the importance of SCM in dairy cattle. This might result in low milk productivity in dairying and may set off losses to dairy farmers. Hence, there is an urgent need to reduce the SCM and CM prevalence by implementing scientific dairy management, good feeding practices, and timely therapeutic interventions for increasing the benefits from dairying to the farmers in the World.

1. Introduction

Mastitis causes economic losses to the dairy farmers in the World and in India, which mainly affects milk production. This production disease is considered an important disease of dairy cows which results in a reduction of milk production, loss in milk quality and quantity, losses due to discarded milk, premature culling, treatment costs, and extra labour cost (Hogeveen et al., 2011). Mastitis is a worldwide production disease of dairy cows (FAO, 2014). Mastitis can be classified into three types, namely sub-clinical, clinical and chronic mastitis, and it depends on causative organisms, breed, age, immunity, and stage of lactation of the animal. Sub-clinical mastitis (SCM) causes major loss to milk production due to the absence of any visible changes in milk and difficulty in detection. Clinical mastitis (CM), characterized by swelling of the udder, milk containing flakes, clots, and watery milk are observed grossly. Chronic mastitis is a rare form but leads to persistent

inflammation of the mammary gland in dairy animals. The estimated annual economic losses due to mastitis were reported to be US\$ 200 per cow per year (Costello, 2004). Viguier et al. (2009) reported an estimated total loss of US\$ 2 billion by the year 2009 in the United States of America (USA) was accounted for due to mastitis. The estimated annual economic loss due to both subclinical and clinical mastitis in India was US\$ 98,228 million [7165.51 crore Indian Rupees] as reported earlier (Bansal and Gupta, 2009).

The meta-analysis, an innovative tool for estimating the prevalence of various livestock diseases from different countries, and many studies have been reported in the field of Veterinary Science in recent times (Krishnamoorthy et al., 2020). Haidich (2010) indicated that meta-analysis is one of the formal, quantitative, epidemiological study design, which was used to systematically evaluate the previously reported studies to develop meaningful assumptions in the same area of research. The main objective of conducting a meta-analysis is to

* Corresponding author at: ICAR- National Institute of Veterinary Epidemiology and Disease Informatics (NIVEDI), Government of India, Post Box No.6450, Ramagondanahalli, Yelahanka, Bengaluru 560064, Karnataka, India.

E-mail address: P.Krishnamoorthy@icar.gov.in (P. Krishnamoorthy).



summarize and integrate results from several previous studies, analyze the differences reported in the prevalence in different studies, to overpower the small sample sizes used in the individual studies. It also helps in analysing the endpoints which require larger sample sizes, increasing the precision in effect estimates, to determine whether the new studies are needed for further and to generate new hypotheses for the studies in the future (Krishnamoorthy et al., 2019a, 2019b). The precarious steps to be considered in a meta-analysis study are the selection of studies, heterogeneity of prevalence reported, availability of evidence, and method of data analysis employed (Walker et al., 2008). There are few studies conducted in the past, by using meta-analysis method for estimating the prevalence of subclinical and clinical mastitis, major mastitis pathogens in India (Krishnamoorthy et al., 2017), anaplasmosis in dairy animals in India and the World (Krishnamoorthy et al., 2019a), and livestock-associated methicillin-resistant *Staphylococcus aureus* in animals in India (Krishnamoorthy et al., 2019b).

The cattle and buffalo population in the World were 1,489 and 206 million, respectively as available online (FAOSTAT, 2020). The annual combined cattle and buffalo milk production was 187.75 million tonnes during the year 2019 in India, which has increased by 6.5% when compared to 2018, and 394 grams per day was the per capita availability of milk as reported (BAHS, 2019). India also ranks first in the buffalo population and second in cattle population after Brazil (193.5 million) in the World (FAOSTAT, 2020). In India, the cattle and buffalo population were 192.5 and 109.8 million out of the total livestock population of 535.8 million, as per the 20th Livestock census (BAHS, 2019). Since the cattle and buffalo population was more in number in India, it was planned to calculate the prevalence estimates of subclinical and clinical mastitis separately and to study in detail as an example to other countries in the World. There are numerous studies on SCM and CM prevalence, reported from various geographical locations in the World. However, the prevalence reported is variable from study to study due to several factors including the period of study, location of the study, animal breed, lactation stage, sample numbers tested, sampling methodology employed and dairy managemental practices followed. Two studies, separately on SCM and CM prevalence in dairy cows in India for the period 1995–2014 by using meta-analysis were reported (Bangar et al., 2015, 2016). Another study from India reported the prevalence estimates for SCM and CM along with subgroup analysis by using studies reported during 1995–2015 based on a meta-analysis (Krishnamoorthy et al., 2017). However, there were no studies on SCM and CM prevalence estimates for a particular country or continents in the World as a whole, based on a meta-analysis, other than India. It is pertinent to collect the information on the prevalence of SCM and CM across different countries in the World, which will act as baseline information for planning mastitis prevention and control strategies in different countries of the World and India. Further, the studies on SCM and CM prevalence will be available in one place, which enables easy retrieval of mastitis studies in near future, by many researchers. Keeping this in mind, the present study was undertaken to do a meta-analysis of available literature on the prevalence of subclinical and clinical mastitis in the World to obtain the mastitis prevalence estimates among dairy cows from the studies reported during the period 1967–2019.

2. Materials and methods

2.1. Search strategy

A systematic literature search was performed by using keywords such as “mastitis prevalence, mastitis AND prevalence, subclinical mastitis, subclinical mastitis AND prevalence, clinical mastitis, clinical mastitis AND prevalence, bovine mastitis, bovine mastitis AND prevalence, prevalence” and followed as per the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) guidelines (Moher et al., 2015) for identifying the studies on the prevalence of subclinical and clinical mastitis in dairy cows in the World. The databases PubMed,

Elsevier, Springer, Indian [journals.com](#), Consortium of e-Resources in Agriculture (CeRA) under the Indian Council of Agricultural Research, and Google Scholar were searched for the publications on SCM and CM prevalence. More than 421 and 215 articles from the World and India were searched and identified from the database search. The studies from different journal publications were from 1967 to 2019 and 1995 to 2019 for the World and India, respectively on SCM and CM prevalence. The studies selected based on the characteristics of the publications which clearly state the following details including author names, year, country, state, number of samples tested, number of positive samples, diagnostic methods employed, and species either cattle/buffalo. Finally, the original articles were reviewed in detail and the references cited from the retrieved articles were searched again for some more relevant studies. The retrieval dates for the studies were limited to the period from January 1967 to December 2019 and the retrieval language was limited to English only.

2.2. Data processing and filtration

The SCM and CM prevalence studies were reviewed in full and included for meta-analysis based on the set inclusion criteria such as cross-sectional and observational studies (prevalence studies) and published in peer-reviewed journals either online or offline. The exclusion criteria for the studies included were case reports, review articles, and consideration of sample size. The study details were extracted from the characteristics reported in each study onto Microsoft Excel data sheets with a designed format. These characteristics include the year of publication, author names, study eligibility criteria, study period, species of animals tested, numbers of samples positive for SCM or CM, the total number of samples tested, prevalence of SCM, and CM, the diagnostic approaches utilized. The methods engaged were California mastitis test (CMT), surf field mastitis test (SFMT), white side test (WST), somatic cell count (SCC) in various studies for SCM, and bacterial culture examination and clinical examination (CE) for CM. The pooled prevalence estimates for SCM and CM were calculated by considering the highest value of prevalence obtained by different methods employed. There are three steps in data filtering, the first step is to ensure that the selected studies are coherent with the purpose of the meta-analysis. The second consisted of a critical and detailed review of each study selected for meta-analysis and authentication of data entries. In the third step, to ensure that a selected publication should not be an outlier concerning the characteristics and studies under consideration for meta-analysis. The above steps mentioned were similar to the PRISMA guidelines as mentioned earlier.

2.3. Meta-analysis methods

In usual terms, a meta-analysis uses specialized techniques for data collection and analysis developed exclusively for new research synthesis (Koricheva et al., 2013). The whole effect may be computed from a group of combined and representative studies or criteria that provides an impartial estimate of the effect, with an increase in the accuracy of this estimate obtained by employing meta-analysis. It provides the total effect estimate from a meta-analysis, normally at the bottom, and often using a diamond to distinguish it from the individual studies (Higgins et al., 2013). The R Open source scripting software version 3.2.5 (Comprehensive R Archive Network) was employed for meta-analysis by using the R package "Meta" (Schwarzer, 2007). The meta-analysis results obtained were represented graphically by forest plots. The forest plots, otherwise known as confidence interval plots, display prevalence estimates and their confidence intervals for each study. The generalized linear mixed model and transformation for proportion used was Logit transformation, i.e., “sm=PLOGIT” for analysis purposes. The prevalence estimate was represented as square and the horizontal line extending on either side of the square indicates the 95% confidence interval (CI). The prediction interval (PI) at a 95% level was given below

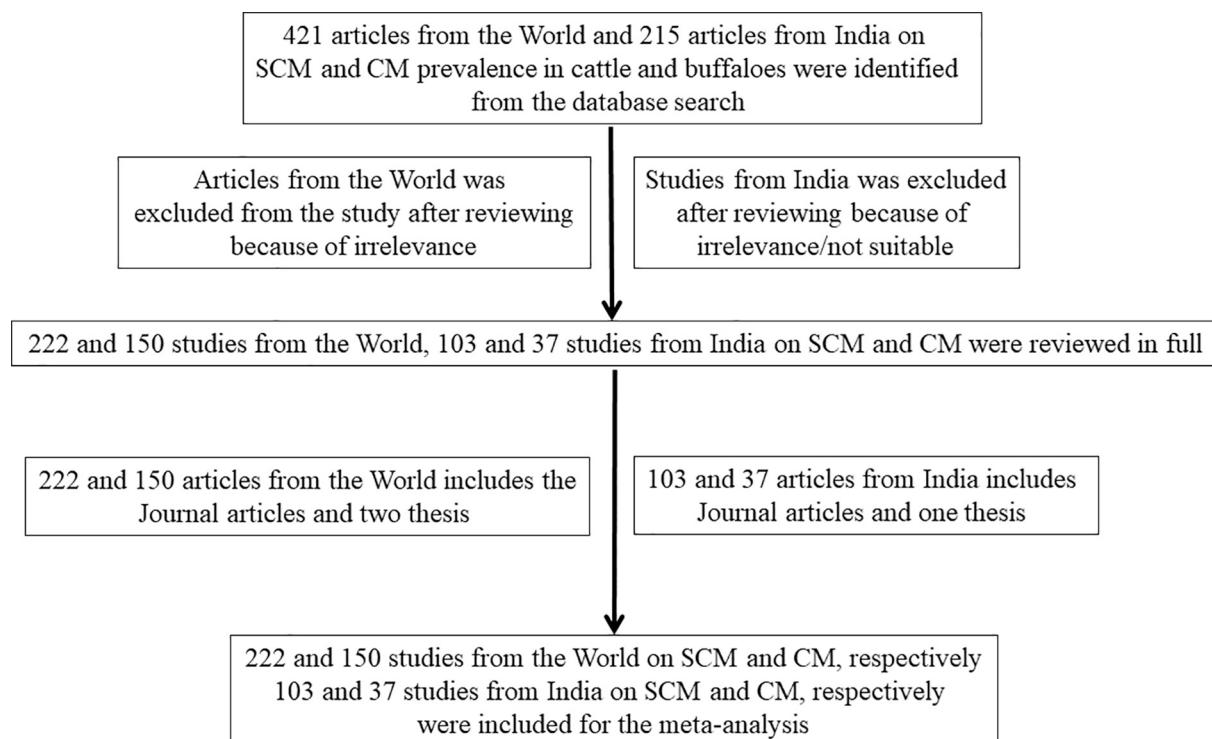


Fig. 1. Diagram showing the Subclinical mastitis (SCM) and Clinical mastitis (CM) prevalence studies from the World and India.

the forest plot as a shaded thick line. The heterogeneity between the studies included for the meta-analysis was determined by the I-Square value, Tau square value, and P values obtained in the forest plot. To reduce the heterogeneity between the selected studies on SCM and CM prevalence in the World and India, the subgroup analysis was done, based on various parameters as described earlier (Krishnamoorthy et al., 2019a) with some modifications. The funnel plot was also obtained for the meta-analysis, which was used to determine the outliers in the SCM and CM prevalence studies. The Cochran Q statistics were calculated as per the method reported earlier (Krishnamoorthy et al., 2017, 2019a, 2019b) and indicated the level of significance. If the P-value obtained was significant, the random effect model values in the forest plot were used for determining the prevalence estimates and CI. The forest plots were prepared for the overall SCM and CM prevalence estimates, continent-wise, period-wise, species-wise, method-wise, country-wise for the World. India was divided into different states and union territories based on the geographically and politically organized under one administrative Government, and further the states were grouped into five zones namely North, East, West, South, and Central zones. The overall SCM and CM prevalence estimates, period-wise, zone-wise, species-wise, method-wise, and state-wise were obtained for India. The prevalence estimates for SCM and CM were expressed as percentages and along with CI and PI at the 95 % level.

3. Results

3.1. Details of SCM and CM prevalence studies

In the current study, the systematic review and meta-analysis of SCM and CM prevalence studies in cattle and buffaloes from the World and India were carried out. The filled Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) 2009 checklist for the present study are given in Supplementary Table 1. The diagram revealing the details of the number of SCM and CM prevalence studies

retrieved, reviewed in full, and considered for a meta-analysis are given in Fig. 1. The details of the studies including continents of the World, countries, author, year, and either SCM or CM prevalence reported are given in Table 1. The number of studies on SCM and CM prevalence from the World included for meta-analysis were 222 and 150, respectively, and have been reported from all the continents except Antarctica. The number of countries included for determining SCM and CM prevalence estimates was 50 and 44, respectively with overall 61 countries reported mastitis prevalence studies. The number of studies reported was more in cattle for both SCM [206] and CM [143] when compared to buffaloes. Method-wise comparison indicated more number of studies used the California mastitis test (CMT) for SCM and clinical examination (CE) for CM diagnosis when compared with other methods. The year-wise and country-wise number of studies reported from the World are given in Fig. 2. The SCM and CM prevalence studies reported during the period 1967–2019 and 1982–2019, respectively from the World were included for meta-analysis. More number of studies reported in the World for SCM [34] and CM [16] was during the year 2013. Ethiopia reported the highest number of studies on SCM and CM prevalence and was 63 and 55, respectively followed by Bangladesh [22] and Pakistan [17] for SCM and Egypt [10] and the United State of America [10] for CM. The SCM and CM studies from India were analyzed separately as mentioned earlier for better comparison with other countries and not included for analysis under Asia [69 SCM and 25 CM studies]. The particulars of the studies included from the different states, zones, and either SCM or CM prevalence reported from India are given in Table 2. The total number of studies used for meta-analysis was 103 and 37 on SCM and CM prevalence, respectively in India. The SCM and CM prevalence studies in India were obtained from 21 states along with one union territory and 15 states, respectively were explored. In cattle, the number of studies on SCM [81] and CM [32] was high when paralleled with buffaloes. Most of the studies employed CMT for SCM and CE for CM diagnosis in India. The details of the year-wise and state-wise number of studies included from India are shown in Fig. 3. More number of studies on SCM [14] and

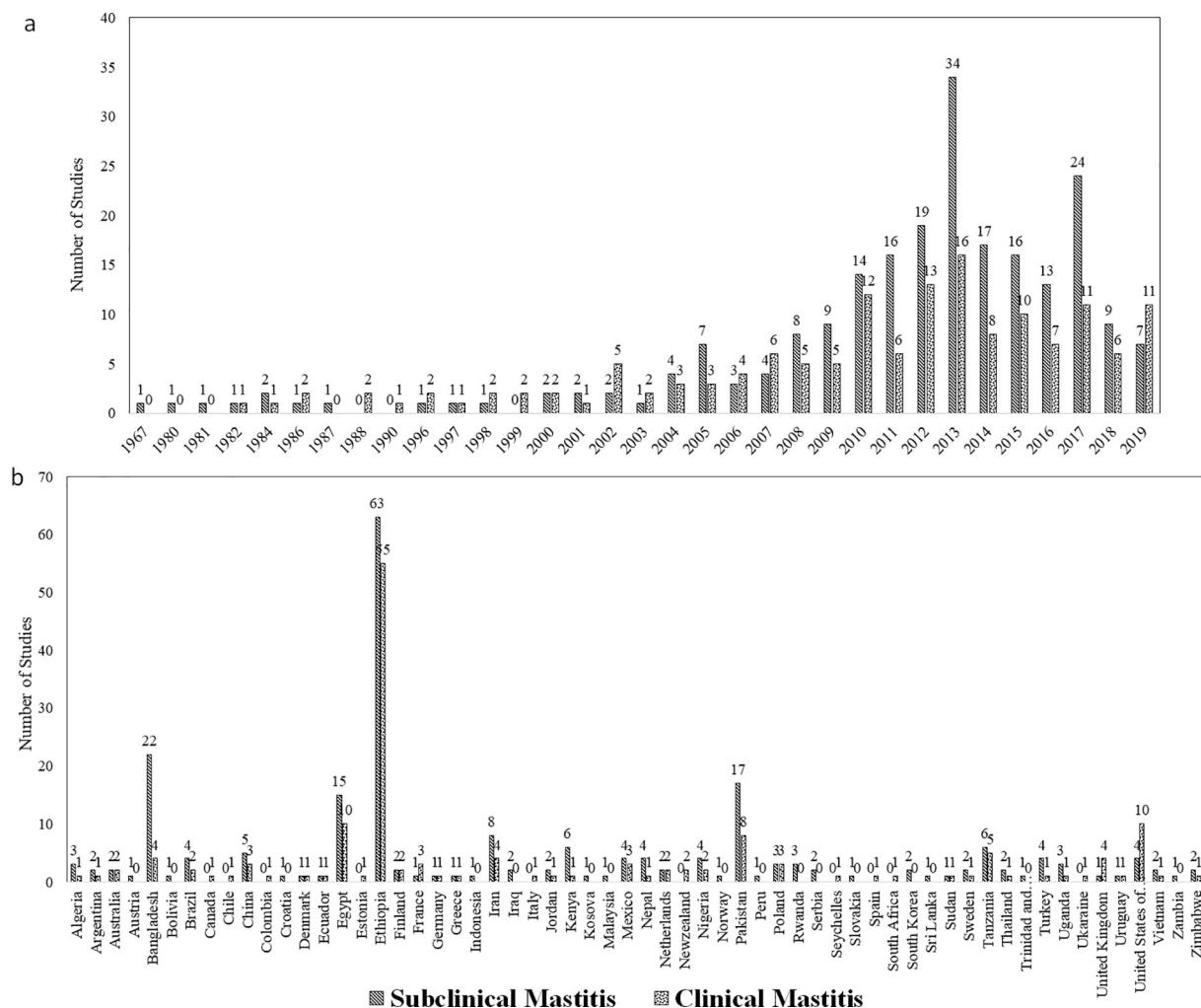


Fig. 2. Year-wise (a) and Country-wise (b) number of studies from the World on Subclinical and Clinical mastitis prevalence included for meta-analysis.

CM [5] were reported during the years 2013 and 2013 & 2017, respectively. To understand the heterogeneity between the various studies, the forest plot was employed for better visualization of meta-analysis results. Further, the subgroup analysis under different categories, the SCM and CM prevalence studies were segregated to obtain the prevalence estimates for superior understanding for both in the World and India.

3.2. Global SCM and CM prevalence

The details of the SCM prevalence estimates obtained by a meta-analysis from the World and the tests for heterogeneity are presented in Table 3. The pooled prevalence estimates for SCM were 42% [95% CI 38–45, PI 10–83%] with an aggregate of 2,51,372 samples tested from six continents of the World. The number of SCM prevalence studies incorporated was 13, 54, and 155 reported during the period 1967–2000, 2000–2010, and 2011–2019, respectively. The highest SCM prevalence [43%] was reported during the period 2011–2019 than other periods and endorses the increased prevalence in recent years. The continent-wise SCM prevalence estimates are depicted in Fig. 4. Based on

continent-wise analysis indicated high SCM prevalence in North America [46%] followed by Africa [44%], Asia [42%], Europe [37%], Oceania [36%], and Latin America [34%]. Period-wise analysis i.e., up to 2000 and after 2000 of the six continents revealed increasing SCM prevalence in recent years except for Africa and Latin America, which exhibited a decreasing trend. The species-wise evaluation indicated the highest SCM prevalence in buffaloes [46%] in comparison to cattle [42%]. The SCM prevalence was highest in SCC [46%] when compared to other methods namely CMT [43%], SFMT [41%], and WST [37%]. The meta-analysis results of CM prevalence estimates obtained for the World are given in Table 4. The CM pooled prevalence estimate was 15% [95% CI 12–19, PI 1–81] with a total of 1,10,066 samples from dairy cattle and buffaloes in the World. The year-wise prevalence estimates revealed a decreasing trend over the years and lowest during 2011–19 [12%] in recent times. High CM prevalence was reported from Europe [29%] followed by other continents in the World and the lowest in Oceania [5%] was observed. Based on the period-wise analysis indicated the decreasing trend in recent years except in Africa which indicated increasing CM prevalence. The buffaloes [28%] showed a high CM prevalence than cattle [14%], based on the studies analysed from the

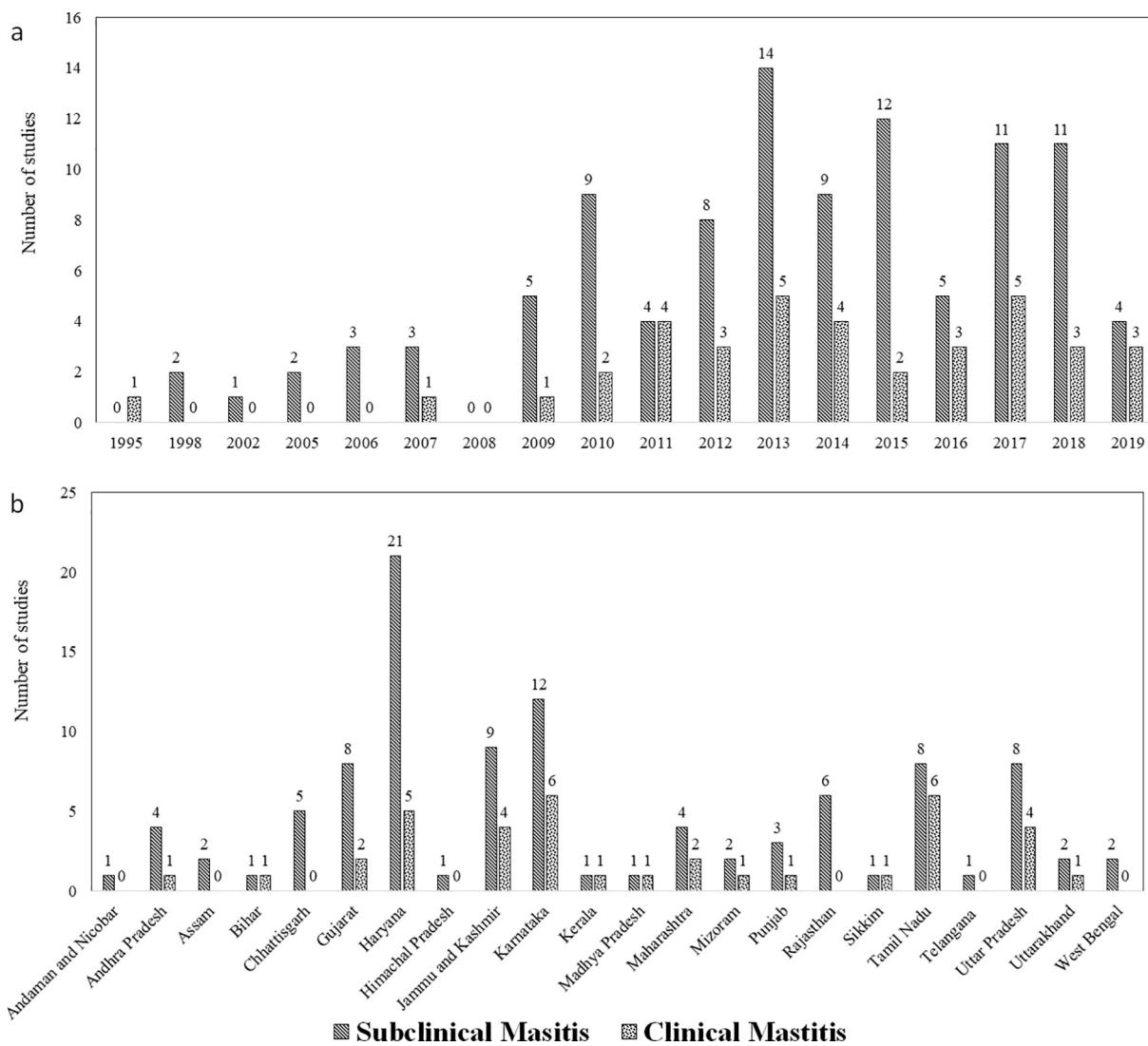


Fig. 3. Year-wise (a) and State-wise (b) number of studies from India on Subclinical and Clinical mastitis prevalence included for meta-analysis.

World. Among the studies reported, the most common diagnostic method used was CE of the udder and milk samples for CM, with a prevalence estimate of 15% [95% CI 12-19, PI 1-80%]. The Cochran Q values revealed a highly significant ($P<0.01$) difference between the studies included for SCM and CM prevalence in the World sub-grouped based on year-wise, continent-wise, period-wise, species-wise, and method-wise showing the heterogeneity between the studies selected for meta-analysis.

3.3. Countrywide SCM and CM prevalence

The details of SCM and CM prevalence values for various countries of the World attained by meta-analysis are presented in Tables 5 and 6, respectively. The number of countries included for meta-analysis was 50 and 44 for SCM and CM prevalence, respectively. The SCM prevalence was highest in Uganda [85%] followed by Indonesia [82%], Malaysia [82%], and other countries, the low prevalence was observed in

Uruguay [1%]. More number of the studies encompassed from Ethiopia for both SCM and CM, when compared to other countries. The highest CM prevalence was in the United Kingdom [74%], followed by Finland [45%], the USA [35%], and other forty-one countries. The lowest CM prevalence was observed in Thailand [1%] and Ukraine [1%]. The countrywide SCM prevalence studies showed a highly significant ($P<0.01$) difference for the various countries except for Argentina, which showed a significant ($P<0.05$) difference and no significant difference was observed for Finland, Netherlands, South Korea, Thailand, and Uganda. The Cochran Q values of CM prevalence studies from various countries revealed a highly significant ($P<0.01$) difference excluding Finland and Nigeria, which showed no significant difference between the studies included for a meta-analysis.

3.4. SCM and CM prevalence in India

The particulars of SCM and CM prevalence estimates in India based

Table 1

Details of Subclinical mastitis (SCM) and Clinical mastitis (CM) studies from the World included for meta-analysis.

No.	Continents	Countries	Studies [Author and Year]	SCM-1, CM-2
Africa	Algeria	Afri et al., 2017	1	Tesfaye and Abera, 2018 Tesfaye et al., 2012 Tesfaye, 2017 Tilahun and Aylate, 2015 Tolosa et al., 2009 Tolosa et al., 2013 Tuke et al., 2017 Workineh et al., 2002 Wubishet et al., 2013 Yohannes and Alemu, 2018
		Akkou et al., 2018	1, 2	1, 2
	Egypt	Saidi et al., 2013	1	1, 2
		Abdel-Rady and Sayed, 2009	1	1, 2
		Abd-Elrahman, 2013	1, 2	1, 2
		Ahmed et al., 2018	1	1, 2
		Amin et al., 2011	1, 2	1, 2
		El-Ashker et al., 2015	1, 2	1, 2
		El-Jakee et al., 2013	1, 2	1, 2
Ethiopia	Ethiopia	Elbably et al., 2013	1, 2	Zeryehun and Abera, 2017
		Elhaig and Selim, 2014	1	Zeryehun et al., 2013
		Elsayed et al., 2015	1, 2	Maichomo et al., 2019
		Hamed and Ziatoun, 2014	1	Mureithi and Njuguna, 2016
		Kotb et al., 2014	1, 2	Mureithi et al., 2017
		Lamey et al., 2013	1, 2	Ndirangu et al., 2019
		Osman et al., 2010	1, 2	Ndirangu et al., 2017
		Sayed et al., 2014	1, 2	Ondiek et al., 2013
		Zaki et al., 2010	1	Junaidu et al., 2011
		Abebe et al., 2016	1, 2	Shittu et al., 2012
Kenya	Kenya	Abera et al., 2012a	1, 2	Suleiman et al., 2012
		Abera et al., 2012b	1, 2	Umaru et al., 2017
		Abera et al., 2013	1, 2	Iraguha et al., 2015
		Abraham and Zeleke, 2017	2	Mpatswenumugabo et al., 2017
		Abunna et al., 2013	1, 2	Ndahetuye et al., 2019
		Adane et al., 2012	1, 2	Watson et al., 1996
		Alemu et al., 2013	1, 2	Brunner et al., 2019
		Almaw et al., 2008	1	Hamid et al., 2012
		Almaw et al., 2009	1	Kivaria et al., 2004
		Amin et al., 2017	1, 2	Mdegeila et al., 2005
Nigeria	Nigeria	Ayano et al., 2013	1	Mdegeila et al., 2009
		Bedacha and Menghistu, 2011	1, 2	Shem et al., 2001
		Belayneh et al., 2013	1, 2	Suleiman et al., 2013
		Beyene and Tolosa, 2017	1, 2	Suleiman et al., 2017
		Biffa et al., 2005	1, 2	Abrahmsén et al., 2014
		Birhanu et al., 2017	1	Björk, 2013
		Bitew et al., 2010	1, 2	Kasozi et al., 2014
		Dego and Tareke, 2003	1, 2	Ssajjakambwe et al., 2017
		Delelesse, 2010	1, 2	Eriksson, 2013
		Demissie et al., 2018	1, 2	Zimbabwé
Rwanda	Rwanda	Duguma et al., 2014	1, 2	Katsande et al., 2013
		Elemo et al., 2017	1, 2	Perry et al., 1987
		Getahun et al., 2008	1, 2	Bangladesh
		Girma et al., 2012	1	Badiuzzaman et al., 2015
		Haftu et al., 2012	1, 2	Bari et al., 2014
		Hailemeskel et al., 2014	1, 2	Barua et al., 2014
		Kebebew and Jorga, 2016	1, 2	Biswas and Sarker, 2017
		Kedir et al., 2016	1, 2	Hoque et al., 2014
		Lakew et al., 2009	1, 2	Hoque et al., 2018
		Lakew et al., 2019	1, 2	Hossain et al., 2016
Seychelles	Seychelles	Madalcho, 2019	1, 2	Islam et al., 2010
		Marama et al., 2016	1, 2	Islam et al., 2011
		Megersa et al., 2012	1, 2	Islam et al., 2014
		Mekibib et al., 2010	1, 2	Kabir et al., 2017
		Mekonnen and Tesfaye, 2010	1, 2	Kayesh et al., 2014
		Mekonnen et al., 2017	1	Meher et al., 2018
		Michael et al., 2013	1, 2	Quaderi et al., 2013
		Moges et al., 2012	1, 2	Rabbani and Samad, 2010
		Mulate et al., 2017	1	Rahman et al., 2009
		Mulshet et al., 2017	1	Rahman et al., 2010
Sudan	Sudan	Pal et al., 2017	1	Rahman et al., 2014
		Regassa et al., 2013	1, 2	Sarker et al., 2013
		Sarba and Tola, 2017	1, 2	Siddique et al., 2015
		Seid et al., 2015	1, 2	Siddique et al., 2013
		Shiferaw and Telila, 2016	1, 2	Sumon et al., 2017
		Sori et al., 2005	1, 2	Tripura et al., 2014
		Tadesse and Chanie, 2012	1, 2	Gao et al., 2017
		Tafa et al., 2015	1, 2	Li et al., 2009
		Tekle and Berihe, 2015	1, 2	Memon et al., 2013
				Wang et al., 2019
Tanzania	Tanzania			Xu et al., 2015
Asia	Asia			
Zambia	Zambia			
Zimbabwe	Zimbabwe			
China	China			

Table 1 (continued)

No.	Continents	Countries	Studies [Author and Year]	SCM-1, CM-2
			Tesfaye and Abera, 2018	1
			Tesfaye et al., 2012	1, 2
			Tesfaye, 2017	1, 2
			Tilahun and Aylate, 2015	1, 2
			Tolosa et al., 2009	1, 2
			Tolosa et al., 2013	1
			Tuke et al., 2017	1, 2
			Workineh et al., 2002	1, 2
			Wubishet et al., 2013	1, 2
			Yohannes and Alemu, 2018	1, 2
			Yohannis and Molla, 2013	1, 2
			Zenebe et al., 2014	1, 2
			Zeryehun and Abera, 2017	1, 2
			Zeryehun et al., 2013	1, 2
			Maichomo et al., 2019	1
			Mureithi and Njuguna, 2016	1
			Mureithi et al., 2017	1
			Ndirangu et al., 2019	1
			Ndirangu et al., 2017	1, 2
			Ondiek et al., 2013	1
			Junaidu et al., 2011	1
			Shittu et al., 2012	1
			Suleiman et al., 2012	1
			Umaru et al., 2017	1, 2
			Iraguha et al., 2015	1
			Mpatswenumugabo et al., 2017	1
			Ndahetuye et al., 2019	1
			Watson et al., 1996	2
			Brunner et al., 2019	2
			Hamid et al., 2012	1, 2
			Kivaria et al., 2004	1, 2
			Mdegeila et al., 2005	1
			Mdegeila et al., 2009	1, 2
			Shem et al., 2001	1, 2
			Suleiman et al., 2013	1, 2
			Suleiman et al., 2017	1
			Abrahmsén et al., 2014	1
			Björk, 2013	2
			Kasozi et al., 2014	1
			Ssajjakambwe et al., 2017	1
			Eriksson, 2013	1
			Katsande et al., 2013	1, 2
			Perry et al., 1987	1
			Badiuzzaman et al., 2015	1
			Bari et al., 2014	2
			Barua et al., 2014	1
			Biswas and Sarker, 2017	1
			Hoque et al., 2014	1
			Hoque et al., 2018	1, 2
			Hossain et al., 2016	1
			Islam et al., 2010	1, 2
			Islam et al., 2011	1
			Islam et al., 2014	1
			Kabir et al., 2017	1, 2
			Kayesh et al., 2014	1
			Meher et al., 2018	1
			Quaderi et al., 2013	1
			Rabbani and Samad, 2010	1
			Rahman et al., 2009	1
			Rahman et al., 2010	1
			Rahman et al., 2014	1
			Sarker et al., 2013	1
			Siddique et al., 2015	1
			Siddique et al., 2013	1
			Sumon et al., 2017	1
			Tripura et al., 2014	1
			Gao et al., 2017	2
			Li et al., 2009	1
			Memon et al., 2013	1, 2
			Wang et al., 2019	1
			Xu et al., 2015	1

(continued on next page)

Table 1 (continued)

No.	Continents	Countries	Studies [Author and Year]	SCM-1, CM-2
Asia	Iran	Indonesia	Yang et al., 2014	1, 2
		Lucia et al., 2017	1	
		Beheshti et al., 2011	1	
		Haghkhah et al., 2010	1, 2	
		Haghkhah et al., 2011	1	
	Iraq	Hashemi et al., 2011	1, 2	
		Jamali et al., 2014	2	
		Kalantari et al., 2013	1	
		Marashifard et al., 2019	1	
		Naghshineh et al., 2015	1	
Asia	Jordan	Reza et al., 2011	1	
		Abdulkadhim, 2012	1	
		Hussein, 2012	1	
		Alekish, 2015	1	
		Hawari and Al-Dabbas, 2008	1, 2	
	Malaysia	Othman and Bahaman, 2005	1	
		Dhakal, 2006	1	
		Dhakal et al., 2007	1, 2	
		Khanal and Pandit, 2013	1	
		Shrestha and Bindari, 2012	1	
Asia	Pakistan	Ali et al., 2011	1	
		Bachaya et al., 2005	1	
		Bachaya et al., 2011	1	
		Baloch et al., 2018	1, 2	
		Chishty et al., 2007	2	
		Farooq et al., 2008	1	
		Hameed et al., 2012	1, 2	
		Hussain et al., 2007	1, 2	
		Hussain et al., 2013	1	
		Khan and Muhammad, 2005	1, 2	
Asia	South Korea	Khan et al., 2004	1, 2	
		Khan et al., 2015	2	
		Muhammad et al., 2010	1	
		Mustafa et al., 2011	1, 2	
		Qayyum et al., 2016	1	
		Rafiullah et al., 2017	1	
		Shahid et al., 2011	1	
		Sharif and Ahmad, 2007	1	
		Umar et al., 2013	1	
		Nam et al., 2010	1	
Asia	Sri Lanka	Sharma et al., 2013	1	
		Sanotharan et al., 2016	1	
		Brunner et al., 2019	2	
		Jarassaeng et al., 2012	1	
		Suriyasathaporn, 2011	1	
		Östensson et al., 2013	1	
		Thanh et al., 2015	1, 2	
		Graaf and Dwinger, 1996	1	
		Maćesić et al., 2016	1	
		Norberg et al., 2004	1, 2	
Europe	Europe	Estonia	Kalmus et al., 2006	2
		Finland	Pyörälä et al., 2011	1, 2
		France	Taponen et al., 2006	1, 2
		Germany	Botrel et al., 2010	1, 2
		Greece	Coulon et al., 2002	2
		Italy	Rupp and Boichard, 2000	2
		Kosovo	Schuberth et al., 2001	1, 2
		Netherlands	Fthenakis, 1998	1, 2
		Norway	Ceniti et al., 2017	2
		Poland	Sylejmani et al., 2016	1
Europe	Europe	Netherlands	Borne et al., 2010	1, 2
		Norway	Miltenburg et al., 1996	2
		Poland	Sampimon et al., 2009	1
		Norway	Bakken, 1981	1
		Poland	Bochniarz and Wawron, 2012	2
		Norway	Bochniarz et al., 2013	1
		Poland	Hameed et al., 2006	1, 2
		Norway	Malinowski et al., 2003	2
		Poland	Sztachańska et al., 2016	1
		Serbia	Marija et al., 2016	1

Table 1 (continued)

No.	Continents	Countries	Studies [Author and Year]	SCM-1, CM-2
Europe	Europe	Slovakia	Zutic et al., 2012	1
		Spain	Idriss et al., 2013	1
		Sweden	Reyes-Jara et al., 2016	2
			Grönlund et al., 2005	1
			Hagnestam et al., 2007	2
			Shitandi and Kihumbu, 2004	1
		Turkey	Baştan et al., 2008	1
			Karahan and Çetinkaya, 2007	1
			Özenç et al., 2008	1
		Ukraine	Türkylimaz et al., 2010	1, 2
Latin America	Latin America	United Kingdom	Brunner et al., 2019	2
		Argentina	Bradley and Green, 2001	2
			Bradley and Green, 2009	2
			Linton and Robinson, 1984	1, 2
		Bolivia	Brunner et al., 2019	2
		Brazil	Dieser et al., 2014	1
			Gonzalez et al., 1980	1
			Edwards et al., 1982	1
			Brunner et al., 2019	2
		Chile	Freitas et al., 2008	1, 2
North America	North America	Colombia	Rall et al., 2014	1
		Ecuador	Silva et al., 2013	1
		Peru	Vieira-da-Motta et al., 2001	1
		Trinidad and Tobago	Brunner et al., 2019	2
		Uruguay	Giannechini et al., 2002	1, 2
		Canada	Lago et al., 2011	2
		Mexico	Brunner et al., 2019	2
			León-Galván et al., 2015	1, 2
			López et al., 2012	1, 2
		United States of America	Olivares-Pérez et al., 2015	1
Oceania	Oceania	Australia	Vázquez et al., 2013	1
			Erskine et al., 2002	2
			Gonzalez et al., 1990	2
			Green et al., 2002	2
			Judge et al., 1997	2
			Morse et al., 1988	2
			Oliver and Mitchell, 1984	1
			Sargeant et al., 1998	2
			White et al., 1986	1, 2
		New Zealand	Wilson et al., 1997	1
Africa	Africa		Wilson et al., 2007	2
			Zeng et al., 2009	1
			Gonzalez et al., 1989	2
			Pinzón-Sánchez and Ruegg, 2010	2
			Daniel et al., 1982	2
			Plozza et al., 2011	1
			Stevenson, 2000	2
			Thompson and Houston, 1967	1
			Brunner et al., 2019	2
			Petrovski et al., 2009	2

on period-wise, zone-wise, state-wise, species-wise, and method-wise are given in Tables 7 and 8. The pooled prevalence estimate of SCM and CM in India were 45% [95% CI 40–49%, PI 11–84%] and 18% [95% CI 14–23%, PI 3–60%] obtained from 50,201 and 26,469 samples, respectively. The forest plot of the studies encompassed for SCM and CM prevalence estimates in India are given in Fig. 5 and Fig. 7. The period-wise scrutiny revealed a higher SCM and CM prevalence during 2011–19 when compared to past periods. The zone-wise and state-wise prevalence of SCM and CM in India are depicted in Figs. 6 and 8 respectively.

Table 2

Details of the mastitis prevalence studies from India included in the meta-analysis.

No.	Zones	Study states	Studies [Author and year]	SCM-1, CM-2
North	Haryana	Bhanot et al., 2012	1	
		Bulla et al., 2006	1	
		Charaya et al., 2013	1	
		Charaya et al., 2014	2	
		Danish et al., 2018	1, 2	
		Guha and Gera, 2011	1	
		Guha et al., 2010	1	
		Guha et al., 2012	1	
		Jain et al., 2009	1	
		Kumar and Sharma, 2002	1	
		Kumar et al., 2007	1	
		Panchal et al., 2016	1	
		Pankaj et al., 2012	1	
		Pankaj et al., 2013	1	
		Sharma and Sindhu, 2007	1, 2	
		Sharma et al., 2006	1	
		Sharma et al., 2012	1	
		Sharma et al., 2018a	1, 2	
		Sharma et al., 2018b	1	
		Sindhu et al., 2009	1	
		Sindhu et al., 2012	1	
		Thakur et al., 2018	1	
		Jingar et al., 2017	2	
Himachal Pradesh Jammu and Kashmir	Himachal Pradesh Jammu and Kashmir	Kaur et al., 2015a	1	
		Ashraf et al., 2018	1	
		Bhat et al., 2016	1	
		Bhat et al., 2017	2	
		Dar et al., 2014	1	
		Qadri et al., 2017	1	
		Sanjolly et al., 2017	1, 2	
		Sharma et al., 2010b	1, 2	
		Sharma et al., 2018c	1, 2	
		Sudhan et al., 2005	1	
Punjab	Punjab	Tufani et al., 2012	1	
		Bansal et al., 2015	1	
		Kaur et al., 2015b	1	
		Mir et al., 2014	1	
		Prabhakar et al., 1995	2	
		Ali et al., 2015	1	
		De and Reena, 2009	1, 2	
		Pachauri et al., 2013	1, 2	
		Prakash et al., 2013	1, 2	
		Sharma et al., 2015	1, 2	
Uttar Pradesh	Uttar Pradesh	Singh et al., 2018	1	
		Tyagi et al., 2013	1	
		Verma et al., 2018	1	
		Uttarakhand	Karabasanavar and Singh, 2013	1
		Mahajan et al., 2011	2	
		Tripathi et al., 2018	1	
		Sunder et al., 2013	1	
		Gogoi et al., 2017	1	
		Sharma and Brintly, 2014	1	
		Bihar	Hardenberg, 2016	1, 2
East	Andaman and Nicobar Assam Bihar Mizoram Sikkim West Bengal	Mizoram	Das et al., 2015	1
		Deka et al., 2013	1, 2	
		Dubal et al., 2010	1, 2	
		Banerjee et al., 2017	1	
		Pranay et al., 2017	1	
		Gujarat	Bhanderi and Garg, 2012	1
		Bhatt et al., 2011	1, 2	
		Devgnia et al., 2015	1	
		Kathiriya et al., 2014	1	
		Patel and Trivedi, 2015	1	
West	Maharashtra	Patel et al., 2012	1	
		Patel et al., 2017	1, 2	
		Thakor and Patel, 2013	1	
		Bhikane et al., 2010	1	
		Chougule et al., 2014	2	
		Joshi and Gokhale, 2006	1	
		Shaikh et al., 2018	1, 2	

Table 2 (continued)

No.	Zones	Study states	Studies [Author and year]	SCM-1, CM-2
South	Rajasthan Andhra Pradesh Karnataka Kerala Tamil Nadu Telangana Central Chhattisgarh Madhya Pradesh	Rajasthan	Swami et al., 2017	1
			Chahar et al., 2005	1
			Jena et al., 2015	1
			Joshi et al., 2013	1
			Langer et al., 2014	1
		Andhra Pradesh	Savita et al., 2018	1
			Sharma et al., 2010a	1
		Karnataka	Kavitha et al., 2009	1
			Manasa et al., 2019	1, 2
			Ramachandraiah et al., 1998	1
Central	Tamil Nadu Telangana Chhattisgarh Madhya Pradesh	Karnataka	Reddy et al., 2015	1
			Harini and Sumathi, 2011	1
			Hegde et al., 2013	1
			Jagadeesh et al., 2016	1
			Kaliwal et al., 2011	1, 2
			Krishnaveni et al., 2014	1, 2
			Kurjogi and Kaliwal, 2011	1, 2
			Kurjogi and Kaliwal, 2014	1, 2
			Prabhu et al., 2015	1, 2
			Preethiran et al., 2015	1
Central	Tamil Nadu Telangana Chhattisgarh Madhya Pradesh	Kerala	Sadashiv and Kaliwal, 2013	1, 2
			Sundareshan et al., 2014	1
			Swamy and Krishnamurthy, 1998	1
			Lakshmi and Jayavardhanan, 2016	1, 2
		Tamil Nadu	Ayyappadas and Renugadevi, 2013	2
			Balaji and Saravanan, 2017	1
			Balaji and Senthilkumar, 2017	1
			Elango et al., 2010	1
			Mubarack et al., 2012	1, 2
			Nithya et al., 2017	1
Central	Telangana Chhattisgarh Madhya Pradesh	Telangana	Selvaraju et al., 2013	1
			Srinivasan et al., 2013	1
			Subramanian et al., 2019	1, 2
			Sukumar and James, 2012	2
			Suresh et al., 2017	2
			Visvanath et al., 2012	2
		Chhattisgarh	Dasohari et al., 2017	1
			Kashyap et al., 2019	1
			Neelesh and Maiti, 2010	1
			Ottawar et al., 2018	1
West	Gujarat Maharashtra	Madhya Pradesh	Sharma et al., 2007	1
			Vishwakarma et al., 2010	1
			Shrivastava et al., 2017	1, 2

The SCM and CM prevalence in India was higher in the Central zone when compared to other zones. Based on state-wise breakdown shown the highest SCM prevalence in West Bengal [75%] followed by Mizoram [65%], Chhattisgarh [63%], Karnataka [58%], and other states. The CM prevalence was high in Andhra Pradesh [44%], Madhya Pradesh [38%], Haryana [31%] followed by the other twelve states in India, and was low in Gujarat [6%] and Maharashtra [6%] states. The cattle species revealed a higher SCM prevalence of 49% than the buffaloes [32%], but for CM prevalence no difference between cattle and buffaloes was observed. The maximum number of studies [85] on SCM prevalence in India employed CMT and indicated SCM prevalence was 43%. By using SCC, the SCM prevalence was 39% and 46% was observed by other methods combined. The method-wise analysis revealed CM prevalence by the CE method and other methods was the same [18%). The SCM prevalence studies from India included for meta-analysis revealed highly significant ($P<0.01$) difference based period-wise, zone-wise, state-wise, species-wise, and method except for the Assam state, which showed no significant difference. The Cochran Q statistics of CM prevalence studies indicated a highly significant ($P<0.01$) difference, which showed the heterogeneity between the studies, except for Maharashtra

Table 3
Subclinical Mastitis (SCM) prevalence estimates in the World based on meta-analysis

No.	Parameters	Period	Number of studies	Total samples	Prevalence (%) [CI at 95% level]	PI (%) at 95% level	Tests for heterogeneity			
							I ² Value (%)	Tau square value	Degrees of Freedom	Cochran Q value
1.	SCM in World	1967-2019	222	2,51,372	42 [38-45]	10-83	99.4	0.931	221	26318.4**
Year-wise										
1.	SCM in World-I	1967-2000	13	1,61,708	40 [27-55]	5-89	99.9	1.157	12	6664.5**
2.	SCM in World-II	2001-2010	54	22,794	38 [31-45]	6-86	98.9	1.379	53	6118.4**
3.	SCM in World-III	2011-2019	155	66,870	43 [40-47]	12-80	98.4	0.739	154	12414.3**
Continent-wise										
1.	SCM in Africa	1987-2019	108	30,324	44 [40-49]	10-84	98.2	0.941	107	4941.7**
2.	SCM in Asia	2004-2019	69	34,491	42 [37-47]	10-82	98.7	0.861	68	7111.5**
3.	SCM in Europe	1981-2016	25	49,209	37 [29-47]	8-80	99.4	0.908	24	2485.7**
4.	SCM in Latin America	1980-2018	11	14,913	34 [17-56]	2-92	99.8	2.391	10	5117.7**
5.	SCM in North America	1984-2015	8	1,10,449	46 [35-56]	15-80	99.2	0.364	7	571.6**
6.	SCM in Oceania	1967 & 2011	2	11,811	36 [21-54]	-	98.1	0.283	1	100.9**
Period-wise										
1.	SCM in Africa-I	1987&2000	2	426	79 [54-93]	-	93.4	0.678	1	28.2**
2.	SCM in Africa-II	2002-2019	106	29,898	43 [39-48]	11-83	98.1	0.897	105	4594.1**
3.	SCM in Asia-II	2004-2019	68	34,463	41 [36-47]	10-81	98.7	0.835	67	7086.3**
4.	SCM in Europe-I	1981-1998	4	38,112	21 [14-30]	9-43	98.4	0.213	3	117.8**
5.	SCM in Europe-II	2001-2016	21	11,097	41 [32-51]	9-82	98.9	0.891	20	2141.9**
6.	SCM in Latin America-I	1980-2000	3	5,435	44 [23-67]	10-84	99.2	0.726	2	548.0**
7.	SCM in Latin America-II	2001-2018	8	9,478	30 [12-58]	1-94	99.7	2.923	7	3857.7**
8.	SCM in North America-I	1984-1997	3	1,06,306	47 [35-59]	0-100	98.2	0.184	2	58.0**
9.	SCM in North America-II	2008-2015	5	4,143	45 [31-60]	16-78	98.0	0.466	4	340.9**
Species-wise										
1.	SCM in Cattle	1967-2019	206	2,42,176	42 [39-45]	10-83	99.4	0.912	205	23489.9**
2.	SCM in Buffaloes	2005-2018	26	8,762	46 [34-57]	7-90	98.7	1.474	25	3025.6**
Method-wise										
1.	SCM by CMT	1980-2019	185	1,14,114	43 [39-46]	10-83	99.0	0.918	184	15121.9**
2.	SCM by SCC	1987-2017	29	1,19,028	46 [36-56]	9-88	99.5	1.193	28	4924.2**
3.	SCM by SFMT	2005-2018	16	5,480	41 [31-52]	10-81	98.1	0.792	15	791.6**
4.	SCM by WST	2008-2018	10	3,360	37 [29-46]	16-65	95.7	0.320	9	199.8**

Note: CI-Confidence Interval, PI-Prediction Interval, CMT-California Mastitis Test, SCC-Somatic Cell Count, SFMT-Surf Field Mastitis Test, WST-White Slide Test, *-Significant ($P<0.05$), **- Highly significant ($P<0.01$).

state showed significant ($P<0.05$) difference and no significant difference for East and West zone, Gujarat state established from the subgroup analysis.

4. Discussion

The studies on SCM and CM prevalence retrieved for this study, from various countries in the World will serve as compiled baseline information at one place for researchers across countries and appropriate stakeholders. In the present report, the SCM and CM prevalence estimates of dairy cows in the World were obtained based on a meta-analysis. The number of studies on SCM prevalence was more than CM from the World and necessitates the importance of SCM in dairying. This might be due to fact that the SCM is not grossly detectable in milk and requires a diagnostic test, so many researchers are working on this, at early detection in dairy animals. More studies on SCM [155] and CM [88] prevalence were reported during 2011-19 signifying the importance of mastitis in dairy animals in the World during the current times.

The number of prevalence studies on SCM and CM was increasing in recent durations and may be attributable to the awareness among the dairy farmers about the quality of milk, which will help them in fetching a good price for the milk produced. Further, the mastitis prevalence studies showed high heterogeneity between the studies included for meta-analysis. This may be owing to various factors comprising of parity of cattle, stage of lactation and genetic breed characters, agroclimatic conditions, weather conditions and practices followed in the management of dairy farms (Joshi and Gokhale, 2006). More SCM studies were reported from the Africa continent than other continents, highlighting the SCM problem there and concurred with a previous report (Abebe et al., 2016). Among African countries, more number of studies was reported from Ethiopia, suggesting mastitis as a major problem in dairy animals due to underdeveloped country. Further, the mastitis studies reported from 61 countries of the World included in this study with more number of countries from the European continent i.e., twenty countries, which suggests the availability of mastitis diagnostic facilities in these countries.

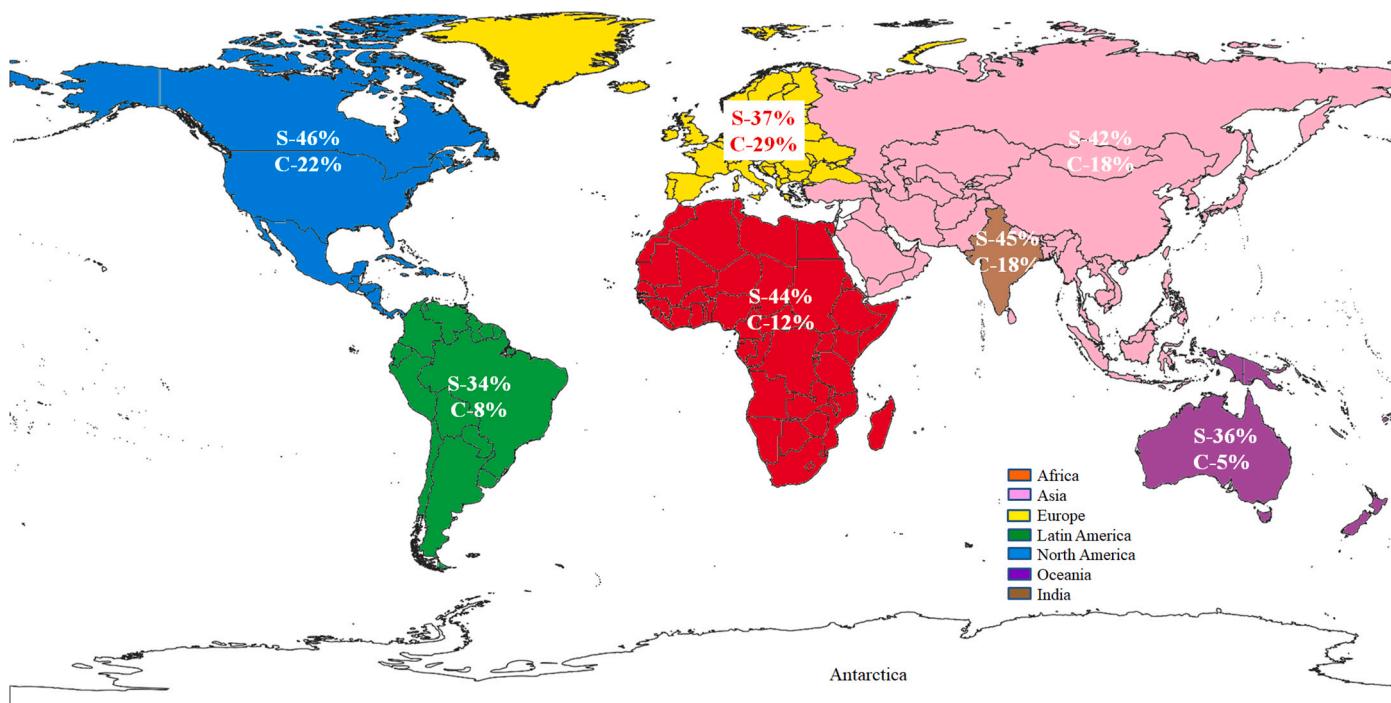


Fig. 4. World map showing prevalence estimates for Subclinical (S) and Clinical (C) mastitis in dairy cattle and buffaloes.

The prevalence estimates were 42% and 15% for SCM and CM in the World and SCM showed the highest prevalence when compared to CM. This indicated the importance of SCM than CM in dairy cattle and buffaloes in the World. In India, the estimated prevalence of SCM was 46.35% obtained from the meta-analysis of studies described during the period 1995–2014 as reported (Bangar et al., 2015) and it was high when compared to the present study. A study reported the SCM prevalence was ranging from 15 to 75 % on a cow basis (Cynthia, 2005) and concurred with the present study. The SCM prevalence estimate was in agreement with the results reported earlier for Bangladesh (Rahman et al., 2010) and Sri Lanka (Sanotharan et al., 2016). However, SCM prevalence was 29.5% (Islam et al., 2011) and 20.2% (Sarker et al., 2013) in Bangladesh as previously reported and considered low compared to this study. The SCM prevalence was higher of late, signifying the need for prevention and control of the same in dairy animals, and might be due to more awareness among the farmers on mastitis. Further, the high prevalence of SCM may be attributed to the low udder immunity in old cows and buffaloes, rearing animals with pendulous udders, and lack of genetic selection of dairy cattle with suitable confirmation of udder for infection prevention as reported (Ahmed et al., 2018). The prevalence estimate was higher in the Africa continent and might be attributed to the lack of implementation of mastitis prevention and control practices by the herd owners in the dairy farms in these African countries as discussed earlier (Abebe et al., 2016). The cattle showed SCM prevalence lesser than the buffaloes and were found to be divergent with a previous study which specified 36% in crossbred cows and 27% in buffaloes in Pakistan (Khan and Muhammad, 2005). In the same way, SCM prevalence was 33% in cows and 8% in buffaloes (Hussain et al., 1984) and differs from the present study. However, Bachaya et al. (2011) described that variations in the SCM prevalence might be due to differences in management practices, diagnostic

methods, breed of the animals, immune response, and prevailing climatic conditions in the particular geographical locations. Method-wise breakdown revealed high SCM prevalence by SCC [46%] and low by WST [37%]. Inflammation of the mammary gland increases the number of somatic cells in milk (Bachaya et al., 2011) and thereby increases the SCC in SCM and CM cases. Thus, the SCC method may be employed for the detection of SCM in dairy cows, but the most commonly followed method for the detection of SCM was CMT and employed in the majority of SCM prevalence studies due to ease of usage in field conditions. However, the SCC may be utilized in the future for accurate diagnosis of SCM in dairy animals, for improved diagnosis and prevention of milk loss due to SCM.

The CM prevalence estimate was 15%, which was less when compared to the SCM, indicating the importance of SCM in the World. The reason may be the easy detection of CM with the physical examination of milk and difficulty in SCM detection, which shows no changes in the milk. Similarly, CM prevalence was reported among dairy cattle in Ethiopia [12.5%] (Zeryehun and Abera, 2017), India [11.5%] (Bhat et al., 2017), and China [3.3%] (Gao et al., 2017) was low in comparison to this study. The year-wise examination indicated the decreasing of CM prevalence in the World over the times and might be due to the awareness among the farmers on CM and easy detection of the same by farmers themselves. In this study, CM prevalence was high in Europe and low in Oceania in comparison to other continents except for Antarctica. The explanations for high prevalence in Europe require further investigation and organisms causing the CM may be different from the other continents, herd-specific factors, milk sampling, and difficult to compare data across dairy farms, regions, and countries as discussed earlier (Brunner et al., 2019). The buffaloes revealed higher CM prevalence than cattle and resulted in the importance of CM in buffaloes in the World. The reason for this might be due to the poor management of

Table 4

Clinical Mastitis (CM) prevalence estimates in the World based on meta-analysis

No.	Parameters	Period	Number of studies	Total samples	Prevalence (%) [CI at 95% level]	PI (%) at 95% level	Tests for heterogeneity			
							I^2 Value (%)	Tau square value	Degrees of Freedom	Cochran Q value
1.	CM in World	1982-2019	150	1,10,066	15 [12-19]	1.81	99.4	2.524	149	29934.3**
Year-wise										
1.	CM in World-I	1982-2000	16	43,442	29 [14-53]	1.97	99.9	3.902	15	7614.3**
2.	CM in World-II	2001-2010	46	25,087	18 [12-27]	1.85	99.4	2.564	45	6293.1**
3.	CM in World-III	2011-2019	88	41,537	12 [9-16]	1.70	98.7	2.024	87	15671.8**
Continent-wise										
1.	CM in Africa	1996-2019	78	26,616	12 [9-15]	1.62	97.8	1.544	77	3978.0**
2.	CM in Asia	2004-2019	25	21,840	18 [10-31]	1.89	99.5	2.966	24	10841.4**
3.	CM in Europe	1988-2019	22	38,976	29 [16-46]	1.95	99.8	3.131	21	4985.2**
4.	CM in Latin America	2002-2019	7	3,581	8 [3-19]	1.75	98.0	1.668	6	1123.6**
5.	CM in North America	1986-2019	14	13,965	22 [9-45]	0.97	99.7	4.213	13	6327.7**
6.	CM in Oceania	1982-2019	3	4,771	5 [1-21]	0.99	98.9	2.109	2	437.5**
Period-wise										
1.	CM in Africa-I	1996-2000	3	5,338	7 [2-29]	0.66	98.8	1.968	2	330.6**
2.	CM in Africa-II	2002-2019	75	21,278	12 [9-15]	1.61	97.7	1.157	74	3107.0**
3.	CM in Asia-II	2004-2019	24	18,075	19 [10-32]	1.90	99.5	3.095	23	10608.2**
4.	CM in Europe-I	1984-1999	5	29,902	39 [12-74]	1.99	99.9	2.808	4	729.4**
5.	CM in Europe-II	2001-2019	17	9,704	26 [13-46]	1.94	99.6	3.180	16	2450.3**
6.	CM in Latin America-II	2002-2019	7	3,581	8 [3-19]	1.75	98.0	1.668	6	1123.6**
7.	CM in North America-I	1986-1998	6	6,421	49 [15-84]	1.99	99.8	4.526	5	3318.2**
8.	CM in North America-II	2002-2019	8	7,544	10 [4-24]	1.82	99.2	2.043	7	985.8**
9.	CM in Oceania-I	1982 & 2000	2	1,781	13 [8-19]	-	82.8	0.095	1	13.1**
Species-wise										
1.	CM in Cattle	1982-2019	143	1,06,181	14 [11-18]	1.79	99.4	2.446	142	28203.7**
2.	CM in Buffaloes	2004-2018	15	3,744	28 [14-47]	1.94	99.0	2.657	14	1199.2**
Method-wise										
1.	CM by CE	1982-2019	146	1,07,177	15 [12-19]	1.80	99.4	2.467	145	28743.9**
2.	CM by OT	2002-2014	4	2,700	13 [2-50]	1.99	99.1	3.816	3	1023.7**

Note: CI-Confidence Interval, PI-Prediction Interval, CE-Clinical Examination, *- Significant ($P<0.05$), **- Highly significant ($P<0.01$).

Table 5
Subclinical Mastitis (SCM) prevalence estimates in various countries in the World based on meta-analysis.

No.	Parameters	Period	Number of studies	Total samples	Prevalence (%) [CI at 95% level]	PI (%) at 95% level	Tests for heterogeneity			
							I^2 Value (%)	Tau square value	Degrees of Freedom	Cochran Q value
Algeria	2013-2018	3	559	46 [31-61]	21-73	91.6	0.265	2	32.1**	
Argentina	1980&2013	2	6,464	56 [54-58]	-	68.9	0.003	1	6.5*	
Australia	1967&2011	2	11,811	36 [21-54]	-	98.1	0.283	1	100.9**	
Austria	1996	1	2,186	24	-	-	-	-	-	
Bangladesh	2009-2018	22	5,363	45 [37-52]	16-77	96.3	0.518	21	687.2**	
Bolivia	1982	1	1,090	19	-	-	-	-	-	
Brazil	2001-2014	4	3,122	27 [11-53]	3-81	99.2	1.197	3	833.5**	
China	2009-2015	5	4,434	39 [26-53]	6-87	98.7	0.449	4	427.5**	
Croatia	2016	1	1,549	49	-	-	-	-	-	
Denmark	2004	1	322	45	-	-	-	-	-	
Ecuador	2018	1	250	60	-	-	-	-	-	
Egypt	2009-2018	15	4,319	41 [30-53]	8-84	97.8	0.829	14	912.0**	
Ethiopia	2002-2019	63	19,806	40 [35-45]	12-76	97.7	0.639	62	2273.0**	
Finland	2006&2011	2	367	54 [48-61]	-	29.0	0.010	1	2.9 ^{NS}	
France	2010	1	1,770	53	-	-	-	-	-	
Germany	2001	1	68	57	-	-	-	-	-	
Greece	1998	1	172	22	-	-	-	-	-	
Indonesia	2017	1	28	82	-	-	-	-	-	
Iran	2009-2019	8	8,613	32 [22-44]	7-75	98.2	0.521	7	211.8**	
Iraq	2012	2	472	50 [36-64]	-	79.1	0.131	1	9.9*	
Jordan	2008&2015	2	353	44 [27-62]	-	91.0	0.264	1	22.7**	
Kenya	2013-2019	6	734	37 [20-57]	2-92	96.2	1.001	5	98.1**	
Kosovo	2016	1	156	26	-	-	-	-	-	
Malaysia	2005	1	60	82	-	-	-	-	-	
Mexico	2012-2015	4	4,042	43 [27-61]	2-96	98.6	0.542	3	340.0**	
Nepal	2006-2013	4	818	22 [8-49]	1-99	97.2	1.510	3	191.4**	
Nigeria	2011-2017	4	1,999	39 [29-50]	7-83	93.8	0.178	3	52.7**	
Netherlands	2009&2010	2	436	24 [20-28]	-	0.0	0.0	1	2.66 ^{NS}	
Norway	1981	1	35,464	31	-	-	-	-	-	
Pakistan	2005-2018	17	11,992	42 [30-55]	6-88	99.3	1.197	16	4162.2**	
Peru	2015	1	459	72	-	-	-	-	-	
Poland	2006-2016	3	1,314	30 [25-36]	2-91	77.8	0.042	2	15.4**	
Rwanda	2015-2019	3	574	60 [46-74]	32-83	91.4	0.259	2	38.5**	
Serbia	2012&2016	2	1,397	25 [19-32]	-	84.5	0.051	1	12.7**	
Slovakia	2013	1	390	74	-	-	-	-	-	
South Korea	2010&2013	2	895	55 [52-58]	-	0.0	0.0	1	0.7 ^{NS}	
Sri Lanka	2016	1	152	43	-	-	-	-	-	
Sudan	2012	1	60	23	-	-	-	-	-	
Sweden	2004&2005	2	403	74 [55-87]	-	92.5	0.344	1	27.3**	
Tanzania	2000-2017	6	959	61 [40-78]	6-97	96.6	1.061	5	151.7**	
Thailand	2011&2012	2	353	35 [30-40]	-	0.0	0.0	1	1.7 ^{NS}	
Trinidad and Tobago	2000	1	177	60	-	-	-	-	-	
Turkey	2007-2010	4	2,925	30 [11-60]	1-99	99.3	1.664	3	852.3**	
Uganda	2014-2017	3	390	85 [81-88]	17-99	34.3	0.033	2	4.9 ^{NS}	
United Kingdom	1984	1	290	11	-	-	-	-	-	
Uruguay	2002	1	3,351	1	-	-	-	-	-	
United States of America	1984-2008	4	1,06,407	49 [39-58]	13-86	96.9	0.147	3	59.4**	
Vietnam	2016&2015	2	1,001	52 [36-67]	-	96.2	0.216	1	53.8**	
Zambia	2013	1	111	49	-	-	-	-	-	
Zimbabwe	1987&2013	2	945	57 [8-95]	-	99.4	3.635	1	540.6**	

Note: CI-Confidence Interval, PI-Prediction Interval, NS-Non significant, *- Significant ($P<0.05$), **- Highly significant ($P<0.01$).

Table 6

Clinical Mastitis (CM) prevalence estimates in various countries in the World based on meta-analysis

No.	Parameters	Period	Number of studies	Total samples	Prevalence (%) [CI at 95% level]	PI (%) at 95% level	Tests for heterogeneity			
							I ² Value (%)	Tau square value	Degrees of Freedom	Cochran Q value
1.	Algeria	2018	1	222	6	-	-	-	-	-
2.	Argentina	2019	1	720	5	-	-	-	-	-
3.	Australia	1982 & 2000	2	1,781	13 [8-19]	-	82.8	0.095	1	13.1**
4.	Bangladesh	2010-2018	4	1,362	8 [3-22]	1.96	95.9	1.414	3	59.8**
5.	Brazil	2008, 2019	2	1,192	3 [1-10]	-	91.7	0.952	1	23.2**
6.	Canada	2011	1	214	22	-	-	-	-	-
7.	Chile	2019	1	159	9	-	-	-	-	-
8.	China	2012-2016	3	4,863	34 [5-85]	0.98	99.8	4.332	2	3210.9**
9.	Colombia	2019	1	183	5	-	-	-	-	-
10.	Denmark	2004	1	322	81	-	-	-	-	-
11.	Ecuador	2018	1	250	12	-	-	-	-	-
12.	Egypt	2010-2015	10	2,721	22 [10-43]	1.92	98.5	2.409	9	815.7**
13.	Estonia	2006	1	2,355	13	-	-	-	-	-
14.	Ethiopia	2002-2019	55	15,822	9 [7-12]	2.41	96.0	0.908	54	1338.5**
15.	Finland	2006 & 2011	2	367	45 [40-52]	-	29.0	0.010	1	2.9 ^{NS}
16.	France	1999-2010	3	22,604	30 [15-50]	7.70	99.6	0.563	2	1118.9**
17.	Germany	2001	1	68	43	-	-	-	-	-
18.	Greece	1998	1	172	17	-	-	-	-	-
19.	Iran	2006-2014	4	6,738	13 [2-49]	0.90	99.1	3.413	3	562.8**
20.	Italy	2017	1	110	47	-	-	-	-	-
21.	Jordan	2008	1	220	16	-	-	-	-	-
22.	Kenya	2017	1	50	54	-	-	-	-	-
23.	Mexico	2012-2019	3	2,864	3 [1-12]	0.34	96.6	1.416	2	109.4**
24.	Nepal	2007	1	355	6	-	-	-	-	-
25.	Netherlands	1996 & 2010	2	9,391	21 [10-39]	-	98.4	0.388	1	110.8**
26.	New Zealand	2009 & 2019	2	6,755	3 [0-25]	-	99.0	2.652	1	546.2**
27.	Nigeria	1999 & 2017	2	5,360	2 [2-3]	-	0.0	0.0	1	1.36 ^{NS}
28.	Pakistan	2004-2018	8	3,794	19 [10-33]	2.77	98.7	1.066	7	371.0**
29.	Poland	2003-2012	3	1,007	4 [1-23]	0.67	95.9	2.974	2	77.4**
30.	Seychelles	1996	1	273	35	-	-	-	-	-
31.	South Africa	2019	1	764	37	-	-	-	-	-
32.	Spain	2016	1	327	33	-	-	-	-	-
33.	Sudan	2012	1	60	77	-	-	-	-	-
34.	Sweden	2007	1	506	59	-	-	-	-	-
35.	Tanzania	2000-2013	5	818	19 [7-46]	1.97	97.7	1.885	4	152.8**
36.	Thailand	2019	1	83	1	-	-	-	-	-
37.	Turkey	2010	1	244	50	-	-	-	-	-
38.	Uganda	2013	1	138	13	-	-	-	-	-
39.	Ukraine	2019	1	164	1	-	-	-	-	-
40.	United Kingdom	1984-2015	4	1,570	74 [64-81]	27.95	90.9	0.158	3	43.9**
41.	United States of America	1986-2007	10	10,887	35 [14-64]	1.98	99.8	3.729	9	5015.5**
42.	Uruguay	2002	1	1,077	52	-	-	-	-	-
43.	Vietnam	2015	1	550	22	-	-	-	-	-
44.	Zimbabwe	2016	1	584	5	-	-	-	-	-

Note: CI-Confidence Interval, PI-Prediction Interval, ^{NS}-Non significant, **- Highly significant ($P<0.01$).

Table 7

Subclinical mastitis (SCM) prevalence estimates in India based on period-wise, zone-wise, states-wise, species-wise and method-wise.

No.	Parameters	Period	Number of studies	Total samples	Prevalence (%) [CI at 95% level]	PI (%) at 95% level	Tests for heterogeneity			
							I ² Value (%)	Tau square value	Degrees of Freedom	Cochran Q value
1.	SCM in India	1998-2019	103	50,201	45 [40-49]	11-84	98.8	0.863	102	7172.1**
2a.	SCM in India-Period I	1998-2010	25	12,238	42 [33-53]	8-87	98.8	1.056	24	1743.9**
2b.	SCM in India-Period II	2011-2019	78	37,963	46 [41-51]	12-83	98.7	0.802	77	5337.7**
3.	SCM in North zone	2002-2018	44	25,522	40 [33-47]	8-83	99.0	0.938	43	4890.8**
3a.	Haryana	2002-2018	21	18,970	35 [26-44]	7-80	99.3	0.892	20	3692.6**
3b.	Himachal Pradesh	2015	1	935	44	-	-	-	-	-
3c.	Jammu and Kashmir	2005-2018	9	3,041	47 [32-62]	8-90	98.1	0.854	8	682.9**
3d.	Punjab	2014 & 2015	3	1,277	30 [14-52]	0-99	97.5	0.665	2	129.7**
3e.	Uttarakhand	2013 & 2018	2	292	39 [20-62]	-	92.9	0.429	1	28.4**
3f.	Uttar Pradesh	2009-2018	8	1,007	49 [31-67]	6-94	96.4	1.108	7	149.2**
4.	SCM in East zone	2010-2017	9	1,675	48 [30-67]	5-94	97.7	1.326	8	449.0**
4a.	Andaman & Nicobar	2013	1	25	12	-	-	-	-	-
4b.	Assam	2014 & 2017	2	213	27 [17-40]	-	71.0	0.129	1	7.2 ^{NS}
4c.	Bihar	2016	1	313	35	-	-	-	-	-
4d.	Mizoram	2013 & 2015	2	60	65 [17-94]	-	87.0	2.245	1	21.3**
4e.	Sikkim	2010	1	276	25	-	-	-	-	-
4f.	West Bengal	2017	2	788	75 [46-91]	-	98.0	0.798	1	112.2**
5.	SCM in West zone	2005-2019	18	14,279	41 [33-50]	12-77	97.6	0.531	17	588.8**
5a.	Gujarat	2011-2016	8	11,796	33 [23-45]	8-76	96.7	0.484	7	296.3**
5b.	Maharashtra	2006-2019	4	1,199	38 [27-50]	6-85	90.6	0.209	3	46.4**
5c.	Rajasthan	2013-2019	7	734	37 [20-57]	6-83	92.1	0.387	6	161.6**
6.	SCM in South zone	1998-2019	26	5,319	51 [43-59]	16-85	96.6	0.633	25	549.9**
6a.	Andhra Pradesh	1998-2019	4	732	30 [15-53]	0-98	95.7	0.865	3	104.0**
6b.	Karnataka	1998-2016	12	3,081	58 [49-67]	24-86	95.8	0.394	11	241.2**
6c.	Kerala	2016	1	80	35	-	-	-	-	-
6d.	Tamil Nadu	2010-2019	8	1,290	51 [39-63]	15-86	94.4	0.478	7	143.8**
6e.	Telangana	2017	1	136	66	-	-	-	-	-
7.	SCM in Central zone	2007-2019	6	3,406	63 [49-75]	17-94	98.0	0.510	5	438.6**
7a.	Chhattisgarh	2007-2018	5	3,321	63 [46-77]	10-96	98.6	0.606	4	436.3**
7b.	Madhya Pradesh	2017	1	85	62	-	-	-	-	-
8a.	SCM in Cattle	1998-2019	81	39,294	49 [44-53]	14-84	98.6	0.747	80	5716.7**
8b.	SCM in Buffaloes	1998-2019	30	10,907	32 [25-39]	6-76	98.1	0.839	29	934.5**
9a.	SCM by CMT	1998-2019	85	34,929	43 [38-47]	12-80	98.3	0.721	84	4889.4**
9b.	SCM by SCC	2006-2019	34	8,712	39 [31-48]	7-85	98.1	1.112	33	2165.4**
9c.	SCM by Other methods	1998-2018	19	13,639	46 [35-57]	10-87	99.0	0.905	18	858.9**

Note: CI-Confidence Interval, PI-Prediction Interval, CMT-California Mastitis Test, NS-Not significant, **- Highly Significant ($P<0.01$).

buffaloes than cows in many countries in the World. In contrast, there is a notion that the buffaloes are resistant to infection due to tight streak canal surrounded by tight muscles and also less population restricted to few continents in the World, with more of the nondescript breed of buffaloes are being reared. Method-wise subgroup analysis revealed a higher CM prevalence by clinical examination and the most commonly used method for CM diagnosis than other methods and concurred with an earlier report (Tuke et al., 2017).

The countrywide SCM prevalence was high in Uganda [85%] and low in Uruguay [1%). The possible explanations for the high prevalence in Uganda could be attributed to the risk factors for SCM prevalence were restriction of cows without grazing, poor udder hygiene practices, and more fraction of cows in late parity and late stages of lactation as described earlier (Abrahmsén et al., 2014). The low SCM prevalence in Uruguay was attributed to the ecological influence on the occurrence of mastitis as discussed earlier (Giannechini et al., 2002). Countrywide breakdown suggested high CM prevalence in the United Kingdom and low in Brazil, New Zealand countries in the World based on meta-analysis was observed. The possible reasons for high prevalence in the

United Kingdom could be the reliability of identification and detection of CM cases by the herdsman because they were well-motivated and with improved awareness of mastitis throughout the study as reported (Bradley and Green, 2001). Petrovski et al. (2009) stated that the number of cows included in the analysis and cows with CM was less in number in the farms sampled in New Zealand as indicated, which might be the reason for low prevalence. Further, fewer studies were available for inclusion in a meta-analysis from Brazil and New Zealand may be the reason for low CM prevalence compared to other countries.

The overall SCM prevalence estimate in India was 45%, and was in agreement with previous studies reported based on meta-analysis were 46.35% (Bangar et al., 2015) and 41% (Krishnamoorthy et al., 2017). In recent times, the SCM prevalence was high during 2011-19 in contrast with 1998-2010. This could be due to better diagnostic methods and awareness of SCM by the dairy farmers. Bangar et al. (2015) indicated that the high SCM prevalence over recent years could be attributed as the principal cause for the low milk productivity of dairy cattle and buffaloes in India. Among the studies, more numbers were from North and South zones in India, which may perhaps be due to the presence of

Table 8

Clinical mastitis (CM) prevalence estimates in India based on period-wise, zone-wise, states-wise, species-wise and method-wise.

No.	Parameters	Period	Number of studies	Total samples	Prevalence (%) [CI at 95% level]	PI (%) at 95% level	Tests for heterogeneity			
							I ² Value (%)	Tau square value	Degrees of Freedom	Cochran Q value
1.	CM in India	1995-2019	37	26,469	18 [14-23]	3.60	98.8	0.847	36	1900.4**
2a.	CM in India-Period I	1995-2010	5	6,759	15 [12-19]	7.31	78.8	0.065	4	27.3**
2b.	CM in India-Period II	2011-2019	32	19,710	19 [14-25]	3.64	98.8	0.969	31	1702.6**
3.	CM in North zone	1995-2018	15	21,834	19 [13-27]	3.62	99.4	0.726	14	980.3**
3a.	Haryana	2007-2018	5	14,010	31 [20-45]	4.81	99.4	0.421	4	604.9**
3b.	Jammu and Kashmir	2010-2018	4	1,165	10 [5-19]	0.74	92.0	0.417	3	21.6**
3c.	Punjab	1995	1	421	9	-	-	-	-	-
3d.	Uttarakhand	2011	1	5,698	21	-	-	-	-	-
3e.	Uttar Pradesh	2009-2015	4	540	21 [11-36]	1.90	89.2	0.513	3	30.4**
4.	CM in East zone	2010-2016	3	601	14 [11-18]	2.54	12.8	0.007	2	3.0 ^{NS}
4a.	Bihar	2016	1	285	12	-	-	-	-	-
4b.	Mizoram	2013	1	40	17	-	-	-	-	-
4c.	Sikkim	2010	1	276	25	-	-	-	-	-
5.	CM in West zone	2011-2019	4	1,472	6 [5-7]	4.9	0.0	0.0	3	4.6 ^{NS}
5a.	Gujarat	2011 & 2016	2	654	6 [4-8]	-	0.0	0.0	1	0.18 ^{NS}
5b.	Maharashtra	2014 & 2019	2	818	6 [5-8]	-	0.0	0.0	1	4.4*
6.	CM in South zone	2011-2019	14	2,477	22 [14-32]	3.71	95.8	0.929	13	509.4**
6a.	Andhra Pradesh	2019	1	61	44	-	-	-	-	-
6b.	Karnataka	2011-2015	6	1,269	18 [9-33]	1.82	96.1	1.011	5	292.3**
6c.	Kerala	2016	1	80	24	-	-	-	-	-
6d.	Tamil Nadu	2012-2019	6	1,067	23 [12-40]	2.84	95.8	0.925	5	204.2**
7.	CM in Central zone	2017	1	85	38	-	-	-	-	-
7a.	Madhya Pradesh	2017	1	85	38	-	-	-	-	-
8a.	CM in Cattle	2007-2019	32	20,319	19 [14-24]	4.58	98.5	0.762	31	1350.5**
8b.	CM in Buffaloes	1995-2019	12	6,150	19 [11-30]	2.75	98.9	1.209	11	636.2**
9a.	CM by CE	1995-2019	30	18,664	18 [14-25]	3.64	98.7	0.997	29	1653.7**
9b.	CM by Other methods	2007-2018	7	7,805	18 [13-24]	5.47	96.1	0.252	6	204.4**

Note: CI-Confidence Interval, PI-Prediction Interval, CE- Clinical Examination, NS-Not Significant, *- Significant ($P<0.05$), **- Highly significant ($P<0.01$).

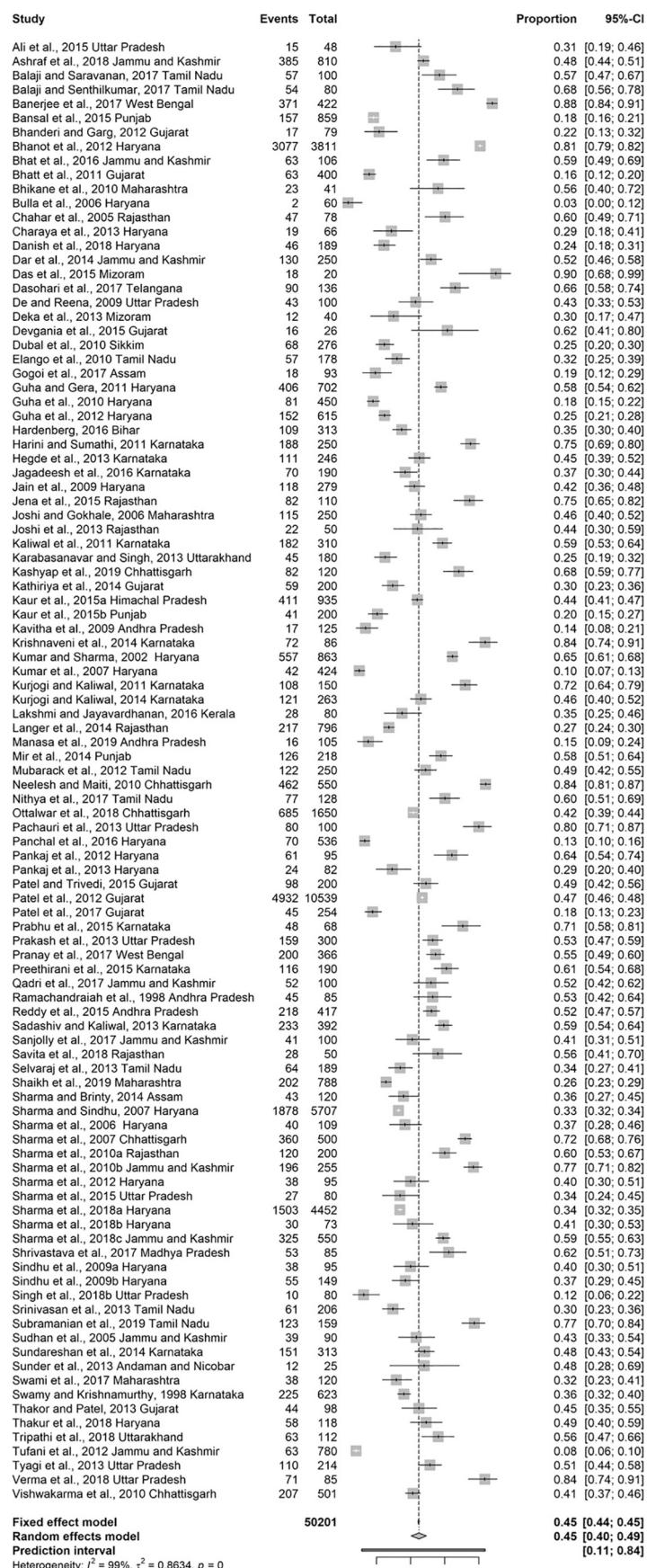


Fig. 5. Forest plot showing the Subclinical mastitis studies from India and their prevalence estimates.

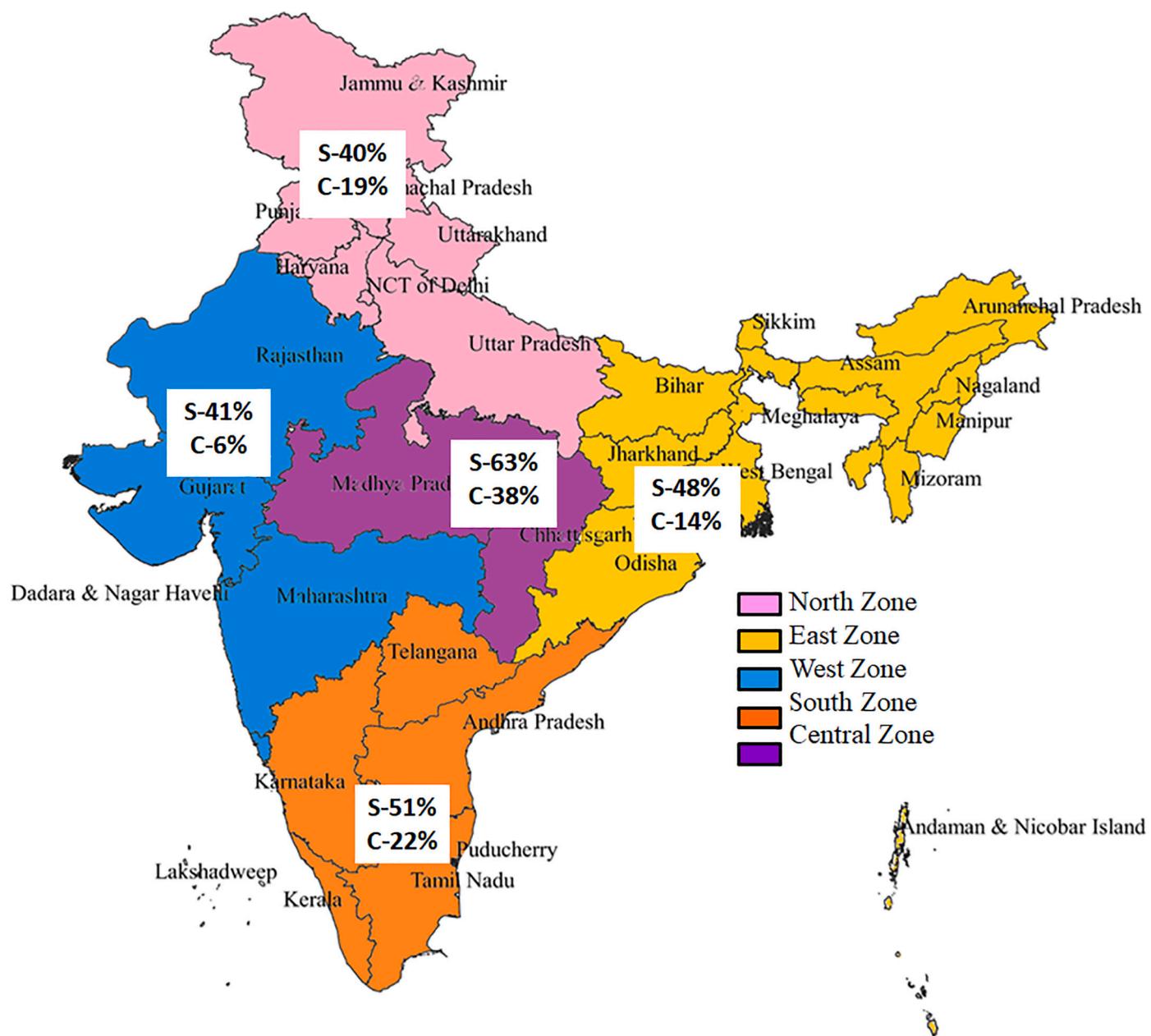


Fig. 6. India map showing the zone-wise prevalence of Subclinical (S) and Clinical (C) mastitis.

more mastitis cases and also more cattle and buffalo population in these zones. Zone-wise determination indicated the high prevalence in the Central zone which includes Chhattisgarh and Madhya Pradesh states. As suggested, the place where the dairy cows were kept, was not suitable for milking, and also the milkman not following the proper methods of clean milk production (Kashyap et al., 2019). Might also due to fact that the manual milking methods in unorganized farms were a major predisposing factor for increased mastitis prevalence (Ottalwar et al., 2018) in these states in the Central zone. The cattle showed high SCM prevalence compared to buffaloes, which confirms the assumption that the buffaloes are less susceptible to mastitis because of tight sphincter

muscles around the streak canal in the teat. In India, mostly the SCM was diagnosed by using CMT, and rarely SCC and bacterial culture examination were followed, and hence the more number of studies encompassed for meta-analysis have used CMT than other methods. Iraguha et al. (2015) stated that the CMT was the most desirable screening test for SCM in field situations and comparable highly to other diagnostic tests. But the SCC and bacterial culture examination are the more sensitive methods for SCM diagnosis from milk containing major mastitis pathogens. Furthermore, the positive CMT reaction depends on the concentration of somatic cells in the milk (Sharma et al., 2018b). The pooled prevalence estimate of CM in India was 18% in this study and

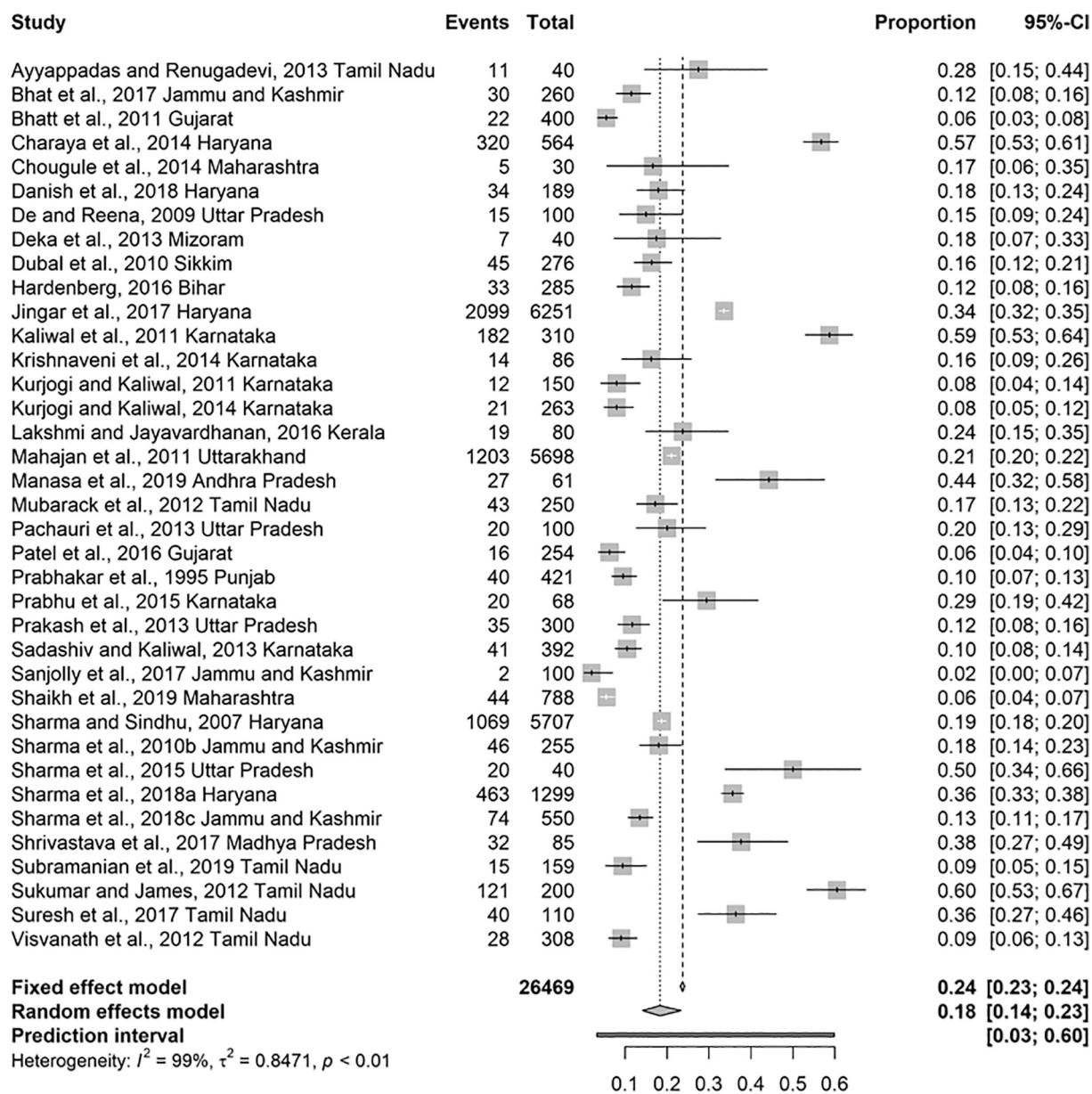


Fig. 7. Forest plot showing the Clinical mastitis studies from India and their prevalence estimates.

concurred with the previous study which reported as 16.08% (Bangar et al., 2016). However, another study reported a CM prevalence of 27% based on a meta-analysis (Krishnamoorthy et al., 2017) and was high in comparison to this study. This could be attributed to the variation in the study period duration considered for meta-analysis which was more in the present study than the previous study. Furthermore, this might be due to the increasing trend in the CM prevalence during 2011-19 of late due to enhanced awareness on the milk quality and mastitis among the dairy farmers. High CM prevalence was observed in the Central zone as that of the SCM and the reasons attributed may be the same as discussed earlier. The variation in the prevalence of mastitis amongst studies may be partly due to diagnostic tests used, sampling strategies followed, and

criteria for mastitis and the risk factors encompassing the stage of lactation, parity number, cattle, and buffalo breeds comprised in the studies (Shrivastava et al., 2017). There was no difference in CM prevalence estimates attained based on species-wise (cattle and buffaloes) and method-wise (CE and other methods) was observed. The clinical examination was the predominantly adopted method in reporting CM prevalence by the majority of the studies in both the World and India. Based on the findings from this study, the high-risk zone for the SCM and CM was the central zone and the states with high prevalence were known. This will be helpful for the policymakers and various stakeholders in implementing preventive measures for mastitis. Further, there is an urgent need for the farmers to adopt the dairy management using

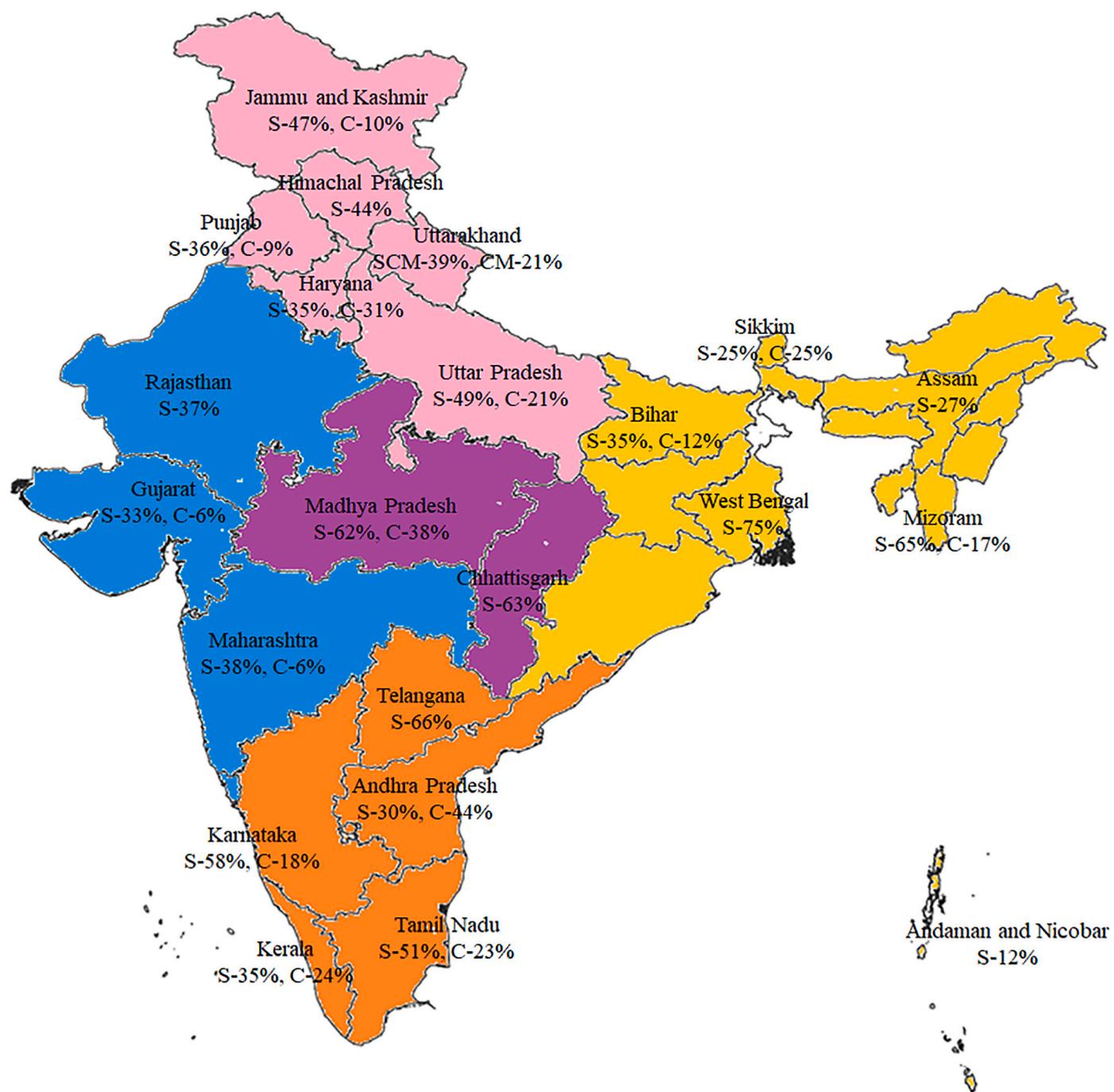


Fig. 8. India map showing the state-wise prevalence of subclinical (S) and clinical (C) mastitis.

scientific approaches by embracing dairy extension methods with mastitis awareness focus, practices of clean milk production, regular screening for mastitis, dry cow therapy, and culling of animals with chronic mastitis from the dairy farms.

In this study, there are limitations or constraints in the prevalence estimates obtained by using meta-analysis, as it might not provide the estimate for the association of SCM and CM prevalence with various risk factors in the studies reported. Further, several factors that might lead to variation between the studies are a breed of the cattle and buffaloes, the genetic makeup of the dairy animals, lactation number, stage of lactation, potential milk yield, dairy managemental practices, weather

conditions, geographical area, spatial and temporal disparities in studies reported from various countries of the World. To compare the results between diverse studies, the common diagnostic test for SCM and CM prevalence were considered as diagnostic criteria and most studies included CMT for SCM and CE for CM diagnosis. Furthermore, this study forms the first report of SCM and CM prevalence in the World and its various countries by meta-analysis method except for India. Thus, the literatures collected will form a database on SCM and CM prevalence studies at one place for easy access to researchers working on mastitis in the future.

5. Conclusion

In conclusion, the present study estimated the pooled prevalence of SCM and CM in the World for the first time by using a meta-analysis method. A high prevalence of SCM than CM was observed in the World, indicated the importance of SCM. It also advocates the need for scientific dairy management methods to be adopted in various countries to reduce the SCM and CM prevalence in the World. The continent-wise and countrywide SCM and CM prevalence estimates will help the various stakeholders concerned with dairying, in developing the infection prevention and control strategies to be followed in dairy farms. In India, the detailed analysis and prevalence estimates will help the various zones and states to take up the mastitis prevention and control methods, based on the severity, and also helps in the clean milk production. The prevalence estimates will help the policymakers and stakeholders to make an informed decision on prevention strategies for mastitis by using scarce resources. Furthermore, the current study provided the metadata on mastitis prevalence studies from 61 countries of the World in this report and will be very useful for the researchers and various stakeholders. However, there is a need for more studies on mastitis prevalence from other countries in the World that are not included in this study, for obtaining more accurate prevalence estimates. To reduce the occurrence of mastitis in the World, there should be a timely diagnosis and therapeutic interventions by field Veterinarians and scientific management of dairy farms both in organized and unorganized sectors, to improve milk production and feed the world population. Furthermore, mastitis is present for a long period of more than 100 years in dairy animals and suitable vaccination or newer strategies may be implemented based on the causative organisms prevalent in particular geographical areas in the World.

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.rvsc.2021.04.021>.

Declaration of Competing Interest

The authors declare no conflict of interest.

Acknowledgment

The authors thank the Indian Council of Agricultural Research (ICAR), New Delhi for providing necessary support in undertaking this research work under the Institute research project.

References

- Abdel-Rady, A., Sayed, M., 2009. Epidemiological studies on subclinical mastitis in dairy cows in Assiut Governorate. *Vet. World* 2, 373–380.
- Abd-Elrahman, A.H., 2013. Mastitis in housed dairy buffaloes: incidence, etiology, clinical finding, antimicrobial sensitivity and different medical treatment against *E. coli* mastitis. *Life Sci. J.* 10, 532–538.
- Abdulkadhim, M.H., 2012. Prevalence of methicillin resistance *Staphylococcus aureus* in cattle and she-camels milk at Al-Qadisyia Province. *Al-Anbar J. Vet. Sci.* 5, 63–67.
- Abeye, R., Hatiya, H., Abera, M., Megersa, B., Asmare, K., 2016. Bovine mastitis: prevalence, risk factors and isolation of *Staphylococcus aureus* in dairy herds at Hawassa milk shed. *Southern Ethiopia.* *BMC Vet. Res.* 12, 270.
- Abera, M., Elias, B., Aragaw, K., Denberga, Y., Amenu, K., Sheferaw, D., 2012a. Major causes of mastitis and associated risk factors in smallholder dairy cows in Shashemene, southern Ethiopia. *Afr. J. Agric. Res.* 7, 3513–3518.
- Abera, M., Habte, T., Aragaw, K., Asmare, K., Sheferaw, D., 2012b. Major causes of mastitis and associated risk factors in smallholder dairy farms in and around Hawassa, Southern Ethiopia. *Trop. Anim. Health Prod.* 44, 1175–1179.
- Abera, B., Lemma, D., Iticha, I., 2013. Study of bovine mastitis in Asella government dairy farm of Oromia Regional state, South Eastern Ethiopia. *Intl. J. Curr. Res. Aca. Rev.* 1, 134–145.
- Abraham, F., Zeleke, M.M., 2017. Prevalence of bovine clinical mastitis and farmer's awareness in and around Wolaita Sodo, Southern Ethiopia. *Adv. Dairy Res.* 5, 3.
- Abrahamsén, M., Persson, Y., Kanyima, B.M., Båge, R., 2014. Prevalence of subclinical mastitis in dairy farms in urban and peri-urban areas of Kampala, Uganda. *Trop. Anim. Health Prod.* 46, 99–105.
- Abunna, F., Fufa, G., Megersa, B., Regassa, A., 2013. Bovine mastitis: prevalence, risk factors and bacterial isolation in small-holder dairy farms in Addis Ababa City. *Ethiopia. Glob. Vet.* 10, 647–652.
- Adane, B., Guyo, K., Tekle, Y., Taddele, H., Bogale, A., Biffa, D., 2012. Study on prevalence and risk factors of bovine mastitis in Borana pastoral and agro-pastoral settings of Yabello district, Borana zone, Southern Ethiopia. *American-Eurasian J. Agri. Environ. Sci.* 12, 1274–1281.
- Afri, F., Fartas, A., Bouzebda, Z., Khamassi, S., 2017. Prevalence and impact of subclinical mastitis on the profitability of dairy cattle in the Far East of Algeria. *Livest. Res. Rural Dev.* 29, 182.
- Ahmed, H.F., Straubinger, R.K., Hegazy, Y.M., Ibrahim, S., 2018. Subclinical mastitis in dairy cattle and buffaloes among small holders in Egypt: Prevalence and evidence of virulence of *Escherichia coli* causative agent. *Trop. Biomed.* 35, 321–329.
- Akkou, M., Bouchiat, C., Antri, K., Bes, M., Tristan, A., Dauwalder, O., Martins-Simões, P., Rasigade, J.P., Etienne, J., Vandenesch, F., Ramdani-Bouguessa, N., 2018. New host shift from human to cows within *Staphylococcus aureus* involved in bovine mastitis and nasal carriage of animal's caretakers. *Vet. Microbiol.* 223, 173–180.
- Alekish, M.O., 2015. The association between the somatic cell count and isolated microorganisms during subclinical mastitis in heifers in Jordan. *Vet. Med.* 60, 71–76.
- Alemu, S., Tamiru, F., Almaw, G., Tsega, A., 2013. Study on bovine mastitis and its effect on chemical composition of milk in and around Gondar Town. *Ethiopia. J. Vet. Med. Anim. Health* 5, 215–221.
- Ali, M.A., Ahmad, M.D., Muhammad, K., Anjum, A.A., 2011. Prevalence of subclinical mastitis in dairy buffaloes of Punjab, Pakistan. *The J. Anim. Plant Sci.* 21, 477–480.
- Ali, Z., Dimri, U., Jhambh, R., 2015. Prevalence and antibiogram of bacterial pathogens from subclinical mastitis in buffaloes. *Buff. Bul.* 34, 41–44.
- Almaw, G., Zerihun, A., Asfaw, Y., 2008. Bovine mastitis and its association with selected risk factors in smallholder dairy farms in and around Bahir Dar. *Ethiopia. Trop. Anim. Health Pro.* 40, 427–432.
- Almaw, G., Molla, W., Melaku, A., 2009. Prevalence of bovine subclinical mastitis in Gondar town and surrounding areas. *Ethiopia. Livest. Res. Rural Dev.* 21, 7.
- Amer, S., Gálvez, F.L.A., Fukuda, Y., Chika, T.A.D.A., Jimenez, I.L., Valle, W.F.M., Nakai, Y., 2018. Prevalence and etiology of mastitis in dairy cattle in El Oro Province. *Ecuador. J. Vet. Med. Sci.* 80, 861–868.
- Amin, A.S., Hamouda, R.H., Abdel-All, A.A., 2011. PCR assays for detecting major pathogens of mastitis in milk samples. *World J. Dairy Food Sci.* 6, 199–206.
- Amin, B., Deneke, Y., Abdela, N., 2017. Bovine Mastitis: Prevalence, Risk Factors and Isolation of *Streptococcus* Species from Small Holders Dairy Farms in and Around Haramaya Town, Eastern Ethiopia. *Glob. J. Med. Res.* 17, 27–38.
- Ashraf, I., Malik, H.U., Muheet, A., Shah, O., Amin, U., Beigh, S.A., Bashir, S., Handoo, N., Hammadani, H., Nazir, N., Jan, A., 2018. Longitudinal study on prevalence of sub clinical mastitis in winter season of cattle from Kashmir valley. *Pharma Innov. J.* 7, 988–989.
- Ayano, A.A., Hiriko, F., Simyalew, A.M., Yohannes, A., 2013. Prevalence of subclinical mastitis in lactating cows in selected commercial dairy farms of Holeta district. *J. Vet. Med. Anim. Health* 5, 67–72.
- Ayyappadas, M.P., Renugadevi, R., 2013. Study on the prevalence, incidence, drug sensitivity and extra chromosomal impact on bovine mastitis pathogens. *J. Agri. Vet. Sci.* 2, 60–62.
- Bachaya, H.A., Iqbal, Z., Muhammad, G., Yousaf, A., Ali, H.M., 2005. Subclinical mastitis in buffaloes in Attock district of Punjab (Pakistan). *Pak. Vet. J.* 25, 134–136.
- Bachaya, H.A., Raza, M.A., Murtaza, S., Akbar, I.U.R., 2011. Subclinical bovine mastitis in Muzaffar Garh district of Punjab (Pakistan). *J. Anim. Plant Sci.* 21, 16–19.
- Badiuzzaman, M., Samad, M.A., Siddiki, S.H.M.F., Islam, M.T., Saha, S., 2015. Subclinical mastitis in lactating cows: Comparison of four screening tests and effect of animal factors on its occurrence. *Bangladesh J. Vet. Med. Dis.* 13, 41–50.
- BAHS, 2019. Basic animal husbandry statistics. Ministry of Fisheries, Animal Husbandry and Dairying, Department of Animal Husbandry and Dairying, Krishi Bhawan, New Delhi, India.
- Bakken, G., 1981. Subclinical mastitis in Norwegian dairy cows: prevalence rates and epidemiological assessments. *Acta Agric. Scand.* 31, 273–286.
- Balaji, S.N., Saravanan, R., 2017. Prevalence of subclinical mastitis in dairy cows of Salem district in Tamilnadu. *Intl. J. Sci. Environ. Technol.* 6, 1772–1776.
- Balaji, N., Senthilkumar, A., 2017. Prevalence of subclinical mastitis in dairy cows of Theni district in Tamilnadu. *Intl. J. Sci. Environ. Technol.* 6, 3427–3432.
- Baloch, H., Rind, R., Rind, M.R., Kumar, V., Baloch, N., Oad, R.K., 2018. Effect of diverse factors on the frequency of clinical and subclinical mastitis in Kundhi buffaloes of Sindh. *Pak. J. Zool.* 50, 1619–1628.
- Banerjee, S., Batabyal, K., Joardar, S.N., Isore, D.P., Dey, S., Samanta, I., Samanta, T.K., Murmu, S., 2017. Detection and characterization of pathogenic *Pseudomonas aeruginosa* from bovine subclinical mastitis in West Bengal, India. *Vet. World* 10, 738–742.
- Bangar, Y.C., Singh, B., Dohare, A.K., Verma, M.R., 2015. A systematic review and meta-analysis of prevalence of sub-clinical mastitis in dairy cows in India. *Trop. Anim. Health Prod.* 47, 291–297.
- Bangar, Y.C., Verma, M.R., Dohare, A.K., Mukherjee, R., 2016. Meta-analysis of prevalence of clinical mastitis in crossbred cows in India (1995–2014). *J. Anim. Res.* 6, 933–938.
- Bansal, B.K., Gupta, D.K., 2009. Economic analysis of bovine mastitis in India and Punjab – a review. *Indian J. Dairy Sci.* 62, 337–345.
- Bansal, B.K., Gupta, D.K., Shafi, T.A., Sharma, S., 2015. Comparative antibiogram of coagulase-negative *Staphylococci* (CNS) associated with subclinical and clinical mastitis in dairy cows. *Vet. World* 8, 421–426.
- Bari, M.S., Alam, M., Uddin, M., Rahman, M.K., 2014. Prevalence and associated risk factors of bovine clinical mastitis in Patiya upazila under Chittagong district of Bangladesh. *Intl. J. Natural Sci.* 4, 5–9.

- Barua, M., Prodhan, M.A.M., Islam, K., Chowdhury, S., Hasanuzzaman, M., Imtiaz, M.A., Das, G.B., 2014. Sub-clinical mastitis prevalent in dairy cows in Chittagong district of Bangladesh: detection by different screening tests. *Vet. World* 7, 483–488.
- Bastan, A., Kaçar, C., Acar, D.B., Şahin, M., Cengiz, M., 2008. Investigation of the incidence and diagnosis of subclinical mastitis in early lactation period cows. *Turk. J. Vet. Anim. Sci.* 32, 119–121.
- Bedacha, B.D., Mengistu, H.T., 2011. Study on prevalence of mastitis and its associated risk factors in lactating dairy cows in Batu and its environs. *Ethiopia. Glob. Vet.* 7, 632–637.
- Beheshti, R., Eshratkhah, B., Shayegh, J., Ghalehkandi, J.G., Dianat, V., Valiei, K., 2011. Prevalence and etiology of subclinical mastitis in buffalo of the Tabriz region. *Iran. J. Am. Sci.* 7, 642–645.
- Belayneh, R., Belihu, K., Wubete, A., 2013. Dairy cows mastitis survey in Adama town. *Ethiopia. J. Vet. Med. Anim. Health* 5, 281–287.
- Beyene, B., Tolosa, T., 2017. Epidemiology and financial impact of bovine mastitis in an animal production and research center and smallholder dairy farms in Horo Guduru Wollega Zone, Western Ethiopia. *J. Dairy Vet. Anim. Res.* 5, 144–151.
- Bhanderi, B.M., Garg, M.R., 2012. A study on reducing the incidence of sub-clinical mastitis in dairy cows by feeding a vitamin and minerals based strategic feed supplement. *Indian J. Dairy Sci.* 65, 388–392.
- Bhanot, V., Chaudhri, S.S., Bisla, R.S., Singh, H., 2012. Retrospective study on prevalence and antibiogram of mastitis in cows and buffaloes of eastern Haryana. *Indian J. Anim. Res.* 46, 160–163.
- Bhat, A.M., Soodan, J.S., Tikoo, A., 2016. Study on prevalence of sub-clinical mastitis in crossbred dairy cattle and its potential risk factors. *J. Anim. Res.* 6, 747–749.
- Bhat, A.M., Soodan, J.S., Singh, R., Dobby, I.A., Hussain, T., Dar, M.Y., Mir, M., 2017. Incidence of bovine clinical mastitis in Jammu region and antibiogram of isolated pathogens. *Vet. World* 10, 984–989.
- Bhatt, V.D., Patel, M.S., Joshi, C.G., Kunjadia, A., 2011. Identification and antibiogram of microbes associated with bovine mastitis. *Anim. Biotechnol.* 22, 163–169.
- Bhikane, A.U., Awandkar, S.P., Hase, P.B., Syed, A.M., Ghoke, S.S., Awaz, K.B., 2010. Prevalence, etiology and antibiogram of subclinical mastitis in crossbred cows. *Vet. Pract.* 11, 122–123.
- Biffa, D., Debelia, E., Beyene, F., 2005. Prevalence and risk factors of mastitis in lactating dairy cows in Southern Ethiopia. *Intl. J. Appl. Res. Vet. Med.* 3, 189–198.
- Birhanu, M., Leta, S., Mamo, G., Tesfaye, S., 2017. Prevalence of bovine subclinical mastitis and isolation of its major causes in Bishoftu Town. *Ethiopia. BMC Res. Notes* 10, 767.
- Biswas, D., Sarker, T., 2017. Prevalence of sub-clinical mastitis at Banaripara upazilla. *Barisal. Bangladesh J. Vet. Med.* 15, 21–26.
- Bitew, M., Tafere, A., Tolosa, T., 2010. Study on bovine mastitis in dairy farms of Bahir Dar and its environs. *J. Anim. Vet. Adv.* 9, 2912–2917.
- Björk, S., 2013. Clinical and subclinical mastitis in dairy cattle in Kampala. In: Uganda, Degree Project in Veterinary Medicine. Swedish University of Agricultural Sciences, Uppsala.
- Bochniarz, M., Wawron, W., 2012. Haemolytic and proteolytic activity of coagulase-negative *Staphylococci* isolated from mastitis cows. *Pol. J. Vet. Sci.* 15, 61–65.
- Bochniarz, M., Wawron, W., Szczubial, M., 2013. Coagulase-negative *Staphylococci* (CNS) as an aetiological factor of mastitis in cows. *Pol. J. Vet. Sci.* 16, 487–492.
- Borne, B.H., Schaik, G., Lam, T.J., Nielen, M., 2010. Variation in herd level mastitis indicators between primi and multiparous in Dutch dairy herds. *Prev. Vet. Med.* 96, 49–55.
- Botrel, M.A., Haenni, M., Mornignat, E., Sulpice, P., Madec, J.Y., Calavas, D., 2010. Distribution and antimicrobial resistance of clinical and subclinical mastitis pathogens in dairy cows in Rhône-Alpes, France. *Foodborne Pathog. Dis.* 7, 479–487.
- Bradley, A.J., Green, M.J., 2001. Aetiology of clinical mastitis in six Somerset dairy herds. *Vet. Rec.* 148, 683–686.
- Bradley, A.J., Green, M.J., 2009. Factors affecting cure when treating bovine clinical mastitis with cephalosporin-based intramammary preparations. *J. Dairy Sci.* 92, 1941–1953.
- Brunner, N., Groeger, S., Raposo, J., Bruckmaier, R.M., Gross, J.J., 2019. Prevalence of subclinical ketosis and production diseases in dairy cows in Central and South America, Africa, Asia, Australia and New Zealand and Eastern Europe. *Transl. Anim. Sci.* 3, 84–92.
- Bulla, T.R., Rana, Y.S., Sharma, A., Beniwal, B.S., 2006. Prevalence of subclinical mastitis in Murrah buffaloes. *Haryana Vet.* 45, 53–56.
- Ceniti, C., Britti, D., Santoro, A.M.L., Musarella, R., Ciambrone, L., Casalinuovo, F., Costanzo, N., 2017. Phenotypic antimicrobial resistance profile of isolates causing clinical mastitis in dairy animals. *Ital. J. Food Saf.* 6, 84–87.
- Chahar, A., Gahlot, A.K., Tanwar, R.K., 2005. Prevalence of sub-clinical mastitis in cattle in Bikaner City. *Vet. Pract.* 6, 84–87.
- Charaya, G., Sharma, A., Singh, M., Tiwari, S., Pankaj, K.A., 2013. Subclinical mastitis at an organized farm: prevalence, etiology and antibiogram. *Haryana Vet.* 52, 30–32.
- Charaya, G., Sharma, A., Kumar, A., Singh, M., Goel, P., 2014. Pathogens isolated from clinical mastitis in Murrah buffaloes and their antibiogram. *Vet. World* 7, 980–985.
- Chishty, M.A., Arshad, M., Avisi, M., Ijaz, M., 2007. Cross-sectional epidemiological studies on mastitis in cattle and buffaloes of Tehsil Gojra. *Pakistan. Buff. Bull.* 26, 50–55.
- Chougule, M.N., Gadge, R.S., Bannalikar, A.S., Majee, S.B., Pharande, R.R., 2014. Characterization of *Staphylococcus aureus* associated with bovine mastitis. *Indian J. Dairy Sci.* 67, 493–497.
- Costelloe, S., 2004. Consultant guide to economics of mastitis. Available at: www.smartstock-usa.com. Accessed on 10th June 2020.
- Coulon, J.B., Gasqui, P., Barnouin, J., Ollier, A., Pradel, P., Pomiès, D., 2002. Effect of mastitis and related-germ on milk yield and composition during naturally-occurring udder infections in dairy cows. *Anim. Res.* 51, 383–393.
- Cynthia, M.K., 2005. The Merck Veterinary Manual. In: Whitehouse station. New Jersey, USA, Merck and Company Inc.
- Daniel, R.C.W., O'boyle, D., Marek, M.S., Frost, A.J., 1982. A survey of clinical mastitis in south-east Queensland dairy herds. *Aust. Vet. J.* 58, 143–147.
- Danish, Z., Bhakat, M., Paray, A.R., Ahmad, S., Lone, A.R., Mohanty, T.K., Sinha, R., 2018. Udder and teat morphology and their relation with incidence of sub-clinical and clinical mastitis in Sahiwal, Karan Fries cows and Murrah buffaloes. *J. Entomol. Zool. Stud.* 6, 2138–2141.
- Dar, K.H., Ansari, M.M., Dar, S.H., Tantary, H.A., Baba, M.A., Mehraj-ud-Din, N., 2014. Studies on subclinical mastitis in dairy cows of Jammu and Kashmir. *Intl. J. Vet. Sci.* 3, 95–99.
- Das, G., Lalnunpuia, C., Sarma, K., Behera, S.K., Dutta, T.K., Bandyopadhyay, S., 2015. Prevalence of *Staphylococcus aureus* associated sub-clinical mastitis in cross bred cows in Mizoram. *Ruminant Sci.* 4, 167–170.
- Dasohari, A., Somasani, A., Nagaraj, P., Gopala, R.A., 2017. Epidemiological studies of subclinical mastitis in cows in and around Hyderabad. *Pharma Innov. J.* 6, 975–979.
- De, U.K., Reena, M., 2009. Prevalence of mastitis in crossbred cows. *Indian Vet. J.* 86, 858–859.
- Dego, O.K., Tareke, F., 2003. Bovine mastitis in selected areas of southern Ethiopia. *Trop. Anim. Health Prod.* 35, 197–205.
- Deka, D., Goswami, R., Motina, E., Ralte, L., 2013. Antibiogram of microbes associated with bovine mastitis in Aizawl, Mizoram. *Indian Vet. J.* 90, 24–26.
- Delelesse, G.D., 2010. Study on prevalence of bovine mastitis on cross breed dairy cow around Holeta areas, West Shewa zone of Oromia. *Ethiopia. Glob. Vet.* 5, 318–323.
- Demissie, T.F., Mengistu, H.T., Mitiku, M.A., 2018. Prevalence of mastitis and identification of its bacterial causative agents in small holder dairy farms in and around Wukro of Tigray region. *Ethiopia. Intl. J. Adv. Res. Biol. Sci.* 5, 10–22.
- Devgnia, B.S., Khordia, D., Chodvadiya, M.B., Patel, R., Patel, D., Kinhekar, A.S., Singh, P.K., Kumar, V., Bhojne, G.R., Ravikumar, R.K., Kumar, V., 2015. Reverence of community towards grassroots livestock innovation: Responding to stakeholders need against sub-clinical mastitis in Amreli District, Gujarat, India. *Adv. Anim. Vet. Sci.* 3, 689–693.
- Dhakal, I.P., 2006. Normal somatic cell count and subclinical mastitis in Murrah buffaloes. *J. Vet. Med. Series B* 53, 81–86.
- Dhakal, I.P., Dhakal, P., Koshihara, T., Nagahata, H., 2007. Epidemiological and bacteriological survey of buffalo mastitis in Nepal. *J. Vet. Med. Sci.* 69, 1241–1245.
- Dieser, S.A., Vissio, C., Lasagno, M.C., Bogni, C.I., Larriestra, A.J., Odierno, L.M., 2014. Prevalence of pathogens causing subclinical mastitis in Argentinean dairy herds. *Pak. Vet. J.* 34, 124–126.
- Dubal, Z.B., Rahman, H., Papri, P., Kumar, A., Kalpana, P., 2010. Characterization and antimicrobial sensitivity of the pathogens isolated from bovine mastitis with special reference to *Escherichia coli* and *Staphylococcus* spp. *Indian J. Anim. Sci.* 80, 1163–1167.
- Duguma, A., Tolosa, T., Yohannes, A., 2014. Prevalence of clinical and sub-clinical mastitis on cross bred dairy cows at Holleta Agricultural Research Center, Central Ethiopia. *J. Vet. Med. Anim. Health* 6, 13–17.
- Edwards, S., Nicholls, M.J., Vallejos, F., Ibata, G., 1982. Mastitis survey in Bolivia. *Trop. Anim. Health Prod.* 14, 93–97.
- Elango, A., Doraisamy, K.A., Rajarajan, G., Kumaresan, G., 2010. Bacteriology of sub clinical mastitis and antibiogram of isolates recovered from cross bred cows. *Indian J. Anim. Res.* 44, 280–284.
- El-Ashker, M., Gwida, M., Tomaso, H., Monecke, S., Ehricht, R., El-Gohary, F., Hotzel, H., 2015. *Staphylococci* in cattle and buffaloes with mastitis in Dakahlia Governorate, Egypt. *J. Dairy Sci.* 98, 7450–7459.
- Elbably, M.A., Emeash, H.H., Asmaa, N.M., 2013. Risk factors associated with mastitis occurrence in dairy herds in Benisuef, Egypt. *World Vet. J.* 3, 5–10.
- Elemo, K.K., Sisay, T., Shiferaw, A., Fato, M.A., 2017. Prevalence, risk factors and multidrug resistance profile of *Staphylococcus aureus* isolated from bovine mastitis in selected dairy farms in and around Asella town, Arsi Zone, South Eastern Ethiopia. *Afr. J. Microbiol. Res.* 11, 1632–1642.
- Elhaig, M.M., Selim, A., 2014. Molecular and bacteriological investigation of subclinical mastitis caused by *Staphylococcus aureus* and *Streptococcus agalactiae* in domestic bovids from Ismailia, Egypt. *Trop. Anim. Health Pro.* 47, 271–276.
- El-Jakee, J.K., Aref, N.E., Gomaa, A., El-Hariri, M.D., Galal, H.M., Omar, S.A., Samir, A., 2013. Emerging of coagulase negative staphylococci as a cause of mastitis in dairy animals: An environmental hazard. *Intl. J. Vet. Sci. Med.* 1, 74–78.
- Elsayed, M.S., Mahmoud, A.E., El-Bagoury, M., Dawoud, A., 2015. Phenotypic and genotypic detection of virulence factors of *Staphylococcus aureus* isolated from clinical and subclinical mastitis in cattle and water buffaloes from different farms of Sadat City in Egypt. *Vet. World* 8, 1051–1058.
- Eriksson, L., 2013. Prevalence of subclinical mastitis and udder pathogens in small holder dairy farms in Mapepe, Batoka and Choma areas in Zambia. Swedish University of Agricultural Sciences, Uppsala, Degree Project in Veterinary Medicine.
- Erskine, R.J., Bartlett, P.C., VanLente, J.L., Phipps, C.R., 2002. Efficacy of systemic ceftiofur as a therapy for severe clinical mastitis in dairy cattle. *J. Dairy Sci.* 85, 2571–2575.
- FAO, 2014. Impact of mastitis in small scale dairy production system. In: Working paper No.13. Rome.
- FAOSTAT, 2020. Population of Live animals in the World - Visualization tool. <http://www.fao.org/faostat/en/#data/QA/visualize>. Accessed online on 22nd June 2020.
- Faroog, A.A., Inayat, S., Akhtar, M.S., Mushtaq, M., 2008. Prevalence of mastitis and antibiotic sensitivity of bacterial isolates recovered from Nili-Ravi buffaloes. *J. Anim. Plant Sci.* 18, 76–77.
- Lakew, B.T., Fayera, T., Ali, Y.M., 2019. Risk factors for bovine mastitis with the isolation and identification of *Streptococcus agalactiae* from farms in and around Haramaya district, Eastern Ethiopia. *Trop. Anim. Health Pro.* 51, 1507–1513.

- Freitas, D.M.F., Luz, I.D.S., Silveira-Filho, V.D.M., Júnior, J., Stamford, T.L., Mota, R.A., Sena, M.J.D., de Almeida, A.M., Balbino, V.D.Q., Leal-Balbino, T.C., 2008. *Staphylococcal* toxin genes in strains isolated from cows with subclinical mastitis. *Pesqui. Vet. Bras.* 28, 617–621.
- Fthenakis, G.G., 1998. Susceptibility to antibiotics of staphylococcal isolates from cases of ovine or bovine mastitis in Greece. *Small Rumin. Res.* 28, 9–13.
- Gao, J., Barkema, H.W., Zhang, L., Liu, G., Deng, Z., Cai, L., Shan, R., Zhang, S., Zou, J., Kastelic, J.P., Han, B., 2017. Incidence of clinical mastitis and distribution of pathogens on large Chinese dairy farms. *J. Dairy Sci.* 100, 4797–4806.
- Getahun, K., Kelay, B., Bekana, M., Lobago, F., 2008. Bovine mastitis and antibiotic resistance patterns in Selale smallholder dairy farms, central Ethiopia. *Trop. Anim. Health Prod.* 40, 261–268.
- Giannechini, R., Concha, C., Rivero, R., Delucci, I., López, J.M., 2002. Occurrence of clinical and sub-clinical mastitis in dairy herds in the West Littoral Region in Uruguay. *Acta Vet. Scand.* 43, 221–230.
- Girma, S., Mammo, A., Bogele, K., Sori, T., Tadesse, F., Jibat, T., 2012. Study on prevalence of bovine mastitis and its major causative agents in West Harerghe zone, Doba district. Ethiopia. *J. Vet. Med. Anim. Health* 4, 116–123.
- Gogoi, S.M., Tamuly, U., Khuman, L.S., 2017. Prevalence of sub-clinical mastitis in areas around Lakhimpur town of Assam. *Intl. J. Agri. Sci. Res.* 7, 501–508.
- Gómez-Quispe, O.E., Santiván-Ballón, C.S., Arauco-Villar, F., Espozza-Flores, O.H., Manrique-Meza, J., 2015. Interpretation criteria for California Mastitis Test in the diagnosis of subclinical mastitis in cattle. *Revista de Investigaciones Veterinarias del Perú* 26, 86–95.
- Gonzalez, R.N., Giraudo, J.A., Busso, J.J., 1980. Studies on subclinical mastitis. II. Bacterial agents. *Revista de Medicina Veterinaria, Argentina* 61, 225–234.
- Gonzalez, R.N., Cullor, J.S., Jasper, D.E., Farver, T.B., Bushnell, R.B., Oliver, M.N., 1989. Prevention of clinical coliform mastitis in dairy cows by a mutant *Escherichia coli* vaccine. *Can. J. Vet. Res.* 53, 301.
- Gonzalez, R.N., Jasper, D.E., Kronlund, N.C., Farver, T.B., Cullor, J.S., Bushnell, R.B., Dellinger, J.D., 1990. Clinical mastitis in two California dairy herds participating in contagious mastitis control programs. *J. Dairy Sci.* 73, 648–660.
- Graaf, D.T., Dwinger, R.H., 1996. Estimation of milk production losses due to sub-clinical mastitis in dairy cattle in Costa Rica. *Prev. Vet. Med.* 26, 215–222.
- Green, M.J., Green, L.E., Medley, G.F., Schukken, Y.H., Bradley, A.J., 2002. Influence of dry period bacterial intra mammary infection on clinical mastitis in dairy cows. *J. Dairy Sci.* 85, 2589–2599.
- Grönlund, U., Sandgren, C.H., Waller, K.P., 2005. Haptoglobin and serum amyloid A in milk from dairy cows with chronic sub-clinical mastitis. *Vet. Res.* 36, 191–198.
- Guha, A., Gera, S., 2011. Eti-prevalence of sub clinical mastitis in Holstein x Haryana crossbred cattle. *Exploratory Anim. Med. Res.* 1, 75–78.
- Guha, A., Gera, S., Sharma, A., 2010. Assessment of chemical and electrolyte profile as an indicator of subclinical mastitis in riverine buffalo (*Bubalus bubalis*). *Haryana Vet.* 49, 19–21.
- Guha, A., Guha, R., Gera, S., 2012. Comparison of somatic cell count, California mastitis test, chloride test and rennet coagulation time with bacterial culture examination to detect subclinical mastitis in riverine buffalo (*Bubalus bubalis*). *Afr. J. Agric. Res.* 7, 5578–5584.
- Haftu, R., Taddele, H., Gugsa, G., Kalayou, S., 2012. Prevalence, bacterial causes, and antimicrobial susceptibility profile of mastitis isolates from cows in large-scale dairy farms of Northern Ethiopia. *Trop. Anim. Health Prod.* 44, 1765–1771.
- Haghkhah, M., Nazifi, S., Jahromi, A.G., 2010. Evaluation of milk haptoglobin and amyloid A in high producing dairy cattle with clinical and subclinical mastitis in Shiraz. *Comp. Clin. Pathol.* 19, 547–552.
- Haghkhah, M., Ahmadi, M.R., Gheisari, H.R., Kadivar, A., 2011. Preliminary bacterial study on subclinical mastitis and teat condition in dairy herds around Shiraz. *Turk. J. Vet. Anim. Sci.* 35, 387–394.
- Hagnestam, C., Emanuelson, U., Berglund, B., 2007. Yield losses associated with clinical mastitis occurring in different weeks of lactation. *J. Dairy Sci.* 90, 2260–2270.
- Haidich, A.B., 2010. Meta-analysis in medical research. *Hippokratia* 14, 29–37.
- Hailemeskel, D., Admasu, P., Alemu, F., 2014. Prevalence and identification of bacterial pathogens causing bovine mastitis from crossbred of dairy cows in North Showa Zone of Ethiopia. *Glob. Vet. Clin.* 13, 189–195.
- Hamed, M.I., Ziatoun, A.M.A., 2014. Prevalence of *Staphylococcus aureus* subclinical mastitis in dairy buffaloes farms. *Intl. J. Livest. Res.* 4, 21–28.
- Hameed, K.G.A., Sender, G., Korwin-Kossakowska, A., 2006. Public health hazard due to mastitis in dairy cows. *Anim. Sci. Paper Rep.* 25, 73–85.
- Hameed, S., Arshad, M., Ashraf, M., Avais, M., Shahid, M.A., 2012. Cross-sectional epidemiological studies on mastitis in cattle and buffaloes of Tehsil Burewala. *Pakistan. J. Anim. Plant Sci.* 22, 371–376.
- Hamid, I.M.B., Shuiq, E.S., El-Zubeir, I.E.M., Saad, A.Z., El Owni, O.A.O., 2012. Influence of *Staphylococcus aureus* mastitis on milk composition of different dairy breeds of cattle in Khartoum State. Sudan. *World Vet. J.* 2, 13–16.
- Hardenberg, F., 2016. Clinical and subclinical mastitis in dairy cattle and buffaloes in Bihar, India Prevalence, major pathogens and risk factors. Degree Project in Veterinary Medicine. Swedish University of Agricultural Sciences, Uppsala.
- Harini, H., Sumathi, B.R., 2011. Screening of bovine milk samples for sub-clinical mastitis and antibiogram of bacterial isolates. *Vet. World* 4, 358–359.
- Hashemi, M., Kafi, M., Safdarian, M., 2011. The prevalence of clinical and subclinical mastitis in dairy cows in the central region of Fars province, south of Iran. *Iran J. Vet. Res.* 12, 236–241.
- Hawari, A.D., Al-Dabbas, F., 2008. Prevalence and distribution of mastitis pathogens and their resistance against antimicrobial agents in dairy cows in Jordan. *Am. J. Anim. Vet. Sci.* 3, 36–39.
- Hegde, R., Isloor, S., Prabhu, K.N., Shome, B.R., Rathnamma, D., Suryanarayana, V.V.S., Yatiraj, S., Prasad, C.R., Krishnaveni, N., Sundareshan, S., Akhila, D.S., 2013. Incidence of subclinical mastitis and prevalence of major mastitis pathogens in organized farms and unorganized sectors. *Indian J. Microbiol.* 53, 315–320.
- Higgins, J.P.T., Lane, P.W., Anagnostelis, B., Anzures-Cabrera, J., Baker, N.F., Cappelleri, J.C., Haugie, S., Hollis, S., Lewis, S.C., Moneuse, P., Whitehead, A., 2013. A tool to assess the quality of a meta-analysis. *Res. Synth. Methods* 4, 351–366.
- Hogeveen, H., Huijps, K., Lam, T.J.G.M., 2011. Economic aspects of mastitis: New developments. *New Zealand Vet. J.* 59, 16–23.
- Hoque, M.N., Das, Z.C., Talukder, A.K., Alam, M.S., Rahman, A.N.M.A., 2014. Different screening tests and milk somatic cell count for the prevalence of subclinical bovine mastitis in Bangladesh. *Trop. Anim. Health Prod.* 47, 79–86.
- Hoque, M.N., Das, Z.C., Rahman, A.N.M.A., Haider, M.G., Islam, M.A., 2018. Molecular characterization of *Staphylococcus aureus* strains in bovine mastitis milk in Bangladesh. *Intl. J. Vet. Sci. Med.* 6, 53–60.
- Hossain, S., Reza, M.A., Hasan, M.N., Sorwar, M.G., Billah, M., 2016. Impact of clinical mastitis in dairy farming at Keshabpur Upazilla in Jessore in Bangladesh. *Bangladesh J. Vet. Med.* 14, 59–64.
- Hussain, A., Shakoor, A., Shahid, M.A., Numam, M., Gulraiz, F., 2007. Clinical and subclinical *Staphylococcus aureus* mastitis in dairy buffaloes: disease characteristics and antibiotic susceptibility profiles of isolates. *Intl. J. Agri. Res.* 2, 804–811.
- Hussain, A., Ahmed, M.D., Mushtaq, M.H., Khan, M.S., Khan, M.A., Nisar, M., Sabir, N., Khan, S.A., 2013. Antibiogram analysis of *Staphylococcus aureus* isolated from mastitic milk samples of buffaloes in district Bhimber Azad Kashmir. *Buffalo Bull.* 32, 1021–1028.
- Hussain, M., Naeem, K., Iqbal, N., 1984. Subclinical mastitis in cows and buffaloes: Identification and drug susceptibility of causative organism. *Pak. Vet. J.* 4, 161–164.
- Hussein, S.A., 2012. Prevalence and bacterial etiology of subclinical mastitis in dairy cows in Al Sulaimaniyah district. *Kufa J. Vet. Med. Sci.* 3, 190–203.
- Idriss, S.E., Folty, V., Tančin, V., Kirchnerová, K., Zaujec, K., 2013. Mastitis pathogens in milk of dairy cows in Slovakia. *Slovak J. Anim. Sci.* 46, 115–119.
- Iraguha, B., Hamudikuwanda, H., Mushonga, B., 2015. Bovine mastitis prevalence and associated risk factors in dairy cows in Nyagatare District. Rwanda. *J. South Afr. Vet. Assoc.* 86, 1–6.
- Islam, M.A., Rahman, A.K.M.A., Rony, S.A., Islam, M.S., 2010. Prevalence and risk factors of mastitis in lactating dairy cows at Baghabari milk shed area of Sirajganj. *Bangladesh J. Vet. Med.* 8, 157–162.
- Islam, M.A., Islam, M.Z., Rahman, M.S., Islam, M.T., 2011. Prevalence of subclinical mastitis in dairy cows in selected areas of Bangladesh. *Bangladesh J. Vet. Med.* 9, 73–78.
- Islam, N.N., Farzana, Z., Chowdhury, A.M.M.A., Mannan, A., Kamaruddin, K.M., Siddiki, A.M.A.M.Z., Uddin, I., 2014. Characterization of bovine subclinical mastitis caused by *Staphylococcus aureus* in southern Bangladesh by bacteriological and molecular approaches. *Asian J. Biol. Sci.* 7, 1–12.
- Jagadeesh, D.S., Puttamallappa, R.K., Keregallolkoppal, H.G., Lakshminarasimhiah, M., 2016. Prevalence of sub-clinical mastitis in cattle and effect on milk quality. *Adv. Anim. Vet. Sci.* 4, 237–240.
- Jain, A.K., Goel, P., Sharma, A., Kumar, A., 2009. Comparison of different tests for diagnosis of bovine sub-clinical mastitis. *J. Immunol. Immunopathol.* 11, 53–56.
- Jamali, H., Radmehr, B., Ismail, S., 2014. Prevalence and antibiotic resistance of *Staphylococcus aureus* isolated from bovine clinical mastitis. *J. Dairy Sci.* 97, 2226–2230.
- Jarassaeng, C., Aiumlamai, S., Wachirapakorn, C., Techakumphu, M., Noordhuizen, J.P., Beynen, A.C., Suadsong, S., 2012. Risk factors of subclinical mastitis in small holder dairy cows in Khon Kaen province. *Thai. J. Vet. Med.* 42, 43–151.
- Jena, B., Pagrut, N.K., Sahoo, A., Ahmed, A., 2015. Subclinical bovine mastitis in rural, peri-urban and suburban regions of Jaipur district of Rajasthan. India. *J. Anim. Res.* 5, 175–182.
- Jingar, S.C., Singh, M., Lawania, P., Roy, A.K., Kumar, A., 2017. Effect of year wise on incidence of mastitis in crossbred, indigenous cattle and Murrah buffaloes. *Int. J. Curr. Microbiol. App. Sci.* 6, 2725–2728.
- Joshi, S., Gokhale, S., 2006. Status of mastitis as an emerging disease in improved and peripubertal dairy farms in India. *Ann. N. Y. Acad. Sci.* 1081, 74–83.
- Joshi, H., Tanwar, R.K., Chahar, A., 2013. Prevalence of sub-clinical mastitis in buffaloes. *Vet. Pract.* 14, 377–378.
- Judge, L.J., Erskine, R.J., Bartlett, P.C., 1997. Recombinant bovine somatotropin and clinical mastitis: incidence, discarded milk following therapy, and culling. *J. Dairy Sci.* 80, 3212–3218.
- Junaidu, A.U., Salihu, M.D., Tambuwal, F.M., Magaji, A.A., Jaafaru, S., 2011. Prevalence of mastitis in lactating cows in some selected commercial dairy farms in Sokoto metropolis. *Adv. Appl. Sci. Res.* 2, 290–294.
- Kabir, M.H., Ershaduzzaman, M., Giasuddin, M., Islam, M.R., Nazir, K.N.H., Islam, M.S., Karim, M.R., Rahman, M.H., Ali, M.Y., 2017. Prevalence and identification of subclinical mastitis in cows at BLRI Regional Station, Sirajganj. Bangladesh. *J. Adv. Vet. Anim. Res.* 4, 295–300.
- Kalantri, A., Safi, S., Foroushani, A.R., 2013. Milk lactate dehydrogenase and alkaline phosphatase as biomarkers in detection of bovine subclinical mastitis. *Ann. Biol. Res.* 4, 302–307.
- Kaliwal, B.B., Sadashiv, S.O., Kurjogi, M.M., Sanakal, R.D., 2011. Prevalence and antimicrobial susceptibility of coagulase-negative staphylococci isolated from bovine mastitis. *Vet. World* 4, 158–161.
- Kalmus, P., Viltrop, A., Aasmäe, B., Kask, K., 2006. Occurrence of clinical mastitis in primiparous Estonian dairy cows in different housing conditions. *Acta Vet. Scand.* 48, 21.
- Karabasanavar, N.S., Singh, S.P., 2013. Isolation and antibiograms of coagulase negative *Staphylococci* from bovine mastitis milk. *J. Foodborne Zoonotic Dis.* 1, 21–23.

- Karahan, M., Çetinkaya, B., 2007. Coagulase gene polymorphisms detected by PCR in *Staphylococcus aureus* isolated from subclinical bovine mastitis in Turkey. *Vet. J.* 174, 428–431.
- Kashyap, D.K., Giri, D.K., Dewangan, G., 2019. Prevalence of sub clinical mastitis (SCM) in she buffaloes at Surajpur district of Chhattishgarh. *India. Buffalo Bull.* 38, 373–381.
- Kasozi, K.I., Tingira, J.B., Vudriko, P., 2014. High prevalence of subclinical mastitis and multidrug resistant *Staphylococcus aureus* are a threat to dairy cattle production in Kiboga District (Uganda). *Open J. Vet. Med.* 4, 35–43.
- Kathiriya, J.B., Kabaria, B.B., Saradava, D.A., Sanepara, D.P., 2014. Prevalence of subclinical mastitis in dairy cows in Rajkot district of Gujarat. *Intl. J. Sci. Nature* 5, 433–436.
- Katsande, S., Matope, G., Ndengu, M., Pfukenyi, D.M., 2013. Prevalence of mastitis in dairy cows from smallholder farms in Zimbabwe. *J. Vet. Res.* 80, 523.
- Kaur, A., Singh, S.G., Singh, V., 2015a. Seasonal prevalence and antibiogram profile of bacterial isolates from bovine mastitis. *J. Anim. Res.* 5, 623–629.
- Kaur, M., Verma, R., Bansal, B.K., Mukhopadhyay, C.S., Arora, J.S., 2015b. Status of sub-clinical mastitis and associated risk factors in Indian water buffalo in Doaba region of Punjab, India. *Indian J. Dairy Sci.* 68, 483–487.
- Kavitha, K.L., Rajesh, K., Stresh, K., Satheesh, K., Sundar, N.S., 2009. Buffalo mastitis-risk factors. *Buffalo Bull.* 28, 134–137.
- Kayesh, M.E.H., Talukder, M., Anower, A.K.M.M., 2014. Prevalence of subclinical mastitis and its association with bacteria and risk factors in lactating cows of Barisal district in Bangladesh. *Intl. J. Biol. Res.* 2, 35–38.
- Kebebew, G., Jorga, E., 2016. Prevalence and risk factors of bovine mastitis in Ambo town of West Shewa Zone, Oromia, Ethiopia. *Ethiop. Vet. J.* 20, 123–134.
- Kedir, J., Disassa, H., Jaleta, H., Zenebe, T., Kebede, G., 2016. A Study on bovine mastitis, isolation and identification of *Staphylococcus* species in Dairy Farms of Dire Dawa City, Eastern Ethiopia. *Glob. Vet.* 16, 222–230.
- Khan, A.Z., Muhammad, G., 2005. Quarter-wise comparative prevalence of mastitis in buffaloes and crossbred cows. *Pak. Vet. J.* 25, 9–12.
- Khan, A.Z., Khan, A., Hayat, C.S., Munir, Z., Ayaz, U., 2004. Prevalence of mastitis in buffaloes and antibiotics sensitivity profiles of isolates. *Pak. J. Life Sci. Sci.* 2, 73–75.
- Khan, A., Mushtaq, M.H., Ahmad, M.U.D., Chaudhry, M., Khan, A.W., 2015. Prevalence of clinical mastitis in bovines in different climatic conditions in KPK, (Pakistan). *Sci. Intl. (Lahore)* 27, 2289–2293.
- Khanal, T., Pandit, A., 2013. Assessment of sub-clinical mastitis and its associated risk factors in dairy livestock of Lamjung, Nepal. *Intl. J. Infect. Microbiol.* 2, 49–54.
- Kivaria, F.M., Noordhuizen, J.P.T.M., Kapaga, A.M., 2004. Risk indicators associated with subclinical mastitis in smallholder dairy cows in Tanzania. *Trop. Anim. Health Prod.* 36, 581–592.
- Koricheva, J., Gurevitch, J., Mengersen, K., 2013. Handbook of meta-analysis in ecology and evolution. Princeton University Press, New Jersey, USA.
- Kotb, E.E., Abu-Seida, A.M., Fadel, M.S., 2014. The correlation between ultrasonographic and laboratory findings of mastitis in buffaloes (*Bubalus bubalis*). *Glob. Vet.* 13, 68–74.
- Krishnamoorthy, P., Suresh, K.P., Saha, S., Govindaraj, G., Shome, B.R., Roy, P., 2017. Meta-analysis of prevalence of subclinical and clinical mastitis, major mastitis pathogens in dairy cattle in India. *Int. J. Curr. Microbiol. App. Sci.* 6, 1214–1234.
- Krishnamoorthy, P., Ashwini, M., Suresh, K.P., Siju, S.J., Roy, P., 2019a. Prevalence of *Anaplasma* species in India and the World in dairy animals: A systematic review and meta-analysis. *Res. Vet. Sci.* 123, 159–170.
- Krishnamoorthy, P., Hamsapriya, S., Ashwini, M., Patil, S.S., Roy, P., Suresh, K.P., 2019b. Systematic review and meta-analysis of livestock associated-methicillin resistant *Staphylococcus aureus* (LA-MRSA) prevalence in animals in India. *Intl. J. Livest. Res.* 9, 179–191.
- Krishnamoorthy, P., Suresh, K.P., Roy, P., 2020. Meta-analysis: An innovative tool for estimating prevalence of livestock diseases. *Res. Rev. J. Vet. Sci. Technol.* 9, 4–7.
- Krishnaveni, N., Isloor, S.K., Hegde, R., Suryanarayanan, V.V.S., Rathnma, D., Veeregowda, B.M., Nagaraja, C.S., Sundaresan, S., 2014. Rapid detection of virulence associated genes in Streptococcal isolates from bovine mastitis. *Afr. J. Microbiol. Res.* 8, 2245–2254.
- Kumar, R., Sharma, A., 2002. Prevalence, etiology and antibiogram of mastitis in cows and buffaloes in Hisar, Haryana. *Indian J. Anim. Sci.* 72, 361–363.
- Kumar, M., Kumar, R., Sharma, A., Jain, V.K., 2007. Investigations on prevalence and oxidative stress aspects of mastitis in buffaloes. *Ital. J. Anim. Sci.* 6, 978–979.
- Kurjogi, M.M., Kaliwal, B.B., 2011. Prevalence and antimicrobial susceptibility of bacteria isolated from bovine mastitis. *Adv. Appl. Sci. Res.* 2, 229–235.
- Kurjogi, M.M., Kaliwal, B.B., 2014. Epidemiology of bovine mastitis in cows of Dharwad district. *Intl. Scholarly Res. Notices* 2014, 1–9.
- Lago, A., Godden, S.M., Bey, R., Ruegg, P.L., Leslie, K., 2011. The selective treatment of clinical mastitis based on on-farm culture results: I. Effects on antibiotic use, milk withholding time, and short-term clinical and bacteriological outcomes. *J. Dairy Sci.* 94, 4441–4456.
- Lakew, M., Tolosa, T., Tigre, W., 2009. Prevalence and major bacterial causes of bovine mastitis in Asella, South Eastern Ethiopia. *Trop. Anim. Health Pro.* 41, 1525–1530.
- Lakshmi, R., Jayavardhanan, K., 2016. Isolation and identification of major causing bacteria from bovine mastitis. *Intl. J. Appl. Pure Sci. Agri.* 2, 45–48.
- Lamey, A.E., Ammar, A.M., Zaki, E.R., Khairy, N., Moshref, B.S., Refai, M.K., 2013. Virulence factors of *Escherichia coli* isolated from recurrent cases of clinical and subclinical mastitis in buffaloes. *Intl. J. Microbiol. Res.* 4, 86–94.
- Langer, A., Sharma, S., Sharma, N.K., Nauriyal, D.S., 2014. Comparative efficacy of different mastitis markers for diagnosis of sub-clinical mastitis in cows. *Intl. J. Appl. Sci. Biotech.* 2, 121–125.
- León-Galván, M., Corona, B.J.E., Arana, L.A.A., Posadas, V.M., Aguayo, D.D., Pelaez, C., Ortega, M.E.A., Chavez, G.A.J., 2015. Molecular detection and sensitivity to antibiotics and bacteriocins of pathogens isolated from bovine mastitis in family dairy herds of central Mexico. *Bio. Med. Res. Int.* 2015, 9.
- Li, J.P., Zhou, H.J., Yuan, L., He, T., Hu, S.H., 2009. Prevalence, genetic diversity, and antimicrobial susceptibility profiles of *Staphylococcus aureus* isolated from bovine mastitis in Zhejiang Province, China. *J. Zhejiang Univ. Sci. B* 10, 753–760.
- Linton, A.H., Robinson, T.C., 1984. Studies on the association of *Escherichia coli* with bovine mastitis. *Br. Vet. J.* 140, 368–373.
- López, A.M.M., Zarco, S.D., García, F.S., Carranza, B.V., Castillo, A.D.C.G., Pliego, A.B., Rojas, M.T., Fresán, M.U.A., Ordóñez, V.V., 2012. *Staphylococcus aureus* biotypes in cows presenting subclinical mastitis from family dairy herds in the Central-Eastern State of Mexico. *Revista Mexicana De Ciencias Pecuarias* 3, 265–274.
- Lucia, M., Rahayu, S., Haerah, D., Wahyuni, D., 2017. Detection of *Staphylococcus aureus* and *Streptococcus agalactiae*: subclinical mastitis causes in dairy cow and dairy buffalo (*Bubalus bubalis*). *Am. J. Biomed. Res.* 5, 8–13.
- Maćesić, N., Bacić, G., Božicević, K., Benić, M., Karadjole, T., Babic, N.P., Lojkic, M., Efendic, M., Bacic, I., Pavlak, M., 2016. Assessment of the Zagreb mastitis test in diagnosis of subclinical mastitis in dairy cattle. *Vet. Arh.* 86, 475–485.
- Madalcho, E.B., 2019. A study on the prevalence of bovine mastitis and isolation of major pathogens associated with it in and around Wolaita Sodo, Southern Ethiopia. *Intl. J. Res. Stud. Biosci.* 7, 40–48.
- Mahajan, S., Bhatt, P., Ramakant, K.A., Dabas, Y.P.S., 2011. Risk and occurrence of bovine mastitis in Tarai region of Uttarakhand. *Vet. Pract.* 12, 244–250.
- Maichomo, M.W., Ndirangu, P.N., Mungube, E.O., Nyongesa, P.K., Wamae, D., On'gala, J.O., Gicheru, N.W., Keya, G., Wesonga, H.O., Siamba, D., 2019. A novel pH-based pen-side test for detection of sub-clinical mastitis: validation in cattle and camels. *Kenya. Livest. Res. Rural Dev.* 31, 7.
- Malinowski, E., Kłosowska, A., Kaczmarowski, M., Kuzma, K., 2003. Prevalence of intra mammary infections in pregnant heifers. *Bull. Vet. Inst. Pulawy* 47, 165–170.
- Manasa, V., Saikumar, T.V., Rao, T.P., Kumar, K.A., 2019. Incidence of clinical and sub-clinical bovine mastitis caused by *Staphylococcus aureus* in Proddatur region of Andhra Pradesh. *Intl. J. Chem. Stud.* 7, 788–792.
- Marama, A., Mamu, G., Birhanu, T., 2016. Prevalence and antibiotic resistance of *Staphylococcus aureus* mastitis in Holeta area, Western Ethiopia. *Glob. Vet.* 16, 365–370.
- Marashifard, M., Aliabad, Z.K., Hosseini, S.A.A.M., Sarokhalil, D.D., Mirzaei, M., Khoramrooz, S.S., 2019. Determination of antibiotic resistance pattern and virulence genes in *Escherichia coli* isolated from bovine with subclinical mastitis in southwest of Iran. *Trop. Anim. Health Prod.* 51, 575–580.
- Marija, P., Stanko, B., Branko, V., Zoran, R., Vera, K., Miodrag, R., Aleksandra, N., Dušan, S., Milijana, B., 2016. Prevalence and molecular characterization of enterotoxin-producing strains of *Staphylococcus aureus* isolated from Serbian dairy cows. *Acta Vet.* 66, 466–477.
- Mdegela, R.H., Karimuribo, E., Kusiluka, L.J.M., Kabula, B., Manjurano, A., Kapaga, A.M., Kambarage, D.M., 2005. Mastitis in smallholder dairy and pastoral cattle herds in the urban and peri-urban areas of the Dodoma municipality in Central Tanzania. *Livest. Res. Rural Dev.* 17, 123.
- Mdegela, R.H., Ryoba, R., Karimuribo, E.D., Phiri, E.J., Løken, T., Reksen, O., Mtengeneti, E., Uriø, N.A., 2009. Prevalence of clinical and subclinical mastitis and quality of milk on smallholder dairy farms in Tanzania. *J. South Afr. Vet. Assoc.* 80, 163–168.
- Megersa, B., Manedo, A., Regassa, M.A.A., Abunna, F., 2012. Mastitis in lactating cows at Hawassa Town: prevalence, risk factors, major bacterial causes and treatment response to routinely used antibiotics. *American-Eurasian J. Sci. Res.* 7, 86–91.
- Meher, M.M., Hasan, A., Afrin, M., 2018. Field investigation on sub-clinical mastitis in cows in different areas of Barisal district in Bangladesh. *Turk. J. Agri. Food Sci. Technol.* 6, 1159–1162.
- Mekibib, B., Furgasa, M., Abunna, F., Megersa, B., Regassa, A., 2010. Bovine mastitis: Prevalence, risk factors and major pathogens in dairy farms of Holeta town, Central Ethiopia. *Vet. Clin. 3, 397–403.*
- Mekonnen, H., Tesfaye, A., 2010. Prevalence and etiology of mastitis and related management factors in market oriented smallholder dairy farms in Adama, Ethiopia. *Rev. Med. Vet.* 161, 574–579.
- Mekonnen, S.A., Koop, G., Melkie, S.T., Getahun, C.D., Hogeveen, H., Lam, T.J., 2017. Prevalence of subclinical mastitis and associated risk factors at cow and herd level in dairy farms in North-West Ethiopia. *Prev. Vet. Med.* 145, 23–31.
- Memon, J., Kashif, J., Yaqoob, M., Liping, W., Yang, Y., Hongjie, F., 2013. Molecular characterization and antimicrobial sensitivity of pathogens from sub-clinical and clinical mastitis in Eastern China. *Pak. Vet. J.* 33, 170–174.
- Michael, L.G., Benti, D., Feyissa, B., Abebe, M., 2013. Study on prevalence of bovine mastitis in lactating cows and associated risk factors in and around Areka town, Southern of Ethiopia. *Afr. J. Microbiol. Res.* 7, 5051–5056.
- Miltenburg, J.D., De Lange, D., Crauwels, A.P.P., Bongers, J.H., Tielen, M.J.M., Schukken, Y.H., Elbers, A.R.W., 1996. Incidence of clinical mastitis in a random sample of dairy herds in the southern Netherlands. *Vet. Rec.* 139, 204–207.
- Mir, A.Q., Bansal, B.K., Gupta, D.K., 2014. Subclinical mastitis in machine milked dairy farms in Punjab: prevalence, distribution of bacteria and current antibiogram. *Vet. World* 7, 291–294.
- Moges, N., Hailemariam, T., Fentahun, T., Chanie, M., Melaku, A., 2012. Bovine mastitis and associated risk factors in small holder lactating dairy farms in Hawassa, Southern Ethiopia. *Glob. Vet.* 9, 41–446.
- Moher, D., Shamseer, L., Clarke, M., Ghersi, D., Liberati, A., Petticrew, M., Shekelle, P., Stewart, L.A., Group, P.P., 2015. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Syst. Rev.* 4, 1–9. <https://doi.org/10.1186/2046-4053-4-1>.
- Morse, D., DeLorenzo, M.A., Wilcox, C.J., Collier, R.J., Natzke, R.P., Bray, D.R., 1988. Climatic effects on occurrence of clinical mastitis. *J. Dairy Sci.* 71, 848–853.

- Mpatswenumugabo, J.P., Bebora, L.C., Gitao, G.C., Mbegi, V.A., Iraguha, B., Kamana, O., Shumbusho, B., 2017. Prevalence of subclinical mastitis and distribution of pathogens in dairy farms of Rubavu and Nyabihu districts. Rwanda. *J. Vet. Med.* 2017, 1–8.
- Mubarack, H.M., Doss, A., Vijayasanthi, M., Venkataswamy, R., 2012. Antimicrobial drug susceptibility of *Staphylococcus aureus* from subclinical bovine mastitis in Coimbatore, Tamilnadu. South India. *Vet. World* 5, 352–355.
- Muhammad, G., Naureen, A., Asi, M.N., Saqib, M., 2010. Evaluation of a 3% surf solution (surf field mastitis test) for the diagnosis of subclinical bovine and bubaline mastitis. *Trop. Anim. Health Prod.* 42, 457–464.
- Mulat, B., Abegaz, S., Nazir, S., 2017. Antibiogram of bacterial pathogens isolated from subclinical mastitis in Kombolcha, South Wollo. Ethiopia. *Trop. Anim. Health Pro.* 65, 37–47.
- Mulshet, Y., Derso, S., Nigus, A., 2017. Prevalence of bovine subclinical mastitis and associated risk factors in Addis Ababa. Central Ethiopia. *J. Anim. Feed Res.* 7, 124–133.
- Mureithi, D.K., Njuguna, M.N., 2016. Prevalence of subclinical mastitis and associated risk factors in dairy farms in urban and peri-urban areas of Thika Sub County. Kenya. *Livest. Res. Rural Dev.* 28, 13.
- Mureithi, D.K., Khang, K.C., Kamau, M.N., 2017. Antimicrobial resistance profile in bacterial isolates from subclinical mastitic milk samples in dairy herds in Kenya. *Bull. Anim. Health Pro. Afr.* 65, 167–173.
- Mustafa, Y.S., Awan, F.N., Zaman, T., Chaudhry, S.R., Zoyfro, V., 2011. Prevalence and antibacterial susceptibility in mastitis in buffalo and cow in and around the district Lahore. Pakistan. *Pak. J. Pharm. Sci.* 24, 29–33.
- Naghshineh, S., Rafat, S.A., Shoja, J., Moghaddam, G.A., Ebrahimi, M., 2015. Prevalence and risk factors of subclinical mastitis in Iranian Holstein cows. *Iran J. Appl. Anim. Sci.* 5, 569–574.
- Nam, H.M., Kim, J.M., Lim, S.K., Jang, K.C., Jung, S.C., 2010. Infectious aetiologies of mastitis on Korean dairy farms during 2008. *Res. Vet. Sci.* 88, 372–374.
- Ndahetuye, J.B., Persson, Y., Nyman, A.K., Tukey, M., Ongol, M.P., Båge, R., 2019. Aetiology and prevalence of subclinical mastitis in dairy herds in peri-urban areas of Kigali in Rwanda. *Trop. Anim. Health Prod.* 51, 2037–2044.
- Ndirangu, P.N., Siamba, D., Wesonga, H.O., Mungube, E.O., Maichomo, M.W., Mugambi, J.M., 2017. Prevalence of bovine mastitis and multi-antibiotic resistant *Staphylococcus* and *Streptococcus* species in a research centre farm at Naivasha. Kenya. *Bull. Anim. Health Pro. Afr.* 65, 291–303.
- Ndirangu, P.N., Mungube, E.O., Maichomo, M.W., Nyongesa, P.K., Wamae, D., On'gala, J.O., Gicheru, N.W., Keya, G., Wesonga, H.O., Siamba, D., 2019. A novel pH-based pen-side test for detection of sub-clinical mastitis: validation in cattle and camels. Kenya. *Livest. Res. Rural Dev.* 31, 7.
- Neelesh, S., Maiti, S.K., 2010. Prevalence and etiology of sub-clinical mastitis in cows. *Indian J. Vet. Res.* 19, 45–54.
- Nithya, P., Sivakumar, K., Aruljothi, A., 2017. A field study on prevalence of sub-clinical mastitis in dairy cows in Coimbatore district, Tamil Nadu. *Intl. J. Sci. Environ. Technol.* 6, 1453–1459.
- Norberg, E., Hogeweijn, H., Korsgaard, I.R., Friggens, N.C., Sloth, K.H.M.N., Løvendahl, P., 2004. Electrical conductivity of milk: ability to predict mastitis status. *J. Dairy Sci.* 87, 1099–1107.
- Olivares-Pérez, J., Khalif, A.E., Hernández, R.S., Elghandour, M.M.M.Y., Salem, A.Z.M., Bastida, A.Z., Velázquez-Reynoso, D., Cipriano-Salazar, M., Camacho-Díaz, L.M., Alonso-Fresán, M.U., DiLorenzo, N., 2015. Prevalence of bovine subclinical mastitis, its etiology and diagnosis of antibiotic resistance of dairy farms in four municipalities of a tropical region of Mexico. *Trop. Anim. Health Prod.* 47, 1497–1504.
- Oliver, S.P., Mitchell, B.A., 1984. Prevalence of mastitis pathogens in herds participating in a mastitis control program. *J. Dairy Sci.* 67, 2436–2440.
- Ondiek, J.O., Ogore, P.B., Shakala, E.K., Kaburu, G.M., 2013. Prevalence of bovine mastitis, its therapeutic and control in Tatton Agriculture Park, Egerton University. Njoro District of Kenya. *Basic Res. J. Agri. Sci.* Rev. 2, 15–20.
- Osman, K.M., Hassan, H.M., Ibrahim, I.M., Mikhail, M.M., 2010. The impact of *Staphylococcal* mastitis on the level of milk IL-6, lysozyme and nitric oxide. *Comp. Immunol. Microbiol. Infect. Dis.* 33, 85–93.
- Ostensson, K., Lam, V., Sjögren, N., Wredle, E., 2013. Prevalence of subclinical mastitis and isolated udder pathogens in dairy cows in Southern Vietnam. *Trop. Anim. Health Prod.* 45, 979–986.
- Orthan, N., Bahaman, A.R., 2005. Prevalence of subclinical mastitis and antibiotic resistant bacteria in three selected cattle farms in Serdang, Selangor and Kluang, Johor. *J. Vet. Malaysia* 17, 27–31.
- Ottalwar, T., Roy, M., Roy, S., Ottalwar, N., 2018. Prevalence of subclinical mastitis in buffaloes (*Bubalus bubalis*) in Chhattisgarh, India. *Intl. J. Agri. Sci. Res.* 8, 9–16.
- Özenç, E., Vural, M.R., Şeker, E., Uçar, M., 2008. An evaluation of subclinical mastitis during lactation in Anatolian buffaloes. *Turk. J. Vet. Anim. Sci.* 32, 359–368.
- Pachauri, S., Varshney, P., Dash, S.K., Gupta, M.K., 2013. Involvement of fungal species in bovine mastitis in and around Mathura, India. *Vet. World* 6, 393–395.
- Pal, M., Lemu, D., Bilato, T., 2017. Isolation, identification and antibiogram of bacterial pathogens from bovine subclinical mastitis in Asella, Ethiopia. *Intl. J. Livest. Res.* 7, 62–70.
- Panchal, I., Sawhney, I.K., Dang, A.K., 2016. Relation between electrical conductivity, dielectric constant, somatic cell count and some other milk quality parameters in diagnosis of subclinical mastitis in Murrah buffaloes. *Indian J. Dairy Sci.* 69, 267–271.
- Pankaj, A.S., Chhabra, R., Sindhu, N., 2012. Prevalence of sub clinical mastitis in cows: Its etiology and antibiogram. *Indian J. Anim. Res.* 46, 348–353.
- Pankaj, A.S., Chhabra, R., Sindhu, N., 2013. Sub-clinical mastitis in Murrah buffaloes with special reference to prevalence, etiology and antibiogram. *Buffalo Bull* 32, 107–115.
- Patel, Y.G., Trivedi, M.M., 2015. Quarter-wise prevalence of subclinical mastitis in crossbred cows. *Trends Biosci.* 8, 4727–4729.
- Patel, J.V., Bhingaradia, B.V., Patel, B.B., Patel, S.B., Patel, P.B., Vahora, S.P., 2012. Study on prevalence of mastitis and antibiotic sensitivity of bacterial isolates recovered from crossbred cows of Anand district of Gujarat. *Indian J. Dairy Sci.* 65, 467–471.
- Patel, K., Joshi, C., Kunjadiya, A., 2017. Isolation and molecular identification of *Staphylococcus* spp. from bovine mastitis milk samples. *Int. J. Adv. Res.* 5, 2320–2347.
- Perry, B.D., Carter, M.E., Hill, F.W.G., Milne, J.A.C., 1987. Mastitis and milk production in cattle in a communal land of Zimbabwe. *Br. Vet. J.* 143, 44–50.
- Petrovski, K.R., Heuer, C., Parkinson, T.J., Williamson, N.B., 2009. The incidence and aetiology of clinical mastitis on 14 farms in Northland, New Zealand. *New Zealand Vet. J.* 57, 109–115.
- Pinzón-Sánchez, C., Ruegg, P.L., 2010. Risk factors associated with short-term post-treatment outcomes of clinical mastitis. *J. Dairy Sci.* 94, 3397–3410.
- Plozza, K., Lievaart, J.J., Potts, G., Barkema, H.W., 2011. Subclinical mastitis and associated risk factors on dairy farms in New South Wales, Australia. *Aust. Vet. J.* 89, 41–46.
- Prabhakar, S.K., Singh, K.B., Joshi, D.V., 1995. Incidence and etiology of clinical mastitis in buffaloes. *Buffalo Bull.* 14, 63–67.
- Prabhu, K.N., Ruban, W.S., Kumar, G.S.N., Sharada, R., Padalkar, R.D., 2015. Sub-clinical mastitis in buffaloes: prevalence, isolation and antimicrobial resistance of *Staphylococcus aureus*. *Buffalo Bull.* 34, 215–222.
- Prakash, V., Bano, S., Yadav, M.P.S., Singh, R.B., Singh, S.P., 2013. Various factors responsible for sub clinical and clinical mastitis and pharmacotherapeutic role of trisodium citrate. *Res. J. Anim. Husbandry Dairy Sci.* 4, 51–53.
- Pranay, B., Champak, B., Japheth, K.P., Pooja, T., 2017. Interdependence and distribution of subclinical mastitis and intra-mammary infection among udder quarters in jersey crossbred cows. *Intl. J. Agri. Sci.* 9, 4235–4237.
- Preethiran, P.L., Isloor, S., Sundareshan, S., Nuthanalahkshmi, V., Deepthikiran, K., Sinha, A.Y., Rathnamma, D., Prabhu, K.N., Sharada, R., Mukkur, T.K., Hegde, N.R., 2015. Isolation, biochemical and molecular identification, and in-vitro antimicrobial resistance patterns of bacteria isolated from bubaline subclinical mastitis in South India. *PLoS One* 10, 1–15.
- Pyörälä, S., Hovinen, M., Simojoki, H., Fitzpatrick, J., Eckersall, P.D., Orro, T., 2011. Acute phase proteins in milk in naturally acquired bovine mastitis caused by different pathogens. *Vet. Rec. Rec.* 168, 535.
- Qadri, S.I.A., Shaheen, M., Baig, S.U., Malik, H., Bhat, Z.I., Bhat, I.A., Dar, P.A., Yousuf, R.W., Iqbal, R.M., Bhat, A.M., Haq, U.A., 2017. Aetio-prevalence study on bovine sub clinical mastitis in lactating Jersey crossbred cows. *Int. J. Curr. Microbiol. App. Sci.* 6, 3354–3357.
- Qayyum, A., Khan, J.A., Hussain, R., Avais, M., Ahmed, N., Khan, A., Khan, M.S., 2016. Prevalence and association of possible risk factors with sub-clinical mastitis in Cholistani cattle. *Pak. J. Zool.* 48, 519–525.
- Quaderi, M.A., Husain, M., Alam, M.G.S., Khatoon, M., Hossain, M.A., 2013. Prevalence of sub-clinical mastitis in dairy farms. *Bangladesh Vet.* 30, 70–77.
- Rabbani, A.F.M.G., Samad, M.A., 2010. Host determinants based comparative prevalence of subclinical mastitis in lactating Holstein-Friesian cross cows and Red Chittagong cows in Bangladesh. *Bangladesh J. Vet. Med.* 8, 17–21.
- Raiullah, R., Khan, M.A., Shafee, M., Akbar, A., Ali, A., Shoaib, M., Ashraf, F., Khan, N., 2017. Occurrence of mastitis and associated pathogens with antibiogram in animal population of Peshawar, Pakistan. *Thai. J. Vet. Med.* 47, 103–108.
- Rahman, M.A., Bhuiyan, M.M.U., Kamal, M.M., Shamsuddin, M., 2009. Prevalence and risk factors of mastitis in dairy cows. *Bangladesh Vet.* 26, 54–60.
- Rahman, M.M., Islam, M.R., Uddin, M.B., Aktaruzzaman, M., 2010. Prevalence of subclinical mastitis in dairy cows reared in Sylhet district of Bangladesh. *Intl. J. Bio. Res.* 1, 23–28.
- Rahman, M.M., Munshi, M.N., Ekram, M.F., Kabir, M.H., Rahman, M.T., Saha, S., 2014. Prevalence of subclinical mastitis in cows at Anwara, a coastal Upazila of Chittagong District in Bangladesh. *J. Vet. Adv.* 4, 594–598.
- Rall, V.L.M., Miranda, E.S., Castilho, I.G., Camargo, C.H., Langoni, H., Guimarães, F.F., Júnior, J.A., Júnior, A.F., 2014. Diversity of *Staphylococcus* species and prevalence of enterotoxin genes isolated from milk of healthy cows and cows with subclinical mastitis. *J. Dairy Sci.* 97, 829–837.
- Ramachandraiah, K., Kumar, K.S., Sreemannarayana, O., 1998. Subclinical mastitis in an organized buffalo farm. *Buffalo Bull.* 17, 85–87.
- Reddy, B.S., Shobhamani, B., Sreedevi, B., Kumari, K.N., Reddy, Y.R., 2015. Diagnosis of subclinical mastitis in cross breed cattle. *Res. Rev. J. Vet. Sci. Technol.* 4, 39–43.
- Regassa, A., Golicha, G., Tesfaye, D., Abunna, F., Megersa, B., 2013. Prevalence, risk factors, and major bacterial causes of camel mastitis in Borana zone, Oromia Regional State, Ethiopia. *Trop. Anim. Health Pro.* 45, 1589–1595.
- Reyes-Jara, A., Cordero, N., Aguirre, J., Troncoso, M., Figueroa, G., 2016. Antibacterial effect of copper on microorganisms isolated from bovine mastitis. *Front. Microbiol.* 7, 626.
- Reza, V.H., Mogaddam, F.M., Sadegh, M.M., Mirzaii, H., 2011. Bacterial pathogens of intra mammary infections in Azeri buffaloes of Iran and their antibiogram. *Afr. J. Agric. Res.* 6, 2516–2521.
- Romain, H.T., Adesiyun, A.A., Webb, L.A., Lauckner, F.B., 2000. Study on risk factors and their association with subclinical mastitis in lactating dairy cows in Trinidad. *J. Vet. Med. Series B* 47, 257–271.
- Rupp, R., Boichard, D., 2000. Relationship of early first lactation somatic cell count with risk of subsequent first clinical mastitis. *Livest. Prod. Sci.* 62, 169–180.

- Sadashiv, S.O., Kaliwal, B.B., 2013. Prevalence of bovine mastitis in north Karnataka, India. *Intl. J. Pharm. Health Care Res.* 1, 169177.
- Saidi, R., Khelef, D., Kaidi, R., 2013. Bovine mastitis: Prevalence of bacterial pathogens and evaluation of early screening test. *Afr. J. Microbiol. Res.* 7, 777–782.
- Sampimon, O.C., Barkema, H.W., Berends, I.M., Sol, J., Lam, T.J., 2009. Prevalence and herd-level risk factors for intra mammary infection with coagulase-negative *Staphylococci* in Dutch dairy herds. *Vet. Microbiol.* 134, 37–44.
- Sanjolly, G., Kotwal, S.K., Singh, S.G., Ahmed, T., Kour, A., Anand, A., 2017. Epidemiological study on mastitis in holstein friesian cattle on organized farm in Jammu, India. *Intl. J. Reprod. All Anim.* 7, 5–11.
- Sanotharan, N., Pagthiathan, M., Nafees, M.S.M., 2016. Prevalence of bovine subclinical mastitis and its association with bacteria and risk factors in milking cows of Batticaloa District in Sri Lanka. *Intl. J. Sci. Res. Innov. Technol.* 3, 2313–3759.
- Sarba, E.J., Tola, G.K., 2017. Cross-sectional study on bovine mastitis and its associated risk factors in Ambo district of West Shewa zone, Oromia, Ethiopia. *Vet. World* 10, 398–402.
- Sargeant, J.M., Scott, H.M., Leslie, K.E., Ireland, M.J., Bashiri, A., 1998. Clinical mastitis in dairy cattle in Ontario: frequency of occurrence and bacteriological isolates. *Can. Vet. J.* 39, 33–38.
- Sarker, S.C., Parvin, M.S., Rahman, A.A., Islam, M.T., 2013. Prevalence and risk factors of subclinical mastitis in lactating dairy cows in north and south regions of Bangladesh. *Trop. Anim. Health Prod.* 45, 1171–1176.
- Savita, A.C., Singh, A.P., Tak, L., Nayak, T.C., 2018. Study of prevalence, somatic cell counts and electrical conductivity in sub-clinical mastitic cattle. *Vet. Pract.* 19, 98–100.
- Sayed, H.R., Salama, S.S., Soliman, T.R., 2014. Bacteriological evaluation of present situation of mastitis in dairy cows. *Glob. Vet.* 13, 690–695.
- Schuberth, H.J., Krueger, C., Zerbe, H., Bleckmann, E., Leibold, W., 2001. Characterization of leukocytotoxic and super antigen-like factors produced by *Staphylococcus aureus* isolates from milk of cows with mastitis. *Vet. Microbiol.* 82, 187–199.
- Schwarzer, G., 2007. Meta: An R package for meta-analysis. *R News* 7, 40–45.
- Seid, U., Zenebe, T., Almaw, G., Edao, A., Disassa, H., Kabetta, T., Gerbi, F., Kebede, G., 2015. Prevalence, risk factors and major bacterial causes of bovine mastitis in west Arsi zone of Oromia region, Southern Ethiopia. *Nature Sci.* 13, 19–27.
- Selvaraju, G., Geetha, M., Saravanan, S., Manicavasaka, D.A., 2013. Evaluation of indirect tests for screening subclinical mastitis in dairy cows. *Indian J. Dairy Sci.* 66, 55–57.
- Shahid, M., Sabir, N., Ahmed, I., Khan, R.W., Irshad, M., Rizwan, M., Ahmed, S., 2011. Diagnosis of subclinical mastitis in bovine using conventional methods and electronic detector. *ARPN J. Agri. Biol. Sci.* 6, 18–22.
- Shaikh, S.R., Digraskar, S.U., Siddiqui, M.F.M.F., Borikar, S.T., Rajurkar, S.R., Suryawanshi, P.R., 2018. Epidemiological studies of mastitis in cows reared under different management system in and around Parbhani. *Pharm. Innov. J.* 8, 1–5.
- Sharif, A., Ahmad, T., 2007. Prevalence of severity of mastitis in buffaloes in district Faisalabad (Pakistan). *J. Agri. Soc. Sci.* 3, 34–36.
- Sharma, I., Brintly, A., 2014. Isolation and identification of *staphylococcus aureus* from bovine mastitis milk and their drug resistance patterns in Silchar town dairy farms, North East India. *Intl. Interdiscip. Res.* 4, 256–259.
- Sharma, A., Sindhu, N., 2007. Occurrence of clinical and subclinical mastitis in buffaloes in the state of Haryana (India). *Ital. J. Anim. Sci.* 6, 965–967.
- Sharma, A., Dhingra, P., Pander, B.L., Kumar, R., 2006. Bovine sub clinical mastitis: prevalence and treatment with homeopathic medicine. *Intl. J. Cow Sci.* 2, 40–44.
- Sharma, N., Maiti, S.K., Sharma, K.K., 2007. Prevalence, etiology of microorganisms associated with sub-clinical mastitis in buffaloes in Durg, Chhattisgarh State (India). *Int. J. Dairy Sci.* 2, 145–151.
- Sharma, D.K., Jallewar, P.K., Sharma, K.K., 2010a. Antibiogram of bacteria isolated from bovine subclinical mastitis. *Indian Vet. J.* 87, 407.
- Sharma, N., Pandey, V., Soodan, J.S., 2010b. Prevalence of mastitis in lactating dairy cows. *Indian J. Vet. Med.* 30, 102–104.
- Sharma, A., Chhabra, R., Sindhu, N., 2012. Prevalence of sub clinical mastitis in cows: Its etiology and antibiogram. *Indian J. Anim. Res.* 46, 348–353.
- Sharma, N., Kang, T.Y., Lee, S.J., Kim, J.N., Hur, C.H., Ha, J.C., Vohra, V., Jeong, D.K., 2013. Status of bovine mastitis and associated risk factors in subtropical Jeju Island, South Korea. *Trop. Anim. Health Pro.* 45, 1829–1832.
- Sharma, L., Verma, A.K., Kumar, A., Rahat, A., Neha Nigam, R., 2015. Incidence and pattern of antibiotic resistance of *Staphylococcus aureus* isolated from clinical and subclinical mastitis in cattle and buffaloes. *Asian Australas. J. Anim. Sci.* 9, 100–109.
- Sharma, A., Chhabra, R., Singh, M., Charaya, G., 2018a. Prevalence, etiology and antibiogram of bacterial isolates recovered from mastitis of buffaloes. *Buffalo Bull.* 37, 313–320.
- Sharma, R., Ashutosh, M., Pandita, S., Yadav, P.S., Parkunan, T., 2018b. Quarter-wise prevalence of subclinical mastitis in crossbred cows. *Indian J. Anim. Res.* 52, 116–120.
- Sharma, N., Singh, S.G., Sharma, S., Gupta, S.K., Hussain, K., 2018c. Mastitis occurrence pattern in dairy cows and importance of related risk factors in the occurrence of mastitis. *J. Anim. Res.* 8, 315–326.
- Shem, M.N., Malole, J.M.L., Machangu, R., Kurwijila, L.R., Fujihara, T., 2001. Incidence and causes of sub-clinical mastitis in dairy cows on smallholder and large scale farms in tropical areas of Tanzania. *Asian Aust. J. Anim. Sci.* 14, 372–377.
- Shiferaw, J., Telila, I., 2016. Prevalence of bovine mastitis and assessment of risk factors in and around Wolayta Sodo, Ethiopia. *Intl. J. Homeopathy Natural Med.* 2, 1–7.
- Shitandi, A., Kihumbu, G., 2004. Assessment of the California mastitis test usage in smallholder dairy herds and risk of violative antimicrobial residues. *J. Vet. Sci.* 5, 5–10.
- Shittu, A., Abdullahi, J., Jibril, A., Mohammed, A.A., Fasina, F.O., 2012. Sub-clinical mastitis and associated risk factors on lactating cows in the Savannah Region of Nigeria. *BMC Vet. Res.* 8, 134.
- Shrestha, S., Bindari, Y.R., 2012. Prevalence of sub-clinical mastitis among dairy cattle in Bhaktapur District, Nepal. *Intl. J. Agri. Biosci.* 1, 16–19.
- Shrivastava, N., Sharma, V., Nayak, A., Shrivastava, A.B., Sarkhel, B.C., Shukla, P.C., Shrivastava, A., 2017. Prevalence and characterization of methicillin-resistant *staphylococcus aureus* (MRSA) mastitis in dairy cattle in Jabalpur, Madhya Pradesh. *J. Anim. Res.* 7, 77–84.
- Siddique, M.Z.F., Islam, S., Islam, S.S., Islam, S., Islam, S., Das, B.C., 2015. Haematobiochemical changes in subclinical mastitis affected high yielding dairy cows in Chittagong district. *Intl. J. Natural Soc. Sci.* 2, 30–34.
- Siddiquee, N.U., Tripura, T.K., Islam, M.T., Bhuiyan, S.A., Rahman, A.K.M.A., Bhuiyan, A.K.F.H., 2013. Prevalence of sub-clinical mastitis in high yielding cross bred cows using Draminski mastitis detector. *Bangladesh J. Vet. Med.* 11, 37–41.
- Silva, N.C.C., Guimaraes, F.F., Manzi, M.P., Budri, P.E., Gómez-Sanz, E., Benito, D., Langoni, H., Rall, V.L.M., Torres, C., 2013. Molecular characterization and clonal diversity of methicillin-susceptible *Staphylococcus aureus* in milk of cows with mastitis in Brazil. *J. Dairy Sci.* 96, 6856–6862.
- Sindhu, N., Sharma, A., Nehra, V., Jain, V.K., 2009. Occurrence of subclinical mastitis in cows and buffaloes at an organized farm. *Haryana Vet.* 48, 85–87.
- Sindhu, N., Sharma, A., Chhabra, R., 2012. Prevalence of subclinical mastitis in cows: Its etiology and antibiogram. *Indian J. Anim. Res.* 46, 348–353.
- Singh, P.K., Diwakar, R.P., Johri, A., 2018. Study the prevalence of *S. aureus* in subclinical mastitis and their antibiogram in buffaloes. *J. Pharmacogn. Phytochem.* 4, 440–442.
- Sori, H., Zerihun, A., Abdicho, S., 2005. Dairy cattle mastitis in and around Sebeta, Ethiopia. *J. Appl. Res. Vet. Med.* 3, 332.
- Srinivasan, P., Jagadeswaran, D., Manoharan, R., Giri, T., Balasubramaniam, G.A., Balachandran, P., 2013. Prevalence and etiology of subclinical mastitis among buffaloes (*Bubalus bubalis*) in Namakkal, India. *Pak. J. Biol. Sci.* 16, 1776–1780.
- Ssajjakambwe, P., Bahizi, G., Setumba, C., Kisaka, S., Vudriko, P., Atuhire, C., Kabasa, J.D., Kaneene, J.B., 2017. Milk hygiene in rural southwestern Uganda: prevalence of mastitis and antimicrobial resistance profiles of bacterial contaminants of milk and milk products. *Vet. Med. Int.* 2017, 8710758.
- Stevenson, M.A., 2000. Disease incidence in dairy herds in the southern highlands district of New South Wales, Australia. *Prev. Vet. Med.* 43, 1–11.
- Subramanian, T., Senthilkumar, T.M.A., Vijayarani, K., Meenakshi Sundaram, S., Venkataramanan, R., 2019. Incidence and factors affecting subclinical and clinical mastitis in selected organised dairy farms located in Tamil Nadu. *J. Anim. Res.* 9, 303–309.
- Sudhan, N.A., Singh, R., Singh, M., Soodan, J.S., 2005. Studies on prevalence, etiology and diagnosis of subclinical mastitis among crossbred cows. *Indian J. Anim. Res.* 39, 127–130.
- Sukumar, K., James, P.C., 2012. Incidence of fungal mastitis in cattle. *Tamilnadu J. Vet. Anim. Sci.* 8, 356–359.
- Suleiman, A.B., Kwaga, J.K.P., Umoh, V.J., Okoloch, E.C., Muhammed, M., Lammier, C., Shaibu, S.J., Akineden, O., Weiss, R., 2012. Macro-restriction analysis of *Staphylococcus aureus* isolated from subclinical bovine mastitis in Nigeria. *Afr. J. Microbiol. Res.* 6, 6270–6274.
- Suleiman, T.S., Karimuribo, E.D., Mdegela, R.H., 2013. Prevalence of mastitis in smallholder dairy cattle in Pemba Island, Tanzania. *Tanzania Vet. J.* 28, 70–81.
- Suleiman, T.S., Karimuribo, E.D., Mdegela, R.H., 2017. Prevalence of bovine subclinical mastitis and antibiotic susceptibility patterns of major mastitis pathogens isolated in Unguja island of Zanzibar, Tanzania. *Trop. Anim. Health Pro.* 50, 259–266.
- Sumon, S.M.M.R., Ehsan, M.A., Islam, M.T., 2017. Subclinical mastitis in dairy cows: Somatic cell counts and associated bacteria in Mymensingh, Bangladesh. *J. Bangladesh Agric. Univ.* 15, 266–271.
- Sundaresan, S., Isloor, S., Babu, Y.H., Awati, B., Hegde, R., Sunagar, R., Waryah, C.B., Gogoi-Tiwari, J., Mukkur, T.K., Hegde, N.R., 2014. A comparative evaluation of four detection tests and the isolation of coagulase-negative *Staphylococci* from subclinical mastitis cases in south Indian cattle. *Indian J. Comp. Microbiol. Immunol. Infect. Dis.* 35, 73–78.
- Sunder, J., De, A.K., Jeyakumar, S., Kundu, A., 2013. Effect of feeding of *Morinda citrifolia* fruit juice on the biophysical parameters of healthy as well as mastitis-affected cow milk. *J. Appl. Anim. Res.* 41, 29–33.
- Suresh, M., Safullah, A.H.M., Kathiravan, G., Narmatha, N., 2017. Incidence of clinical mastitis among small holder dairy farms in India. *Atatürk Univ. J. Vet. Sci.* 12, 1–13.
- Suriyathaporn, W., 2011. Epidemiology of subclinical mastitis and their antibacterial susceptibility in smallholder dairy farms, Chiang Mai province, Thailand. *J. Anim. Vet. Adv.* 10, 316–321.
- Swami, S.V., Patil, R.A., Gadekar, S.D., 2017. Studies on prevalence of subclinical mastitis in dairy animals. *J. Entomol. Zool. Stud.* 5, 1297–1300.
- Swamy, M.C.M., Krishnamurthy, G.V., 1998. Prevalence of *Staphylococcus* species in California mastitis test positive cows. *Indian Vet. J.* 75, 101–103.
- Sylejmani, D., Ramadani, N., Robaj, A., Hamidi, A., 2016. Prevalence and antimicrobial susceptibility of bacterial isolates recovered from subclinical mastitis in dairy farms in Kosovo. *Bulg. J. Vet. Med.* 19, 299–307.
- Sztachańska, M., Barański, W., Zduńczyk, S., Janowski, T., Pogorzelska, J., Zdunczyk, S., 2016. Prevalence and etiological agents of subclinical mastitis at the end of lactation in nine dairy herds in North-East Poland. *Pol. J. Vet. Sci.* 19, 119–124.
- Tadesse, A., Chanie, M., 2012. Study on the occurrence of bovine mastitis in Addis Ababa dairy farms and associated risk factors. *Adv. Biol. Res.* 6, 151–158.
- Tafa, F., Terefe, Y., Tamerat, N., Zewdu, E., 2015. Isolation, identifications and antimicrobial susceptibility pattern of coagulase positive *Staphylococcus* from

- subclinical mastitic dairy cattle in and around Haramaya University. *Ethiop. Vet. J.* 19, 41–53.
- Taponen, S., Simojoki, H., Haveri, M., Larsen, H.D., Pyörälä, S., 2006. Clinical characteristics and persistence of bovine mastitis caused by different species of coagulase-negative *Staphylococci* identified with API or AFLP. *Vet. Microbiol.* 115, 199–207.
- Tekle, Y., Berihe, T., 2015. Bovine mastitis: prevalence, risk factors and major pathogens in Sidama Zone SNNPRS. *Ethiopia. Int'l. J. Innov. Sci. Maths* 3, 230–238.
- Tesfaye, B., 2017. Bovine mastitis: Prevalence, risk factors, major pathogens and antimicrobial susceptibility test on the isolates around addis Ababa. *Central Ethiopia. Open Access J. Vet. Sci.* Res. 2, 1–6.
- Tesfaye, B., Abera, A., 2018. Prevalence of mastitis and associated risk factors in Jimma town dairy farms. *Western Ethiopia. J. Vet. Sci. Anim. Husbandry* 6, 1–8.
- Tesfaye, A., Mekonnen, H., Rahwa, S., Dendin, S., 2012. The effect of management practices on prevalence of mastitis in small scale dairy farms in Nazareth. *Ethiopia. World J. Agri. Sci.* 8, 218–222.
- Thakor, D., Patel, D., 2013. Prevalence and antibiogram pattern of subclinical mastitis in crossbred cows, Natural Remedies Private Limited. <https://en.engormix.com/dairy-cattle/articles/prevalence-antibiogram-pattern-sub-t35804.htm>. Accessed 10 June 2020.
- Thakur, S., Singh, M., Aseri, G.K., Verma, A., Khan, S.S., 2018. Isolation and characterization of mastitis pathogens and milk composition changes in Murrah buffaloes (*Bubalus bubalis*) during winter season. *Indian J. Anim. Res.* 52, 276–280.
- Thanh, N.V., Hai, N.T., Son, N.N., Dung, B.V., Atsushi, M., 2015. A study about mastitis infection characteristics in dairy cow of Bavi, Hanoi. *Vietnam. Asian J. Pharma. Clin. Res.* 8, 165–168.
- Thompson, W.H., Houston, B.A., 1967. A survey on the incidence of bovine mastitis in dairy herds in two districts in Victoria. *Aust. Vet. J.* 43, 558–563.
- Tilahun, A., Aylate, A., 2015. Prevalence of bovine mastitis in lactating cows and its public health implications in selected commercial dairy farms of Addis Ababa. *Glob. J. Med. Res. (G)* 15, 16–24.
- Tolosa, T., Gebretedik, Z., Regassa, F., 2009. Bovine mastitis and its associated risk factors in lactating cows in Wolaita Soddo. *Southern Ethiopia. Bull. Anim. Health Pro. Afr.* 57, 123–129.
- Tolosa, T., Verbeke, J., Piepers, S., Supré, K., De Vliegher, S., 2013. Risk factors associated with subclinical mastitis as detected by California mastitis test in smallholder dairy farms in Jimma, Ethiopia using multilevel modelling. *Prev. Vet. Med.* 112, 68–75.
- Tripathi, S., Arora, N., Shekhar, S., Rajora, V.S., 2018. Etio-prevalence of sub clinical mastitis in crossbred cattle. *J. Entomol. Zool. Stud.* 6, 778–780.
- Triputra, T.K., Sarker, S.C., Roy, S.K., Parvin, M.S., Sarker, R.R., Rahman, A.K.M.A., Islam, M.T., 2014. Prevalence of subclinical mastitis in lactating cows and efficacy of intra mammary infusion therapy. *Bangladesh J. Vet. Med.* 12, 55–61.
- Tufani, N.A., Makhdoomi, D.M., Hafiz, A., 2012. Epidemiology and therapeutic management of bovine mastitis. *Indian J. Anim. Res.* 46, 148–151.
- Tuke, M., Kassaye, D., Muktar, Y., Negeye, T., Nigusu, K., 2017. Bovine mastitis: prevalence and associated risk factors in Alage ATVET college dairy farm, Southern Ethiopia. *J. Vet. Sci. Technol.* 8, 1–5.
- Türkylimaz, S., Yıldız, Ö., Oryaşın, E., 2010. Molecular identification of bacteria isolated from dairy herds with mastitis. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi* 16, 1025–1032.
- Tyagi, S.P., Joshi, R.K., Joshi, N., 2013. Characterization and antimicrobial sensitivity of *Staphylococcus aureus* isolates from subclinical bovine mastitis. *J. Anim. Health Pro.* 1, 20–23.
- Umar, S., Sarwar, F., Usman, M., Shah, M.A.A., Ghafar, A., Ali, A., Asif, S., 2013. In vitro antimicrobial sensitivity pattern of mastitis causing bacterial pathogens isolated from cattle in arid zones of Punjab, Pakistan. *Sci. Lett.* 1, 17–20.
- Umaru, G.A., Kwaga, J.K.P., Bello, M., Raji, M.A., Maitala, Y.S., 2017. Occurrence of bovine mastitis and isolation of *Staphylococcus* species from fresh cow milk in settled Fulani herds in Kaduna State. Nigeria. *Bayero J. Pure Appl. Sci.* 10, 259–263.
- Vázquez, H.C., Jäger, S., Wolter, W., Zschöck, M., Vazquez, C., El-Sayed, A., 2013. Isolation and identification of main mastitis pathogens in Mexico. *Arquivo Brasileiro de Medicina Veterinaria e Zootecnia* 65, 377–382.
- Verma, H., Rawat, S., Sharma, N., Jaiswal, V., Singh, R., 2018. Prevalence, bacterial etiology and antibiotic susceptibility pattern of bovine mastitis in Meerut. *J. Entomol. Zool. Stud.* 6, 706–709.
- Vieira-da-Motta, O., Folly, M.M., Sakyama, C.C.H., 2001. Detection of different *Staphylococcus aureus* strains in bovine milk from subclinical mastitis using PCR and routine techniques. *Braz. J. Microbiol.* 32, 27–31.
- Viguier, C., Arora, S., Gilmartin, N., Welbeck, N., O'Kennedy, R., 2009. Mastitis detection: current trends and future perspectives. *Trends Biotechnol.* 27, 486–493.
- Vishwakarma, P., Sushovan, R., Manju, R., Shakya, S., 2010. Occurrence of sub clinical mastitis in buffaloes of Chhattisgarh. *Indian J. Vet. Med.* 30, 94–96.
- Visvanath, K., Shoba, K., Dorairajan, N., Srithar, A., Chandran, N.D.J., 2012. Clinical and sub-clinical bovine mastitis in Chelliyaeplayam, Namakkal district. In: Indian Association of Veterinary Microbiologists, Immunologists and Specialists in Infectious Diseases conference 6–8th September, 2012. India, Bangalore.
- Walker, E., Hernandez, A.V., Kattan, M.W., 2008. Meta-analysis: its strengths and limitations. *Cleve. Clin. J. Med.* 75, 431–439.
- Wang, L., Yang, F., Wei, X.J., Luo, Y.J., Guo, W.Z., Zhou, X.Z., Guo, Z.T., 2019. Prevalence and risk factors of subclinical mastitis in lactating cows in Northwest China. *Isr. J. Vet. Med.* 74, 17–22.
- Watson, D.L., McColl, M.L., Davies, H.I., 1996. Field trial of a staphylococcal mastitis vaccine in dairy herds: clinical, subclinical and microbiological assessments. *Aust. Vet. J.* 74, 447–450.
- White, M.E., Glickman, L.T., Barnes-Pallese, F.D., Stem, E.S., Dinsmore, P., Powers, M. S., Powers, P., Smith, M.C., Jasko, D., 1986. Accuracy of clinicians in predicting the bacterial cause of clinical bovine mastitis. *Can. Vet. J.* 27, 218–220.
- Wilson, D.J., Gonzalez, R.N., Das, H.H., 1997. Bovine mastitis pathogens in New York and Pennsylvania: prevalence and effects on somatic cell count and milk production. *J. Dairy Sci.* 80, 2592–2598.
- Wilson, D.J., Grohn, Y.T., Bennett, G.J., González, R.N., Schukken, Y.H., Spatz, J., 2007. Comparison of J5 vaccines and controls for incidence, etiologic agent, clinical severity, and survival in the herd following naturally occurring cases of clinical mastitis. *J. Dairy Sci.* 90, 4282–4288.
- Workneh, S., Bayleyegn, M., Mekonnen, H., Potgieter, L.N.D., 2002. Prevalence and aetiology of mastitis in cows from two major Ethiopian dairies. *Trop. Anim. Health Prod.* 34, 19–25.
- Wubishet, Z., Ararsa, D., Alemayehu, L., 2013. Bovine mastitis in selected districts of Borena zone. *Southern Ethiopia. Bull. Anim. Health Pro. Afr.* 61, 173–179.
- Xu, J., Tan, X., Zhang, X., Xia, X., Sun, H., 2015. The diversities of *Staphylococcal* species, virulence and antibiotic resistance genes in the subclinical mastitis milk from a single Chinese cow herd. *Microb. Pathog.* 88, 29–38.
- Yang, F.L., Shen, C., He, B.X., Yang, Y.Y., Li, X.S., 2014. The prevalence of heifer mastitis and its associated risk factors in Huanggang, Central China. *Trop. Anim. Health Prod.* 47, 87–92.
- Yohannes, K., Alemu, B., 2018. Prevalence of Bovine Mastitis in lactating Cows and Associated risk factors in and around Wolaita Soddo, Southern Ethiopia. *Int'l. J. Adv. Res. Biol. Sci.* 5, 60–69.
- Yohannis, M., Molla, W., 2013. Prevalence, risk factors and major bacterial causes of bovine mastitis in and around Wolaita Sodo, Southern Ethiopia. *Afr. J. Microbiol. Res.* 7, 5400–5405.
- Zaki, M.S., El-Batrawy, N., Mostafa, S.O., 2010. Some biochemical Studies on Friesian suffering from subclinical mastitis. *Nat. Sci.* 8, 143–146.
- Zenebe, N., Habtam, T., Endale, B., 2014. Study on bovine mastitis and associated risk factors in Adigrat, Northern Ethiopia. *Afr. J. Microbiol. Res.* 8, 327–331.
- Zeng, R., Bequette, B.J., Vinyard, B.T., Bannerman, D.D., 2009. Determination of milk and blood concentrations of lipopolysaccharide-binding protein in cows with naturally acquired subclinical and clinical mastitis. *J. Dairy Sci.* 92, 980–989.
- Zeryehun, T., Abera, G., 2017. Prevalence and bacterial isolates of mastitis in dairy farms in selected districts of eastern Harrarghe Zone, Eastern Ethiopia. *J. Vet. Med.* 2017, 1–7.
- Zeryehun, T., Aya, T., Bayeche, R., 2013. Study on prevalence, bacterial pathogens and associated risk factors of bovine mastitis in small holder dairy farms in and around Addis Ababa. Ethiopia. *J. Anim. Plant Sci.* 23, 50–55.
- Zutic, M., Cirkovic, I., Pavlovic, L., Zutic, J., Asanin, J., Radanovic, O., Pavlovic, N., 2012. Occurrence of methicillin-resistant *Staphylococcus aureus* in milk samples from Serbian cows with subclinical mastitis. *Afr. J. Microbiol. Res.* 6, 5887–5889.