

<b>STANDARD OPERATING PROCEDURE</b>			
<i>Title:</i>	<b>DETECTION OF PORCINE CIRCO VIRUS (PCV) BY PCR</b>		
<i>Reference:</i>	<b>G Saikumar, IVRI</b>	<i>Author:</i>	<b>Dr N N Barman, Dr(Ms) Arpita Bharali</b>

## 1. INTRODUCTION

### Purpose/Scope of this SOP

- 1.1. This SOP outlines the method for the detection of PCV DNA from tissue samples. DNA is isolated from samples using the DNASure Tissue Mini Kit.

**All PCR reagents are stored at -20°C. Primers purified either by desalting or HPLC purification are acceptable for this protocol.**

## 2. MATERIALS

2.1	<b>Consumables</b>	<b>Supplier</b>
	0.2ml PCR tubes	GENAXY
2.2	<b>Equipment</b>	<b>Supplier, Model</b>
	Class 2 safety microbiological safety cabinet	LABCHEM & LABORTENIK INSTRUMENTS
	Micro-centrifuge	TARSONS
	VERITI 96-well Thermalcycler	APPLIED BIOSYSTEMS
	Micropipette (0.5-10 µl, 10-100 µl, 20-200 µl, 100-1000 µl)	GENAXY (Nichipette)
	Filter Tips	GENAXY

### 2.3 TABLE 1: Primer sets used:

<b>Primer designation</b>	<b>Primer sequence</b>
PCV2F	CGGATATTGTAGTCCTGGTCG
PCV2R	ACTGTCAAGGCTACCACAGTCA

**F=** Forward Primer; **R=** Reverse Primer

**3. PROCEDURE/METHOD**

**3.1. Isolation of DNA from samples using kit**

- 3.1.1. Isolate DNA from samples using DNASure Tissue Mini Kit.
- 3.1.2. Once the DNA has been eluted, store at -18°C or below or keep on ice and proceed to detection of PCV by PCR.

**4. Preparation of PCR master mix**

- 4.1 Preparation of PCR master mix must be carried out in the PCR clean room.
- 4.2. Prepare a master mix containing the following reagents and preferably add the reagents in the order given.

<b>Reagent</b>	<b>Volume per reaction</b>
PCR MasterMix (Fermentas)	12.5 µl
Forward primer (10uM)	2 µl
Reverse primer (10uM)	2 µl
Nuclease free water	6.5 µl

- 4.3. Thoroughly mix the master mix and aliquot 22µl into the appropriate number of Optical Flat Cap 8/Strip or plates.

**4.4. Addition of templates**

- 4.4.1. Add 2µl of the DNA samples to the 22µl master mix.
- 4.4.2. Once DNA is added, fit caps to all wells. It is important that the caps are fitted firmly and correctly onto the wells before being used on the PCR machine.

## 5. PCR THERMAL CYCLING CONDITION

- 5.5.1. Place the reaction tubes in PCR system (Applied Biosystems).
- 5.5.2. Incubate the reactions with the following thermo cycling profile:

PCR step:

1. 94<sup>0</sup>C for 5 min
2. 40 cycles
  - 94<sup>0</sup>C for 30 sec
  - 56<sup>0</sup>C for 45 sec
  - 72<sup>0</sup>C for 30 sec
3. 72<sup>0</sup>C for 5 min and hold at 4<sup>0</sup>C

## 6. RESULTS

### 6.1. Confirmation of PCR amplicons : Gel Electrophoresis

The confirmation of PCR amplicons was carried out by their sizes in agarose gel. The PCR products were electrophoresed in 1.7% agarose gel containing ethidium bromide in 0.5X tris borate EDTA and visualized on a UV transilluminator as per standard procedures. For size comparison, a 100bp DNA ladder marker was run parallel to the PCR amplicons.

**The PCR amplicon PCV shows a product size of 481 bp.**

**Interpretation:** The amplified product visualized as a single compact fluorescent band of 481bp under UV light and documented by gel documentation system is considered positive for BVDV.